

Clinical Research Protocol
Clearance of 25-hydroxyvitamin D in chronic kidney disease

Protocol Number:	CLEAR-01
Version:	2.1
Investigational Product:	Deuterated 25-dihydroxyvitamin D ₃
IND Number:	115016
Development Phase:	Phase 1
Sponsor:	Ian H. de Boer, MD, MS
Funding Organization:	National Institute of Diabetes and Digestive and Kidney Diseases (R01DK099199)
Principal Investigator:	Name: Ian H. de Boer, MD, MS Telephone: (206) 616-5403 Fax: (206) 685-9399 E-mail: deboer@u.washington.edu

TABLE OF CONTENTS

1	BACKGROUND	9
1.1	Overview of Clinical Studies.....	9
2	STUDY RATIONALE	9
2.1	Risk / Benefit Assessment	11
3	STUDY OBJECTIVES	11
3.1	Primary Objective.....	11
3.2	Secondary Objectives	11
4	STUDY DESIGN	11
5	CRITERIA FOR EVALUATION	12
5.1	Primary Efficacy Endpoint	12
5.2	Secondary Efficacy Endpoints.....	12
5.3	Safety Evaluations	12
6	SUBJECT SELECTION	13
6.1	Study Population.....	13
6.2	Inclusion Criteria	14
6.3	Exclusion Criteria.....	14
7	CONCURRENT MEDICATIONS	15
8	STUDY TREATMENTS	15
8.1	Method of Assigning Subjects to Treatment Groups.....	15
8.2	Blinding	15
8.3	Formulation of Test and Control Products	16
8.4	Supply of Study Drug at the Site	16
8.5	Supply of Study Drug at the Site	17
8.6	Study Drug Accountability	17
8.7	Measures of Treatment Compliance.....	17
9	STUDY PROCEDURES AND GUIDELINES	17
9.1	Clinical Assessments	18
9.2	Clinical Laboratory Measurements.....	18
9.3	Pharmacokinetic Measurements	19
9.4	Research Laboratory Measurements.....	19
10	EVALUATIONS BY VISIT	19
10.1	Screening Visit (Study Visit 1).....	19
10.2	Baseline Visit (Study Visit 2, Day 0)	20
10.3	Study Visit 3 (Day 1).....	20
10.4	Study Visit 4 (Day 4).....	20
10.5	Study Visit 5 (Day 7).....	20
10.6	Study Visits 6-10 (Day 14, 21, 28, 42, and 56)	21
11	ADVERSE EXPERIENCE REPORTING AND DOCUMENTATION	21
11.1	Adverse Events	21

11.2 Serious Adverse Experiences (SAE) 22

12 DISCONTINUATION AND REPLACEMENT OF SUBJECTS 24

12.1 Early Discontinuation of Study Drug 24

12.3 Withdrawal of Subjects from the Study..... 24

12.4 Replacement of Subjects 25

13 PROTOCOL VIOLATIONS 25

14 STATISTICAL METHODS AND CONSIDERATIONS 25

14.1 Data Sets Analyzed..... 25

14.2 Demographic and Baseline Characteristics 25

14.3 Analysis of Primary Endpoint 25

14.4 Analysis of Secondary Endpoints 26

14.5 Interim Analysis 26

14.6 Sample Size and Randomization 26

15 DATA COLLECTION, RETENTION AND MONITORING 26

15.1 Data Collection Instruments 26

15.2 Data Management Procedures 26

15.3 Data Quality Control and Reporting..... 26

15.4 Archival of Data 27

15.5 Availability and Retention of Investigational Records..... 27

15.6 Monitoring 27

15.7 Subject Confidentiality 27

16 ADMINISTRATIVE, ETHICAL, REGULATORY CONSIDERATIONS 27

16.1 Protocol Amendments 28

16.2 Institutional Review Boards and Independent Ethics Committees..... 28

16.3 Informed Consent Form..... 28

16.4 Publications 29

16.5 References 30

LIST OF ABBREVIATIONS

AE	adverse event
ALT	alanine aminotransferase
AST	aspartate aminotransferase
BUN	blood urea nitrogen
CFR	Code of Federal Regulations
CRF	case report form
FDA	Food and Drug Administration
GCP	Good Clinical Practice
HIPAA	Health Insurance Portability and Accountability Act of 1996
ICF	informed consent form
ICH	International Conference on Harmonisation
IEC	Independent Ethics Committee
IRB	Institutional Review Board
IV	intravenous
PI	Principal Investigator
PK	pharmacokinetic
SAE	serious adverse experience
SGOT	serum glutamic oxaloacetic transaminase
SGPT	serum glutamate pyruvate transaminase

PROTOCOL SYNOPSIS

TITLE	Clearance of 25-hydroxyvitamin D ₃ in chronic kidney disease
SPONSOR	Ian H. de Boer, MD, MS
FUNDING ORGANIZATION	National Institute of Diabetes and Digestive and Kidney Diseases (R01DK099199)
NUMBER OF SITES	Single-site study
RATIONALE	<p>Patients with chronic kidney disease (CKD) are at high risk of adverse health outcomes, including progression to end stage renal disease, cardiovascular disease events, and death. Impaired vitamin D metabolism is commonly observed in CKD and may contribute to the morbidity of this disease. In healthy individuals, the kidney produces the majority of circulating 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃, or calcitriol). Calcitriol binds the vitamin D receptors in diverse target tissues and is known to modulate inflammation and fibrosis, suppress the renin-angiotensin system, and regulate proliferation and differentiation. In CKD, renal production of 1,25(OH)₂D₃ is reduced, resulting in secondary hyperparathyroidism and dysregulation of other organs and tissues. Our preliminary data suggest that clearance of vitamin D metabolites is also reduced in CKD.</p> <p>The goal of this study is to better understand vitamin D catabolism and how it is affected by CKD and race. Specifically, we will evaluate the metabolic clearance of D6-25-hydroxyvitamin D₃ in individuals with varying degrees of CKD and among participants who self-report race as Caucasian, African American or African. The long-term goal of this work is to enhance the clinical evaluation and treatment of impaired vitamin D metabolism.</p>
STUDY DESIGN	<p>This is a single-dose, open-label pharmacokinetic study that uses intravenous administration of a stable deuterium-labeled 25(OH)D₃ to evaluate the metabolic clearance of 25(OH)D₃.</p> <p>Allocation: Non-Randomized Intervention Model: Single Group Assignment Masking: Open Label</p>
PRIMARY OBJECTIVE	<p>The goal of this study is to better understand vitamin D catabolism and how it is affected by CKD and race. Specifically, we will evaluate the metabolic clearance of 25(OH)D₃ in individuals with varying degrees of CKD and among participants who self-report their race as Caucasian, African American, or African.</p>

SECONDARY OBJECTIVES	<ul style="list-style-type: none"> To explore the biochemical pathways through which 1,25(OH)D₃ is metabolized, and the effects of CKD and race on these pathways, by evaluating the appearance and disappearance of labeled metabolites of 25(OH)D₃. To determine clinical and biochemical factors that are correlated with the metabolic clearance of 25(OH)D₃. To evaluate biomarkers of 25(OH)D₃ clearance.
NUMBER OF SUBJECTS	Up to 120
SUBJECT SELECTION CRITERIA	<p><u>Inclusion Criteria:</u></p> <ul style="list-style-type: none"> Age ≥ 18 years Self-reported race Caucasian, African American, or African Serum total 25(OH)D 10-50 ng/mL Chronic kidney disease status: Estimated GFR ≥60 mL/min/1.73m² (N=40) Estimated GFR <60 mL/min/1.73m², not treated with dialysis (N=40) End stage renal disease treated with hemodialysis (N=40) <p><u>Exclusion Criteria:</u></p> <ul style="list-style-type: none"> Primary hyperparathyroidism Gastric bypass Tuberculosis or sarcoidosis Current pregnancy Child-Pugh Class B or C cirrhosis (i.e. cirrhosis with ascites, hepatic encephalopathy, bilirubin ≥2 mg/dL, serum albumin ≤3.5 g/dL, or PT ≥ 4 seconds) History of kidney transplantation (unless failed transplant now treated with hemodialysis) Use of vitamin D₃, or vitamin D₂ supplements exceeding a mean daily dose of 400 IU, within 3 months (wash-out allowed) Use of 1,25(OH)₂D₃ or an analogue, calcimimetics, or medications known to induce CYP24A1 within 4 weeks (wash-out allowed) Serum calcium > 10.1 mg/dL Hemoglobin < 9 g/dL
TEST PRODUCT, DOSE, AND ROUTE OF ADMINISTRATION	<p>We will administer D₆-25-hydroxyvitamin D₃ Solution for Injection, 20 µg/mL intravenously. The deuterated 25(OH)D₃ is formulated using cosolvents to enhance solubility. Each mL contains 20 µg D₆-25(OH) D₃, 60% propylene glycol, 10% ethanol (ETOH) and water (30%) for injection, to volume. To ensure reliable detection of circulating deuterated 25(OH)D₃, without administering a dose that might alter the underlying</p>

	<p>vitamin D metabolism, we aim to administer a dose that results in a peak deuterated 25(OH)D₃ concentration of approximately 5 ng/mL. The administered dose will be calculated as the targeted peak serum deuterated 25(OH)D₃ (5 ng/mL) multiplied by blood volume.</p>
CONTROL PRODUCT, DOSE AND ROUTE OF ADMINISTRATION	There is no control product.
DURATION OF SUBJECT PARTICIPATION AND DURATION OF STUDY	<p>Subjects will be on study for approximately 12 weeks</p> <p>Screening: approximately 4 weeks</p> <p>Treatment: 1 day (subjects to the University of Washington Clinical Research Center)</p> <p>Follow-up: 8 weeks</p> <p>The total duration of the study is expected to be up to 3 years for subject recruitment and an additional 12 weeks for final subject follow-up.</p>
CONCOMMITANT MEDICATIONS	<p>Allowed:</p> <ul style="list-style-type: none"> Vitamin D supplements (cholecalciferol, ergocalciferol) not to exceed a mean daily dose of 400 IU, within 3 months (wash-out allowed) <p>Prohibited (within last 4 weeks, wash-out allowed):</p> <ul style="list-style-type: none"> 1,25(OH)₂D₃ or an analogue (e.g. paricalcitol, hectorol) Calcimimetics (e.g. cinaclacet) Medications known to potentially induce or inhibit CYP24A1 or CYP3A4
EFFICACY EVALUATIONS	None
PRIMARY ENDPOINT	<ul style="list-style-type: none"> Metabolic clearance of labeled 25(OH)D₃ (administered dose/AUC)
SECONDARY ENDPOINTS	<ul style="list-style-type: none"> AUC of labeled 25(OH)D₃ Terminal half-life of labeled 25(OH)D₃ Volume of distribution in the central compartment of labeled 25(OH)D₃
OTHER EVALUATIONS	Metabolic formation clearance (metabolite/parent AUC ratio) for metabolites of labeled 25(OH)D ₃
SAFETY EVALUATIONS	<p>Change in the serum concentrations of calcium, creatinine, AST, and ALT and from baseline to 7 days after 25(OH)D₃ administration</p> <p>Incidence of adverse events</p>

PLANNED INTERIM ANALYSES	This is not a clinical trial with an efficacy outcome, and it is neither randomized nor blinded. Formal interim analyses are not planned.
STATISTICS Primary Analysis Plan	We will examine the distribution of D ₆ -25(OH)D ₃ clearance by CKD status (3 categories) and race (2 categories) using box plots and summary statistics. We will test associations of CKD status and race with D ₆ -25(OH)D ₃ clearance (continuous outcome variable) using linear regression. Models will be adjusted for age, gender, diabetes status, body size, baseline 25(OH)D concentration, and other covariates strongly related to 25(OH)D ₃ clearance. We will test whether associations of CKD status with clearance are modified by race by including a CKD x race interaction term.
Rationale for Number of Subjects	Our power calculations conservatively assume an interaction between CKD status and race, for which each cell in the 3x2 recruitment grid would require individual comparison. With this approach, we estimate that 17 people in each of two groups will provide 80% power to detect a difference in D ₆ -25(OH)D ₃ half-life of 2 days, assuming a standard deviation of 2 days. We will recruit 20 people per cell to allow for multivariable adjustment.

1 BACKGROUND

Among people with chronic kidney disease (CKD), impaired vitamin D metabolism may contribute to cardiovascular disease, progression to end stage renal disease, and premature death.¹ In healthy individuals, the kidney produces the majority of circulating 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃, calcitriol), which binds to the vitamin D receptors in diverse target tissues. Activation of Vitamin D₃ signaling pathways is known to modulate inflammation and fibrosis, suppress the renin-angiotensin system, and attenuate secondary hyperparathyroidism. The evaluation and treatment of impaired vitamin D metabolism is a major focus of current clinical care for CKD patients however, clinical decision making is limited by the lack of an effective measure of functional 1,25(OH)₂D₃ deficiency. 25-hydroxyvitamin D₃ (25(OH)D₃) is a relatively inactive substrate form of vitamin D; circulating 1,25(OH)₂D₃ concentration is tightly regulated and poorly reflects tissue levels; and circulating parathyroid hormone reflects functional 1,25(OH)₂D₃ deficiency at only one of many relevant biological sites.

1.1 Overview of Clinical Studies

Deuterated and tritiated 25(OH)D₃ have been used previously to study metabolism of vitamin D₃ in humans.²⁻⁶ In addition deuterium is has been used extensively to study metabolism and is considered safe.⁷

2 STUDY RATIONALE

Patients with chronic kidney disease (CKD) are at high risk of adverse health outcomes, including progression to end stage renal disease, cardiovascular disease events, and death.⁸ Impaired vitamin D metabolism is commonly observed in CKD and may contribute to the morbidity of this disease.¹ In healthy individuals, the kidney produces the majority of circulating 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃, or calcitriol). Calcitriol binds the vitamin D receptors in diverse target tissues, and is known to modulate inflammation and fibrosis, suppress the renin-angiotensin system, and regulate proliferation and differentiation. In CKD, renal production of 1,25(OH)₂D₃ is reduced, resulting in secondary hyperparathyroidism and dysregulation of other organs and tissues. Our preliminary data suggest that clearance of vitamin D metabolites is also reduced in CKD.

Concentrations of vitamin D metabolites in blood and tissues must necessarily represent a balance of production and catabolism. While basic mechanisms of vitamin D catabolism have been defined,⁹ the clinical evaluation of vitamin D catabolism has received remarkably little attention. In health, CYP24A1 is the major enzyme responsible for catabolism of both 25(OH)D₃ and 1,25(OH)₂D₃.^{1,9-15} CYP24A1 is found in most tissues in the body and is rapidly induced by 1,25(OH)₂D₃ to prevent 1,25(OH)₂D₃ intoxication.¹⁶⁻¹⁸ As a result, CYP24A1 expression is commonly used as a marker of tissue 1,25(OH)₂D₃ activity in basic science.¹⁹⁻³⁵ However, the ability of vitamin D catabolism to reflect 1,25(OH)₂D₃ function has not been similarly harnessed for clinical use. Evaluation of circulating 24,25(OH)₂D₃ concentration may offer this opportunity. 24,25(OH)₂D₃ is the most abundant product of 25(OH)D₃ catabolism by CYP24A1.⁹ When viewed in the context of prevailing 25(OH)D₃, 24,25(OH)₂D₃ reflects CYP24A1 activity. 24,25(OH)₂D₃ circulates at high concentrations (generally 1-10 ng/mL)^{36,37} and has a circulating half-life of approximately 7 days.³⁸ Circulating 24,25(OH)₂D₃ appears to

come from both renal and non-renal sources.^{15,22,39-41} In the kidney proximal tubule, CYP24A1 transcription is induced by FGF-23 and suppressed by PTH.^{17,42}

To study vitamin D catabolism in humans, we developed a novel high-throughput mass spectrometry assay for 24,25(OH)₂D₃, measured concurrently with 25(OH)D₃, 25(OH)D₂, 1,25(OH)₂D₃, and 1,25(OH)₂D₂.⁴³ We applied this assay to the serum of 278 participants in the Seattle Kidney Study (SKS), a single-center cohort study of pre-dialysis CKD.³⁶ Serum 25(OH)D₃ did not vary by eGFR, but mean serum concentrations of 24,25(OH)₂D₃ were lower with lower eGFR: 3.6, 3.2, 2.6, 2.6, and 1.7 ng/mL for eGFR ≥60, 45-59, 30-44, 15-29, and <15 mL/min/1.73m², respectively (p<0.001). These data are consistent with smaller CKD studies that observed low circulating concentrations of 24,25(OH)₂D₃ measured by immunoassay^{6,44-48} and with one study suggesting substantially reduced metabolic clearance of 25(OH)D in ESRD.⁴⁹ In SKS, serum 24,25(OH)₂D₃ correlated moderately with 25(OH)D (r²=0.56) but not with 1,25(OH)₂D (r²<0.01).

In addition to 24-hydroxylation, 25(OH)D₃ also undergoes hydroxylation at the 4-position and sulfation at the 3-O-position. The resulting metabolites, 4β,25(OH)₂D₃ and 25(OH)D₃-3-sulfate, respectively, are generated in the liver by CYP3A4⁵⁰ and SULT2A1 (unpublished results). As with CYP24A1, CYP3A4 expression is induced by 1,25(OH)₂D₃.⁵¹ In a preliminary analysis of plasma from 20 prevalent hemodialysis patients and 14 normal controls, we found that the relationships of plasma 4β,25(OH)₂D₃ and 25(OH)D₃-3-sulfate with plasma 25(OH)D₃ were distinctly different for the two populations. Specifically, slopes were less steep in dialysis patients, with lower metabolite concentrations at any given concentration of 25(OH)D₃. This parallels the relationship we previously observed for 24,25(OH)₂D₃ and suggests that hepatic 25(OH)D₃ metabolism is also reduced in CKD. In addition, plasma 25(OH)D₃-3-sulfate concentrations were comparable in magnitude to plasma 25(OH)D₃ concentrations, suggesting that sulfation may be a quantitatively important unrecognized route of vitamin D catabolism. Because 25(OH)D₃-3-sulfate is probably eliminated by the kidney, the degree to which 25(OH)D₃ sulfation is reduced in CKD may be underestimated by steady state plasma 25(OH)D₃-3-sulfate concentrations.

Vitamin D metabolism also differs substantially by race.⁵² The circulating concentration of 25(OH)D is substantially lower in people who define their race as African American or African compared to Caucasian populations, presumably due to differences in skin pigmentation and cutaneous cholecalciferol synthesis. Interestingly, the circulating concentrations of 1,25(OH)₂D₃ do not vary by race, although there are noted differences in bone density between the two populations. Data from our cohort suggest associations between low 25(OH)D levels and coronary heart disease among Caucasians, but not in African American or African populations. This data suggests that previously unrecognized differences in vitamin D catabolism may contribute to racial differences in disease susceptibility.

The goal of this study is to better understand vitamin D catabolism and how it is affected by CKD and race. Specifically, we will evaluate the metabolic clearance of 25-hydroxyvitamin D₃ in individuals with varying degrees of CKD and among participants who self-report their race as Caucasian, African American, or African. The long-term goal of this work is to enhance the clinical evaluation and treatment of impaired vitamin D metabolism.

2.1 Risk / Benefit Assessment

This study will not provide any direct benefit to the study subjects. Individual subjects may have a modestly reduced chance of adverse health events because their care will be monitored more closely than it might otherwise. Future patients may benefit from these studies by virtue of knowledge gained about the pathophysiology and ascertainment of impaired vitamin D metabolism.

The risks of administering deuterated 25(OH)D₃ in tracer quantities are low. 25(OH)D₃ is a naturally occurring substance and generally circulates in concentrations of 10-50 ng/mL. The deuterated 25(OH)D₃ we propose to administer differs from the naturally occurring form only by the substitution of 6 hydrogen atoms with deuterium. This isotope is stable and has metabolic and biologic characteristics identical to the naturally occurring form. We will administer quantities intended to make small changes in total circulating 25(OH)D₃ concentration, i.e., an increase of 5 ng/mL.

There is some risk of hypercalcemia with the administration 25(OH)D₃, but this risk is minimized by excluding participants with baseline 25(OH)D > 50 ng/mL and by administering quantities that raise 25(OH)D by only approximately 5 ng/mL. We will monitor for hypercalcemia during our research. If instances of hypercalcemia are observed, we will change our study protocol accordingly to prevent hypercalcemia in future participants. This study also includes placement of peripheral intravenous catheters and blood draws. Risks include discomfort and minor bleeding associated with catheter placements and anemia due to blood sampling.

3 STUDY OBJECTIVES

3.1 Primary Objective

The goal of this study is to better understand vitamin D catabolism and how it is affected by CKD and race. Specifically, we will evaluate the metabolic clearance of 25(OH)D₃ across stages of CKD and among participants who self-report race as Caucasian, African American or African. The long-term goal of this work is to enhance the clinical evaluation and treatment of impaired vitamin D metabolism.

3.2 Secondary Objectives

- To explore the biochemical pathways through which 25(OH)D₃ is metabolized, and the effects of CKD and race on these pathways, by evaluating the appearance and disappearance of labeled metabolites of 25(OH)D₃.
- To determine clinical and biochemical factors that are correlated with the metabolic clearance of 25(OH)D₃.
- To evaluate potential biomarkers of 25(OH)D₃

4 STUDY DESIGN

This is a cross-sectional, observational study that uses the intravenous administration of stable isotope-labeled D₆-25(OH)D₃ to evaluate the metabolic clearance of 25(OH)D₃. It is a single-dose, open-label PK study of 25(OH)D₃.

We will recruit a total of up to 120 study subjects, with recruitment stratified by CKD status and race. Subjects who provide written informed consent and qualify at screening will receive a single dose of intravenous labeled 25(OH)D₃. Subsequent blood draws and urine collections will be used to determine the metabolic clearance of 25(OH)D₃ and related parameters.

5 CRITERIA FOR EVALUATION

5.1 Primary Efficacy Endpoint

The primary outcome of interest is the metabolic clearance of labeled 25(OH)D₃.

We will measure serum concentrations of 24,25(OH)₂D₃, 25(OH)D₃, 25(OH)D₂, 1,25(OH)₂D₃ and 1,25(OH)₂D₂ at the University of Washington by mass spectrometry. For each participant and time point, a single serum aliquot is used to measure this panel of vitamin D metabolites using immunoaffinity extraction and HPLC-mass spectrometry (Xevo TQ, Waters Corp., Milford, MA) with deuterated internal standards.⁵³⁻⁵⁶

For each subject, non-compartmental analysis of plasma concentration versus time data will be performed using Phoenix software (Pharsight, Cary, NC). Clearance will be calculated as administered 25(OH)D₃ dose divided by 25(OH)D₃ AUC. We focus on clearance as our primary outcome because it reflects the metabolism of 25(OH)D₃ accounting for circulating 25(OH)D₃ concentration (units of volume/time, akin to creatinine clearance). Clearance is independent of volume of distribution, in contrast to $t_{1/2}$, which will be evaluated as a secondary outcome. Clearance will be evaluated with and without adjustment for body size.

5.2 Secondary Efficacy Endpoints

- AUC of labeled D₆-25(OH)D₃
- Terminal half-life of labeled D₆-25(OH)D₃
- Volume of distribution in the central compartment of labeled D₆-25(OH)D₃
- Metabolic formation clearance (metabolite/parent AUC ratio) for metabolites of labeled D₆-25(OH)D₃

5.3 Safety Evaluations

An abnormal clinical laboratory value will be documented as an adverse event if one of the following applies:

- The abnormality is not contradicted by a repeat test to confirm the abnormality.
- The abnormality suggests a disease and/or organ toxicity.
- The abnormality is of a degree that requires active management (e.g., requires a medication change, more frequent follow-up, or further diagnostic evaluation).

Change in clinical laboratory findings (if there are specific labs, then why they are appropriate to measure, e.g., BUN or Creatinine for an aminoglycoside)

An adverse event (AE) is any symptom, sign, illness or experience that develops or worsens in severity during the course of the study. Intercurrent illnesses or injuries should be regarded as adverse events. Abnormal results of diagnostic procedures are considered to be adverse events if the abnormality:

- results in study withdrawal
- is associated with a serious adverse event
- is associated with clinical signs or symptoms
- leads to additional treatment or to further diagnostic tests
- is considered by the investigator to be of clinical significance

A serious adverse event is any AE that is:

- fatal
- life-threatening
- requires or prolongs hospital stay
- results in persistent or significant disability or incapacity
- a congenital anomaly or birth defect
- an important medical event

Important medical events are those that do not fit the other definitions of serious adverse events, but the event may jeopardize the patient and may require treatment to prevent one of the listed serious adverse events.

Adverse events will be reported if they occur between the time of informed consent and 30 days after the last study visit. All unresolved adverse events will be followed by the PI until resolution, the adverse event is otherwise explained, or the participant is lost to follow-up. At the last study visit, the investigator will instruct each participant to report any subsequent event that the participant, or participant’s personal physician, reasonably believes may be related to the study. The investigator will notify the study sponsor of any death or adverse event occurring after the participant has discontinued participation if the event can be reasonably related to the study.

6 SUBJECT SELECTION

6.1 Study Population

Recruitment for this study will be stratified by CKD status and race (Table 6.1). Study participants will be recruited from the Seattle Kidney Study (an established, active cohort study of people with CKD) and other Kidney Research Institute cohorts and trial, Nephrology clinics that refer to the Kidney Research Institute, the Healthy Kidney Study (a cohort study of people without known kidney disease, conducted in parallel to the Seattle Kidney Study), primary care clinics within the University of Washington system, dialysis units within the Northwest Kidney Centers and Puget Sound Kidney Centers, and the University of Washington Kidney Research Institute participant registry.

Table 6.1 Target recruitment by chronic kidney disease status.		
	Self-reported race	
	Caucasian	African American or African
End state renal disease treated with hemodialysis	20	20

Estimated GFR <60 ml/min/1.73m ² , not treated with dialysis	20	20
Estimated GFR ≥60 ml/min/1.73m ²	20	20

6.2 Inclusion Criteria

We will study only adults (Table 6.2). In addition to eligibility criteria stratified by CKD status and race, we will limit to a range of total serum 25(OH)D.

Table 6.2. Participant Eligibility – Inclusion criteria
Target chronic kidney disease status as described in table 6.1
Race as described in table 6.1
Age ≥ 18 years
Serum 25(OH)D concentration 10 – 50ng/ml

6.3 Exclusion Criteria

Exclusion criteria will be

- inability to give informed consent
- Primary hyperparathyroidism
- Gastric bypass
- Tuberculosis or sarcoidosis
- pregnancy
- Child-Pugh Class B or C cirrhosis (i.e. cirrhosis with ascites, hepatic encephalopathy, bilirubin ≥2 mg/dL, serum albumin ≤3.5 g/dL, or PT ≥ 4 seconds)
- History of kidney transplantation (unless failed transplant now treated with hemodialysis)
- hemoglobin <9 mg/dL
- serum calcium >10.1 mg/dL
- use of vitamin D₃ or vitamin D₂ supplements (e.g. ergocalciferol or cholecalciferol > 400 IU/day) within 3 months
- use of active vitamin D receptor agonists (e.g. 1,25(OH)₂D₃ or an analogue, calcitriol) or cinacalcet within 4 weeks
- use of a cytochrome P-450 (CYP) inhibitor or inducer within 4 weeks (table 6.4)

If people do not meet eligibility criteria based on medication use, they will be allowed to participate in the study after an appropriate washout period if their primary physician agrees. For ergocalciferol and cholecalciferol, the washout period will be 3 months. For active vitamin D compounds, cinacalcet, and CYP inhibitors and inducers the washout period will be 4 weeks. If excluding for ergocalciferol or cholecalciferol supplementation >400 IU/day proves to be a major impediment to successful recruiting, we will consider liberalizing this exclusion criterion to >800 IU/day or >1000 IU/day.

Table 6.3. Patient Eligibility – Exclusion criteria
--

Ergocalciferol in last 3 months, cholecalciferol >400IU/day in last 3 months
Active vitamin D receptor agonist (e.g. calcitriol) or cinacalcet in last 4 weeks
Child-Pugh Class B or C cirrhosis
Hemoglobin < 9 mg/dl
Serum calcium concentration > 10.1 mg/dl
Pregnancy
Inability to give informed consent
Medications known to strongly induce or suppress the CYP enzymes which metabolize vitamin D (see example list below)

Table 6.4. CYP inhibitors and inducers

CYP Inhibitors		CYP Inducers
Amprenavir	Imatinib	Avasimibe
Aprepitant	Indinavir	Bosentan
Atazanavir	Itraconazole	Carbamazepine
Casopitant	Ketoconazole	Efavirenz
Cimetidine	Lopinavir	Etravirine
Ciprofloxacin	Nefazodone	Modafinil
Clarithromycin	Nelfinavir	Nafcillin
Conivaptan	Posaconazole	Phenobarbital
Darunavir	Ritonavir	Phenytoin
Diltiazem	Saquinavir	Rifabutin
Dronedarone	Shisandra	Rifampin
Elvitegravir	Telithromycin	St. John’s Wort
Erythromycin	Tipranavir	
Fluconazole	Verapamil	
Grapefruit Juice*	Voriconazole	

* Greater than 8 ounces per day

7 CONCURRENT MEDICATIONS

All subjects should be maintained on the same medications throughout the entire study period, as medically feasible, with no introduction of new chronic therapies. Specific allowed and disallowed medications are detailed in Section 6 (Eligibility Criteria).

8 STUDY TREATMENTS

8.1 Method of Assigning Subjects to Treatment Groups

All consenting and qualified participants will be assigned to the single treatment arm.

8.2 Blinding

Neither participants nor study investigators will be blinded to the study intervention.

8.3 Formulation of Test and Control Products

We will administer labeled D₆-25(OH)D₃ Solution for Injection, 20 µg/mL intravenously. D₆-25(OH) D₃ Solution for Injection, 20 µg/mL has been manufactured according to current Good Manufacturing Practices by SRI International (Palo Alto, CA).

8.3.1 Formulation of Test Product

The labeled D₆-25(OH)D₃ Solution for Injection, 20 µg/mL prepared by SRI International is formulated in cosolvents to enhance solubility. Each mL contains 20 µg D₆-25(OH) D₃, 60% propylene glycol (60%), 10% ethanol (ETOH) and water (30%) for injection, to volume.

8.3.2 Packaging and Labeling

The formulated drug product has been aliquoted into single-use vials by SRI International and frozen at <60°C for storage. Vial labels include the drug product name, concentration, batch number, manufacture date, and storage conditions, similar to the draft label shown below.

<p>D₆-25-Hydroxyvitamin D₃ Solution for Injection, 20 µg/mL, 5 mL single use vial</p> <p>Subject ID: _____</p> <p>Batch No.: 16504-08 Manuf. Date: Sept 2016</p> <p>Store at ≤ -60°C (- 76°F)</p> <p>See protocol for preparation for administration instructions</p> <p><i>Caution: New Drug -- Limited by Federal (US) law to investigational use. Keep out of the reach of children.</i></p> <p>Manufactured by SRI International, Menlo Park, CA 94025</p>
--

8.4 Supply of Study Drug at the Site

Single-use vials will be stored at <60°C at the University of Washington Investigational Drug Services or Kidney Research Institute. Individual doses of D₆-25(OH) D₃ Solution for Injection will be prepared for intravenous administration by the University of Washington Investigational Drug Services by thawing and drawing the appropriate volume into a syringe. The prepared dose of will be calculated according to estimated blood volume based on body weight. The prepared dosing syringe will be labeled with the participant's full name, date of birth, and medical record number along with the drug contents. Once thawed, the drug product will be infused within eight hours of preparation.

8.4.1 Dosage/Dosage Regimen

Each participant will receive a single intravenous dose of D₆-25(OH) D₃ Solution for Injection, 20 µg/mL. We will use estimated blood volume based on body weight to achieve a target concentration of 5 ng/mL D₆-25(OH) D₃. The target of 5 ng/mL was selected because it will allow precise tracking of circulating deuterated D₆-25(OH) D₃ concentration throughout follow-up (based on known limits of detection for our assay) without substantially perturbing underlying vitamin D status. Theoretically, the dose required to achieve a 5 ng/mL increment in deuterated 25(OH)D₃ concentration can be calculated as the targeted peak concentration multiplied by the volume of distribution, which for 25(OH)D₃ is expected to be equal to blood volume. We will use the formula of Nadler et al (1962) to estimate blood volume for each participant. We will begin the study by administering to each participant a dose of 5 ng/mL x estimated blood

volume. A typical dose would be 32 µg (1.6 mL) for an 85 kg individual. We will monitor achieved D₆-25(OH) D₃ concentrations throughout the course of the study. If our initial approach fails to achieve peak D₆-25(OH) D₃ concentrations near 5 ng/mL, we will revise our approach for future participants, as indicated. For example, if mean peak achieved 25(OH)D₃ concentrations are near 2.5 ng/mL with our initial approach, we will increase our dose by a factor of two. We will not exceed a dose of 100 µg (5 mL) D₆-25(OH) D₃ for any participant.

8.4.2 Dispensing

All study drug will be dispensed by the University of Washington Investigational Drug Services.

8.4.3 Administration Instructions

The study drug will be administered intravenously on a single occasion over a period of 5 minutes. Subsequent to thawing, the drug product vial may be stored refrigerated (2-8 °C) for up to 24 hours prior to preparation for administration in a syringe. Once prepared for administration and held at room temperature, the solution should be administered within 8 hours.

8.5 Supply of Study Drug at the Site

All single-use vials of formulated D₆-25(OH)D₃ will be shipped overnight on dry ice to the University of Washington Investigational Drug Services, with temperature monitoring. The University of Washington is the sole clinical site for this study.

8.5.1 Storage

Study drug will be securely stored at a temperature <-60°C at the University of Washington Investigational Drug Services or Kidney Research Institute.

8.6 Study Drug Accountability

An accurate and current accounting of the dispensing and return of study drug for each subject will be maintained on an ongoing basis by the University of Washington Investigational Drug Services. The number of study drug dispensed and returned by the subject will be recorded on the Investigational Drug Accountability Record.

8.7 Measures of Treatment Compliance

Each study drug administration will be directly observed by study staff.

9 STUDY PROCEDURES AND GUIDELINES

A Schedule of Events representing the required testing procedures to be performed for the duration of the study is provided in Section 10 (Evaluations by Visit) below and diagrammed in Appendix 1.

Prior to conducting any study-related activities, written informed consent and the Health Insurance Portability and Accountability Act (HIPAA) authorization must be signed and dated by the subject or subject's legal representative. If appropriate, assent must also be obtained prior to conducting any study-related activities.

9.1 Clinical Assessments

9.1.1 Concomitant Medications

All concomitant medication and concurrent therapies will be documented at Screening, Baseline (Study Day 0), and the final Study Visit (Study Visit 10). Dose, route, unit frequency of administration, and indication for administration and dates of medication will be captured.

9.1.2 Demographics

Demographic information (date of birth, gender, race) will be recorded at Screening.

9.1.3 Medical History

Relevant medical history, including history of current disease, other pertinent history, and information regarding underlying diseases will be recorded at Baseline (Study Day 0).

9.1.4 Physical Examination

Height and weight will be measured at the Screening visit and at Baseline (Study Day 0). When prompted by reports or suspicion of potential adverse effects during follow-up, physical exam findings will be documented by qualified staff (MD, NP, RN, or PA) and will be followed by a physician or other qualified staff immediately or at the next scheduled visit, as indicated.

9.1.5 Vital Signs

Body temperature, blood pressure, pulse and respirations will be measured after resting for 5 minutes at Baseline (Study Day 0), prior to administration of study drug.

9.1.6 Other Clinical Procedures

Physical activity will be ascertained by questionnaire and accelerometry between Study days 1 and 7. Vitamin D intake will be estimated by food frequency questionnaire at Baseline. Serum and plasma will be collected at Baseline to measure calcium and phosphorus, intact parathyroid hormone, fibroblast growth factor-23, and vitamin D binding protein. Two 24-hour urine collections will be obtained, immediately preceding and following administration of D₆-25(OH) D₃, to quantify urinary excretion of albumin and the metabolites of deuterated D₆-25(OH) D₃, respectively. Dialysis patients will be exempt from all urine collections. Adiposity will be measured by Dual energy X-ray absorptiometry at the baseline visit.

9.1.7 Adverse Events

Information regarding occurrence of adverse events will be captured throughout the study. Duration (start and stop dates and times), severity/grade, outcome, treatment and relation to study drug will be recorded on the case report form (CRF).

9.2 Clinical Laboratory Measurements

9.2.1 Hematology

A complete blood count will be obtained at the Screening Visit.

9.2.2 Blood Chemistry Profile

Serum sodium, potassium, chloride, bicarbonate, random glucose, BUN, creatinine, aspartate aminotransferase (AST/SGOT), alanine aminotransferase (ALT/SGPT), albumin, calcium, and phosphorous will be obtained at the Screening Visit.

Repeat serum sodium, potassium, chloride, bicarbonate, random glucose, BUN, creatinine, aspartate aminotransferase (AST/SGOT), alanine aminotransferase (ALT/SGPT), albumin, calcium, and phosphorous will be obtained at Baseline and Study Day 7.

Intact parathyroid hormone (PTH) will be measured at Visits 2 and 5 (Day 0 and 7 gauged from administration of deuterated 25(OH)D₃).

9.2.3 Pregnancy Test

A urine pregnancy test will be obtained from female subjects who are of childbearing age at the Baseline Visit prior to administration of the study drug. For anuric (dialysis) participants, blood pregnancy tests will be obtained to rule out pregnancy during the week preceding the administration of the study drug.

9.3 Pharmacokinetic Measurements

Blood for determination of serum concentrations of deuterated 25(OH)D₃ and its metabolites will be collected at Study Visits 2-10.

At Baseline, blood will be drawn prior to administration of study drug as well as 5 minutes and 4 hours after administration.

9.4 Research Laboratory Measurements

Novel products of 25(OH)D₃ catabolism, including 4β,25(OH)₂D₃ and 25(OH)D₃-3-sulfate will be measured using LC-MS/MS, as previously described.⁵⁷ 25(OH)D₃-3-sulfate will be extracted from plasma.

10 EVALUATIONS BY VISIT

10.1 Screening Visit (Study Visit 1)

The following activities will occur during the screening visit:

- Collect signed consent form
- Perform focused medical history restricted to study eligibility criteria
- Complete medication inventory through EMR and patient report.
- Physical Exam (height and weight), and Vital Signs (blood pressure, heart rate)
- Collect non-fasting blood sample for measurement of complete blood count and total 25(OH)D.
- Collect non-fasting blood sample for serum sodium, potassium, chloride, bicarbonate, random glucose, BUN, creatinine, AST/SGOT, ALT/SGPT, albumin, calcium, and phosphorous.
- Collect urine for measurement of albumin and creatinine (except for hemodialysis patients).
- Participants who meet eligibility criteria at the screening visit will be invited to return for the remaining study visits.
- Provide selected subjects with supplies required for first 24-hour urine collection

10.2 Baseline Visit (Study Visit 2, Day 0)

Visit 2 will take place at the University of Washington Clinical Research Center (CRC) or a suitable alternate facility and will last approximately 5 hours. Visit 2 will take place no more than 90 days after Visit 1. If more than 90 days elapse between Visits 1 and 2, screening procedures may be repeated to verify continued eligibility. The following activities will occur during the baseline visit:

- Perform a complete medical history
- Review current medications.
- Physical Exam (height and weight), and Vital Signs (blood pressure, heart rate)
- Complete physical activity and food frequency questionnaires.
- Pregnancy test for women of childbearing potential
- Collect blood **prior to administration of study drug** for measurement of basic chemistries, vitamin D metabolites, parathyroid hormone (PTH), and other measurements related to kidney disease and vitamin D metabolism.
- Collect urine sample from first 24-hour collection period. Sample will be used for measurement of albumin, creatinine, vitamin D binding protein, and vitamin D metabolites:
- Perform dual energy x-ray absorptiometry (DEXA) to measure adiposity.
- Administer D₆₋₂₅(OH) D₃ intravenously.
- Collect blood 5 minutes and 4 hours post-infusion for measurement of vitamin D metabolites.
- Distribute supplies for second 24-hour urine collection.

10.3 Study Visit 3 (Day 1)

This visit is expected to last less than one hour. The following activities will occur during Visit 3:

- Collect urine sample from second 24-hour collection period.
- Collect blood for measurement of serum vitamin D metabolite concentrations.
- Distribute Accelerometer for measurement of physical activity

10.4 Study Visit 4 (Day 4)

This visit is expected to last less than one hour. A window of one day before or after the scheduled date will be allowed when needed. Blood will be collected for measurement of serum vitamin D metabolite concentrations.

10.5 Study Visit 5 (Day 7)

This visit is expected to last less than one hour. A window of one day before or after the scheduled date will be allowed when needed. The following activities will occur during Visit 7:

- Collect blood for measurement of serum vitamin D metabolite concentrations.
- Collect blood for serum sodium, potassium, chloride, bicarbonate, random glucose, BUN, creatinine, aspartate aminotransferase (AST/SGOT), alanine aminotransferase (ALT/SGPT), albumin, calcium, and phosphorous at Study Visit 5 (Day 7) to monitor study safety.
- Collect Accelerometer

10.6 Study Visits 6-10 (Day 14, 21, 28, 42, and 56)

Each of these visits is expected to last less than one hour. A window of up to 3 days before or after the scheduled date will be allowed when needed. Blood will be collected at each visit for measurement of serum vitamin D metabolite concentrations. Medication inventory will be repeated at Visit 10 (Day 56).

11 ADVERSE EXPERIENCE REPORTING AND DOCUMENTATION

11.1 Adverse Events

An adverse event (AE) is any symptom, sign, illness or experience that develops or worsens in severity during the course of the study. Intercurrent illnesses or injuries should be regarded as adverse events. Abnormal results of diagnostic procedures are considered to be adverse events if the abnormality:

- results in study withdrawal
- is associated with a serious adverse event
- is associated with clinical signs or symptoms
- leads to additional treatment or to further diagnostic tests
- is considered by the investigator to be of clinical significance

11.1.1 AE Severity

The National Cancer Institute's Common Terminology Criteria for Adverse Events (CTCAE) Version 3.0 should be used to assess and grade AE severity, including laboratory abnormalities judged to be clinically significant. The modified criteria can be found in the study manual. If the experience is not covered in the modified criteria, the guidelines shown in Table 11.1 below should be used to grade severity. It should be pointed out that the term "severe" is a measure of intensity and that a severe AE is not necessarily serious.

Table 11.1 AE Severity Grading

Severity (Toxicity Grade)	Description
Mild (1)	Transient or mild discomfort; no limitation in activity; no medical intervention or therapy required. The subject may be aware of the sign or symptom but tolerates it reasonably well.
Moderate (2)	Mild to moderate limitation in activity, no or minimal medical intervention/therapy required.
Severe (3)	Marked limitation in activity, medical intervention/therapy required, hospitalizations possible.
Life-threatening (4)	The subject is at risk of death due to the adverse experience as it occurred. This does not refer to an experience that hypothetically might have caused death if it were more severe.

11.1.2 AE Relationship to Study Drug

The relationship of an AE to the study drug should be assessed using the following the guidelines in Table 11.2.

Table 11.2 AE Relationship to Study Drug

Relationship to Drug	Comment
Definitely	Previously known toxicity of agent; or an event that follows a reasonable temporal sequence from administration of the drug; that follows a known or expected response pattern to the suspected drug; that is confirmed by stopping or reducing the dosage of the drug; and that is not explained by any other reasonable hypothesis.
Probably	An event that follows a reasonable temporal sequence from administration of the drug; that follows a known or expected response pattern to the suspected drug; that is confirmed by stopping or reducing the dosage of the drug; and that is unlikely to be explained by the known characteristics of the subject's clinical state or by other interventions.
Possibly	An event that follows a reasonable temporal sequence from administration of the drug; that follows a known or expected response pattern to that suspected drug; but that could readily have been produced by a number of other factors.
Unrelated	An event that can be determined with certainty to have no relationship to the study drug.

11.2 Serious Adverse Experiences (SAE)

An SAE is defined as any AE occurring at any dose that results in any of the following outcomes:

- death
- a life-threatening adverse experience
- inpatient hospitalization or prolongation of existing hospitalization
- a persistent or significant disability/incapacity
- a congenital anomaly/birth defect

Other important medical events may also be considered an SAE when, based on appropriate medical judgment, they jeopardize the subject or require intervention to prevent one of the outcomes listed.

A serious adverse event is any AE that is:

- fatal
- life-threatening
- requires or prolongs hospital stay
- results in persistent or significant disability or incapacity
- a congenital anomaly or birth defect
- an important medical event

Important medical events are those that do not fit the other definitions of serious adverse events, but the event may jeopardize the patient and may require treatment to prevent one of the listed serious adverse events.

Adverse events will be collected if they occur between the time of informed consent and up to 30 days after the last study visit. All unresolved adverse events will be followed by the PI until resolution, the adverse event is otherwise explained, or the participant is lost to follow-up. At the last study visit, the investigator will instruct each participant to report any subsequent event that the participant, or participant's personal physician, reasonably believes may be related to the study. The investigator will notify the study sponsor of any death or adverse event occurring after the participant has discontinued participation if the event can be reasonably related to the study.

11.3 Abnormal lab values

An abnormal clinical laboratory value will be documented as an adverse event if one of the following applies:

1. The abnormality is not contradicted by a repeat test to confirm the abnormality.
2. The abnormality suggests a disease and/or organ toxicity.
3. The abnormality is of a degree that requires active management (e.g., requires a medication change, more frequent follow-up, or further diagnostic evaluation).

11.4 Hospitalization, Prolonged Hospitalization or Surgery

Any adverse event that results in hospitalization or prolonged hospitalization will be documented and reported as a serious adverse event unless specifically instructed otherwise in this protocol. Any condition responsible for surgery will be documented as an adverse event if the condition meets the criteria for an adverse event. Neither the condition, hospitalization, prolonged hospitalization, nor surgery are reported as an adverse event in the following circumstances:

1. Hospitalization or prolonged hospitalization for diagnostic or elective surgical procedures for a preexisting condition. Surgery should not be reported as an outcome of an adverse event if the purpose of the surgery was elective or diagnostic and the outcome was uneventful.
2. Hospitalization or prolonged hospitalization required to allow efficacy measurement for the study.
3. Hospitalization or prolonged hospitalization for therapy of the target disease of the study, unless it is a worsening or increase in frequency of hospital admissions as judged by the clinical investigator.

11.2.1 Serious Adverse Experience Reporting

The PI will document all SAEs that occur (whether or not related to study drug) on an SAE Report Form. The collection period for all SAEs will begin after informed consent is obtained until all procedures for the final study visit have been completed.

All SAE Report Forms will be reviewed by the PI. SAE reports that are both related and unexpected will be sent to FDA within one business day of learning of the event. SAE reports that are both related and unexpected will also be forwarded to the UW Institutional Review Board, in accordance with UW Standard Operating Procedures. Because this is an observational study, no data safety and monitoring board will be created for this trial. Adverse events and recruitment will be monitored by the PI and reported to the UW IRB, as described above.

12 DISCONTINUATION AND REPLACEMENT OF SUBJECTS

12.1 Early Discontinuation of Study Drug

A subject may be discontinued from study treatment at any time if the subject, the investigator, or the Sponsor feels that it is not in the subject's best interest to continue. The following is a list of possible reasons for study treatment discontinuation:

- Subject withdrawal of consent
- Subject is not compliant with study procedures
- Adverse event that in the opinion of the investigator would be in the best interest of the subject to discontinue study treatment
- Protocol violation requiring discontinuation of study treatment
- Lost to follow-up
- Sponsor request for early termination of study
- Positive pregnancy test (females)

If a subject is withdrawn from treatment due to an adverse event, the subject will be followed and treated by the Investigator until the abnormal parameter or symptom has resolved or stabilized.

All subjects who discontinue study treatment should come in for an early discontinuation visit as soon as possible and then should be encouraged to complete all remaining scheduled visits and procedures.

All subjects are free to withdraw from participation at any time, for any reason, specified or unspecified, and without prejudice.

Reasonable attempts will be made by the investigator to provide a reason for subject withdrawals. The reason for the subject's withdrawal from the study will be specified in the subject's source documents Refer to Section 10 for early termination procedures.

12.3 Withdrawal of Subjects from the Study

A subject may be withdrawn from the study at any time if the subject, the investigator, or the Sponsor feels that it is not in the subject's best interest to continue.

All subjects are free to withdraw from participation at any time, for any reason, specified or unspecified, and without prejudice.

Reasonable attempts will be made by the investigator to provide a reason for subject withdrawals. The reason for the subject's withdrawal from the study will be specified in the subject's source documents. Subjects who withdraw after Visit 2 but prior to Visit 10 will be encouraged to come in for a final visit (and the procedures to be followed would include those for their next scheduled visit).

12.4 Replacement of Subjects

Subjects who withdraw from the study treatment will be replaced.

Subjects who withdraw from the study will be replaced.

13 PROTOCOL VIOLATIONS

A protocol violation occurs when the subject or Sponsor-Investigator fails to adhere to significant protocol requirements affecting the inclusion, exclusion, subject safety and primary endpoint criteria. Protocol violations for this study include, but are not limited to, the following:

- Failure to meet inclusion/exclusion criteria
- Use of a prohibited concomitant medication
- Failure to comply with Good Clinical Practice (GCP) guidelines will also result in a protocol violation. The Sponsor-Investigator will determine if a protocol violation will result in withdrawal of a subject.

When a protocol violation occurs, it will be discussed with the Sponsor-Investigator and a Protocol Violation Form detailing the violation will be generated. This form will be signed by a Sponsor-Investigator. A copy of the form will be maintained in the regulatory binder and in the Sponsor-Investigator's files.

14 STATISTICAL METHODS AND CONSIDERATIONS

14.1 Data Sets Analyzed

All participants who receive the study drug (D₆-25(OH) D₃ Solution for Injection) will be included in the safety analysis.

14.2 Demographic and Baseline Characteristics

The following demographic variables at screening will be summarized by CKD status and race: age, gender, medical comorbidities, medications used, height, weight, body mass index, laboratory values (including calcium, phosphorus, albumin, vitamin D binding protein, parathyroid hormone, fibroblast growth factor-23), and adiposity measured by DXA.

14.3 Analysis of Primary Endpoint

We will examine the distribution of D₆-25(OH) D₃ clearance by CKD status (3 categories) and race (2 categories) using box plots and summary statistics. We will explore the distribution of D₆-25(OH) D₃ clearance by key covariates (age, gender, diabetes status, and baseline concentrations of 25(OH)D, PTH, FGF-23, and vitamin D binding protein) using locally weighted scatterplot smoothing (LOWESS), cubic splines, and box plots. We will pay particular attention to the associations of PTH and FGF-23 with D₆-25(OH) D₃ clearance, including multivariable modeling to determine independent associations. We will test associations of CKD status and race with D₆-25(OH) D₃ clearance (continuous outcome variable) using linear regression. Models will be adjusted for age, gender, diabetes status, vitamin D supplement use (within eligibility criteria), body size, baseline 25(OH)D concentration, and other covariates strongly related to D₆-25(OH) D₃ clearance. We will test whether associations of CKD status with clearance are modified by race by including a CKD x race interaction term. If we find evidence of interaction, we will analyze each CKD x race category separately. Absent

interaction, we will test associations of each exposure in parallel (adjusted for each other) to maximize precision and power.

14.4 Analysis of Secondary Endpoints

Analyses of secondary endpoints will parallel those of the primary endpoint.

14.5 Interim Analysis

This is not a clinical trial with an efficacy outcome, and it is neither randomized nor blinded. No formal interim analyses are planned.

14.6 Sample Size and Randomization

Our power calculations conservatively assume an interaction between CKD status and race, for which each cell in the 3x2 recruitment grid would require individual comparison. With this approach, we estimate that 17 people in each of two groups will provide 80% power to detect a difference in D₆-25(OH) D₃half-life of 2 days, assuming a standard deviation of 2 days. We will recruit 20 people per cell to allow for multivariable adjustment.

15 DATA COLLECTION, RETENTION AND MONITORING

15.1 Data Collection Instruments

The PI will prepare and maintain adequate and accurate source documents designed to record all observations and other pertinent data for each subject treated with the study drug. Study personnel at each site will enter data from source documents corresponding to a subject's visit into the protocol-specific electronic Case Report Form (eCRF) when the information corresponding to that visit is available. Subjects will not be identified by name in the study database or on any study documents to be collected by the Sponsor (or designee), but will be identified by a subject number. The Investigator is responsible for all information collected on subjects enrolled in this study. All data collected during the course of this study must be reviewed and verified for completeness and accuracy by the Investigator.

15.2 Data Management Procedures

The data will be entered into a validated database. Database lock will occur once quality assurance procedures have been completed.

All procedures for the handling and analysis of data will be conducted using good computing practices meeting FDA guidelines for the handling and analysis of data for clinical trials.

15.3 Data Quality Control and Reporting

After data have been entered into the study database, a system of computerized data validation checks will be implemented and applied to the database on a regular basis. Queries are entered, tracked, and resolved through the EDC system directly. The study database will be updated in accordance with the resolved queries. All changes to the study database will be documented.

15.4 Archival of Data

The database is safeguarded against unauthorized access by established security procedures; appropriate backup copies of the database and related software files will be maintained. Databases are backed up by the database administrator in conjunction with any updates or changes to the database.

At critical junctures of the protocol (e.g., production of interim reports and final reports), data for analysis is locked and cleaned per established procedures.

15.5 Availability and Retention of Investigational Records

The PI will make study data accessible to the UW monitor, other authorized representatives of the University of Washington IRB, and Regulatory Agency (e.g., FDA) inspectors upon request. A file for each subject will be maintained that includes the signed Informed Consent and HIPAA Authorization and copies of all source documentation related to that subject. The Sponsor-Investigator will ensure the reliability and availability of source documents from which the information on the CRF was derived.

All study documents (patient files, signed informed consent forms, copies of CRFs, Study File Notebook, etc.) will be maintained for at least two years after the study is completed.

15.6 Monitoring

Study Monitoring Plan. The PI will allocate adequate time for such monitoring activities. The PI will also ensure that any compliance or quality assurance reviewer is given access to all the above noted study-related documents and study related facilities (e.g. diagnostic laboratory), and has adequate space to conduct a monitoring visit, if requested.

Monitoring visits will be conducted by representatives of the University of Washington according to the U.S. CFR Title 21 Parts 50, 56, and 312 and ICH Guidelines for GCP (E6).

15.7 Subject Confidentiality

In order to maintain subject confidentiality, only a subject number will identify all study subjects on CRFs and other documentation submitted to the Sponsor.

16 ADMINISTRATIVE, ETHICAL, REGULATORY CONSIDERATIONS

The study will be conducted according to the Declaration of Helsinki, Protection of Human Volunteers (21 CFR 50), Institutional Review Boards (21 CFR 56), and Obligations of Clinical Investigators (21 CFR 312).

To maintain confidentiality, all laboratory specimens, evaluation forms, reports and other records will be identified by a coded number and initials only. All study records will be kept in a locked file cabinet and code sheets linking a patient's name to a patient identification number will be stored separately in another locked file cabinet. Clinical information will not be released without written permission of the subject, except as necessary for monitoring by the FDA. The Investigator must also comply with all applicable privacy regulations (e.g., Health Insurance Portability and Accountability Act of 1996, EU Data Protection Directive 95/46/EC).

16.1 Protocol Amendments

Any amendment to the protocol will be written by the Investigator-Sponsor. Protocol amendments will not be implemented without prior submission to FDA and prior written IRB approval except as necessary to eliminate immediate safety hazards to patients. A protocol amendment intended to eliminate an apparent immediate hazard to patients may be implemented immediately, provided the IRBs are notified within five working days.

16.2 Institutional Review Boards and Independent Ethics Committees

The protocol and consent form will be reviewed and approved by the University of Washington Institutional Review Board prior to study initiation. Serious adverse experiences regardless of causality will be reported to the IRB in accordance with the standard operating procedures and policies of the IRB, and the Investigator will keep the IRB informed as to the progress of the study. The Investigator will obtain assurance of IRB compliance with regulations.

Any documents that the IRB may need to fulfill its responsibilities (such as protocol, protocol amendments, Investigator's Brochure, consent forms, information concerning patient recruitment, payment or compensation procedures, or other pertinent information) will be submitted to the IRB. The IRB's written unconditional approval of the study protocol and the informed consent form will be in the possession of the Investigator before the study is initiated. The IRB's unconditional approval statement will be transmitted by the Investigator to the Sponsor or designee prior to the shipment of study supplies to the site. This approval must refer to the study by exact protocol title and number and should identify the documents reviewed and the date of review.

Protocol and/or informed consent modifications or changes may not be initiated without prior written IRB approval except when necessary to eliminate immediate hazards to the patients or when the change(s) involves only logistical or administrative aspects of the study. Such modifications will be submitted to the IRB and written verification that the modification was submitted and subsequently approved should be obtained.

The IRB must be informed of revisions to other documents originally submitted for review; serious and/or unexpected adverse experiences occurring during the study in accordance with the standard operating procedures and policies of the IRB; new information that may affect adversely the safety of the patients of the conduct of the study; an annual update and/or request for re-approval; and when the study has been completed.

16.3 Informed Consent Form

Informed consent will be obtained in accordance with the Declaration of Helsinki, ICH GCP, US Code of Federal Regulations for Protection of Human Subjects (21 CFR 50.25[a,b], CFR 50.27, and CFR Part 56, Subpart A), the Health Insurance Portability and Accountability Act (HIPAA, if applicable), and local regulations.

The Investigator will prepare the informed consent form, assent and HIPAA authorization and provide the documents to the Sponsor or designee for approval prior to submission to the IRB/IEC. The consent form generated by the Sponsor-Investigator must be approved by the IRB/IEC. The written consent document will embody the elements of informed consent as described in the International Conference on Harmonisation and will also comply with local regulations.

A properly executed, written, informed consent will be obtained from each subject prior to entering the subject into the trial. Information should be given in both oral and written form and subjects must be given ample opportunity to inquire about details of the study. If a subject is unable to sign the informed consent form (ICF) and the HIPAA authorization, a legal representative may sign for the subject. A copy of the signed consent form (and assent) will be given to the subject and the original will be maintained with the subject's records.

16.4 Publications

The publication or presentation of any study results shall comply with all applicable privacy laws, including, but not limited to, the Health Insurance Portability and Accountability Act of 1996.

16.5 References

1. Bosworth C, de Boer IH. Impaired vitamin D metabolism in CKD. *Seminars in nephrology*. 2013;33(2):158-68. Epub 2013/03/08. doi: 10.1016/j.semnephrol.2012.12.016. PubMed PMID: 23465502; PubMed Central PMCID: PMC3592201.
2. Vicchio D, Yergey A, O'Brien K, Allen L, Ray R, Holick M. Quantification and kinetics of 25-hydroxyvitamin D3 by isotope dilution liquid chromatography/thermospray mass spectrometry. *Biol Mass Spectrom*. 1993;22(1):53-8. doi: 10.1002/bms.1200220107. PubMed PMID: 8431502.
3. Compston JE, Merrett AL, Ledger JE, Creamer B. Faecal tritium excretion after intravenous administration of 3H-25-hydroxyvitamin D3 in control subjects and in patients with malabsorption. *Gut*. 1982;23(4):310-5. PubMed PMID: 7076008; PubMed Central PMCID: PMC1419735.
4. Davies M, Heys SE, Selby PL, Berry JL, Mawer EB. Increased catabolism of 25-hydroxyvitamin D in patients with partial gastrectomy and elevated 1,25-dihydroxyvitamin D levels. Implications for metabolic bone disease. *J Clin Endocrinol Metab*. 1997;82(1):209-12. PubMed PMID: 8989260.
5. Batchelor AJ, Compston JE. Reduced plasma half-life of radio-labelled 25-hydroxyvitamin D3 in subjects receiving a high-fibre diet. *Br J Nutr*. 1983;49(2):213-6. doi: S0007114583000318 [pii]. PubMed PMID: 6299329.
6. Gray RW, Weber HP, Dominguez JH, Lemann J. The metabolism of vitamin D3 and 25-hydroxyvitamin D3 in normal and anephric humans. *J Clin Endocrinol Metab*. 1974;39(6):1045-56. PubMed PMID: 4372244.
7. Speakman JR. The history and theory of the doubly labeled water technique. *Am J Clin Nutr*. 1998;68(4):932S-8S. PubMed PMID: 9771875.
8. Matsushita K, van der Velde M, Astor BC, Woodward M, Levey AS, de Jong PE, Coresh J, Gansevoort RT. Association of estimated glomerular filtration rate and albuminuria with all-cause and cardiovascular mortality in general population cohorts: a collaborative meta-analysis. *Lancet*. 2010;375(9731):2073-81. Epub 2010/05/21. doi: 10.1016/S0140-6736(10)60674-5. PubMed PMID: 20483451.
9. Jones G, Prosser DE, Kaufmann M. 25-Hydroxyvitamin D-24-hydroxylase (CYP24A1): its important role in the degradation of vitamin D. *Arch Biochem Biophys*. 2012;523(1):9-18. Epub 2011/11/22. doi: 10.1016/j.abb.2011.11.003. PubMed PMID: 22100522.
10. Akiyoshi-Shibata M, Sakaki T, Ohyama Y, Noshiro M, Okuda K, Yabusaki Y. Further oxidation of hydroxycalcidiol by calcidiol 24-hydroxylase. A study with the mature enzyme expressed in *Escherichia coli*. *Eur J Biochem*. 1994;224(2):335-43. Epub 1994/09/01. PubMed PMID: 7925346.
11. Sakaki T, Sawada N, Komai K, Shiozawa S, Yamada S, Yamamoto K, Ohyama Y, Inouye K. Dual metabolic pathway of 25-hydroxyvitamin D3 catalyzed by human CYP24. *Eur J Biochem*. 2000;267(20):6158-65. Epub 2000/09/30. PubMed PMID: 11012668.
12. Beckman MJ, Tadikonda P, Werner E, Prah J, Yamada S, DeLuca HF. Human 25-hydroxyvitamin D3-24-hydroxylase, a multicatalytic enzyme. *Biochemistry*. 1996;35(25):8465-72. Epub 1996/06/25. doi: 10.1021/bi960658i. PubMed PMID: 8679605.
13. Knutson JC, DeLuca HF. 25-Hydroxyvitamin D3-24-hydroxylase. Subcellular location and properties. *Biochemistry*. 1974;13(7):1543-8. Epub 1974/03/26. PubMed PMID: 4150615.
14. Annalora AJ, Goodin DB, Hong WX, Zhang Q, Johnson EF, Stout CD. Crystal structure of CYP24A1, a mitochondrial cytochrome P450 involved in vitamin D metabolism. *Journal of molecular biology*. 2010;396(2):441-51. Epub 2009/12/08. doi: 10.1016/j.jmb.2009.11.057. PubMed PMID: 19961857; PubMed Central PMCID: PMC2830900.
15. Kumar R, Schaefer J, Grande JP, Roche PC. Immunolocalization of calcitriol receptor, 24-hydroxylase cytochrome P-450, and calbindin D28k in human kidney. *The American journal of physiology*. 1994;266(3 Pt 2):F477-85. Epub 1994/03/01. PubMed PMID: 8160797.
16. Schuster I. Cytochromes P450 are essential players in the vitamin D signaling system. *Biochim Biophys Acta*. 2010. doi: S1570-9639(10)00181-0 [pii] 10.1016/j.bbapap.2010.06.022. PubMed PMID: 20619365.
17. Shinki T, Jin CH, Nishimura A, Nagai Y, Ohyama Y, Noshiro M, Okuda K, Suda T. Parathyroid hormone inhibits 25-hydroxyvitamin D3-24-hydroxylase mRNA expression stimulated by 1 alpha,25-dihydroxyvitamin D3 in rat kidney but not in intestine. *J Biol Chem*. 1992;267(19):13757-62. PubMed PMID: 1618870.

18. Akeno N, Saikatsu S, Kawane T, Horiuchi N. Mouse vitamin D-24-hydroxylase: molecular cloning, tissue distribution, and transcriptional regulation by 1 α ,25-dihydroxyvitamin D₃. *Endocrinology*. 1997;138(6):2233-40. PubMed PMID: 9165006.
19. Tomon M, Tenenhouse HS, Jones G. Expression of 25-hydroxyvitamin D₃-24-hydroxylase activity in Caco-2 cells. An in vitro model of intestinal vitamin D catabolism. *Endocrinology*. 1990;126(6):2868-75. Epub 1990/06/01. PubMed PMID: 2351099.
20. Goff JP, Reinhardt TA, Engstrom GW, Horst RL. Effect of dietary calcium or phosphorus restriction and 1,25-dihydroxyvitamin D administration on rat intestinal 24-hydroxylase. *Endocrinology*. 1992;131(1):101-4. Epub 1992/07/01. PubMed PMID: 1611988.
21. Akeno N, Saikatsu S, Kawane T, Horiuchi N. Mouse vitamin D-24-hydroxylase: molecular cloning, tissue distribution, and transcriptional regulation by 1 α ,25-dihydroxyvitamin D₃. *Endocrinology*. 1997;138(6):2233-40. Epub 1997/06/01. PubMed PMID: 9165006.
22. Liu PT, Stenger S, Li H, Wenzel L, Tan BH, Krutzik SR, Ochoa MT, Schaubert J, Wu K, Meinken C, Kamen DL, Wagner M, Bals R, Steinmeyer A, Zugel U, Gallo RL, Eisenberg D, Hewison M, Hollis BW, Adams JS, Bloom BR, Modlin RL. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science (New York, NY)*. 2006;311(5768):1770-3. PubMed PMID: 16497887.
23. Flanagan JN, Zheng S, Chiang KC, Kittaka A, Sakaki T, Nakabayashi S, Zhao X, Spanjaard RA, Persons KS, Mathieu JS, Holick MF, Chen TC. Evaluation of 19-nor-2 α -(3-hydroxypropyl)-1 α ,25-dihydroxyvitamin D₃ as a therapeutic agent for androgen-dependent prostate cancer. *Anticancer research*. 2009;29(9):3547-53. Epub 2009/08/12. PubMed PMID: 19667147.
24. Stubbs JR, Idiculla A, Slusser J, Menard R, Quarles LD. Cholecalciferol supplementation alters calcitriol-responsive monocyte proteins and decreases inflammatory cytokines in ESRD. *Journal of the American Society of Nephrology : JASN*. 2010;21(2):353-61. Epub 2009/12/17. doi: 10.1681/ASN.2009040451. PubMed PMID: 20007751; PubMed Central PMCID: PMC2834546.
25. Gal-Tanamy M, Bachmetov L, Ravid A, Koren R, Erman A, Tur-Kaspa R, Zemel R. Vitamin D: an innate antiviral agent suppressing hepatitis C virus in human hepatocytes. *Hepatology*. 2011;54(5):1570-9. Epub 2011/07/28. doi: 10.1002/hep.24575. PubMed PMID: 21793032.
26. Koszewski NJ, Horst RL, Goff JP. Importance of apical membrane delivery of 1,25-dihydroxyvitamin D₃ to vitamin D-responsive gene expression in the colon. *Am J Physiol Gastrointest Liver Physiol*. 2012. Epub 2012/07/28. doi: 10.1152/ajpgi.00149.2012. PubMed PMID: 22837344.
27. Goff JP, Koszewski NJ, Haynes JS, Horst RL. Targeted delivery of vitamin D to the colon using beta-glucuronides of vitamin D: therapeutic effects in a murine model of inflammatory bowel disease. *Am J Physiol Gastrointest Liver Physiol*. 2012;302(4):G460-9. Epub 2011/11/25. doi: 10.1152/ajpgi.00156.2011. PubMed PMID: 22114117.
28. Bacchetta J, Sea JL, Chun RF, Lisse TS, Wesseling-Perry K, Gales B, Adams JS, Salusky IB, Hewison M. FGF23 inhibits extra-renal synthesis of 1,25-dihydroxyvitamin D in human monocytes. *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research*. 2012. Epub 2012/08/14. doi: 10.1002/jbmr.1740. PubMed PMID: 22886720.
29. Swami S, Krishnan AV, Wang JY, Jensen K, Horst R, Albertelli MA, Feldman D. Dietary vitamin D(3) and 1,25-dihydroxyvitamin D(3) (calcitriol) exhibit equivalent anticancer activity in mouse xenograft models of breast and prostate cancer. *Endocrinology*. 2012;153(6):2576-87. Epub 2012/03/29. doi: 10.1210/en.2011-1600. PubMed PMID: 22454149; PubMed Central PMCID: PMC3359605.
30. Sacco RE, Nonnecke BJ, Palmer MV, Waters WR, Lippolis JD, Reinhardt TA. Differential expression of cytokines in response to respiratory syncytial virus infection of calves with high or low circulating 25-hydroxyvitamin D₃. *PLoS One*. 2012;7(3):e33074. Epub 2012/03/14. doi: 10.1371/journal.pone.0033074. PubMed PMID: 22412984; PubMed Central PMCID: PMC3297628.
31. Tarroni P, Villa I, Mrak E, Zolezzi F, Mattioli M, Gattuso C, Rubinacci A. Microarray analysis of 1,25(OH)(2)D(3) regulated gene expression in human primary osteoblasts. *J Cell Biochem*. 2012;113(2):640-9. Epub 2011/10/01. doi: 10.1002/jcb.23392. PubMed PMID: 21956231.
32. Alimirah F, Peng X, Yuan L, Mehta RR, von Knethen A, Choubey D, Mehta RG. Crosstalk between the peroxisome proliferator-activated receptor gamma (PPARgamma) and the vitamin D receptor (VDR) in human breast cancer cells: PPARgamma binds to VDR and inhibits 1 α ,25-dihydroxyvitamin D(3) mediated

- transactivation. *Exp Cell Res.* 2012;318(19):2490-7. Epub 2012/08/14. doi: 10.1016/j.yexcr.2012.07.020. PubMed PMID: 22884583.
33. Jiang Y, Fleet JC. Phorbol esters enhance 1alpha,25-dihydroxyvitamin D3-regulated 25-hydroxyvitamin D-24-hydroxylase (CYP24A1) gene expression through ERK-mediated phosphorylation of specific protein 3 (Sp3) in Caco-2 cells. *Mol Cell Endocrinol.* 2012;361(1-2):31-9. Epub 2012/08/09. doi: 10.1016/j.mce.2012.03.008. PubMed PMID: 22871965; PubMed Central PMCID: PMC3414851.
34. Liu K, Meng H, Hou J. Characterization of the autocrine/paracrine function of vitamin D in human gingival fibroblasts and periodontal ligament cells. *PLoS One.* 2012;7(6):e39878. Epub 2012/07/05. doi: 10.1371/journal.pone.0039878. PubMed PMID: 22761920; PubMed Central PMCID: PMC3382579.
35. Bennett RG, Wakeley SE, Hamel FG, High RR, Korch C, Goldner WS. Gene expression of vitamin d metabolic enzymes at baseline and in response to vitamin d treatment in thyroid cancer cell lines. *Oncology.* 2012;83(5):264-72. Epub 2012/09/21. doi: 10.1159/000342093. PubMed PMID: 22992568.
36. Bosworth CR, Levin G, Robinson-Cohen C, Hoofnagle AN, Ruzinski J, Young B, Schwartz SM, Himmelfarb J, Kestenbaum B, de Boer IH. The serum 24,25-dihydroxyvitamin D concentration, a marker of vitamin D catabolism, is reduced in chronic kidney disease. *Kidney international.* 2012. Epub 2012/06/01. doi: 10.1038/ki.2012.193. PubMed PMID: 22648296.
37. de Boer IH, Sachs MC, Cleary PA, Hoofnagle AN, Lachin JM, Molitch ME, Steffes MW, Sun W, Zinman B, Brunzell JD. Circulating vitamin D metabolites and kidney disease in type 1 diabetes. *The Journal of clinical endocrinology and metabolism.* 2012;97(12):4780-8. Epub 2012/09/20. doi: 10.1210/jc.2012-2852. PubMed PMID: 22990096; PubMed Central PMCID: PMC3513544.
38. Leeuwenkamp OR, van der Wiel HE, Lips P, van der Vijgh WJ, Barto R, Greuter H, Netelenbos JC. Human pharmacokinetics of orally administered (24 R)-hydroxycalcidiol. *Eur J Clin Chem Clin Biochem.* 1993;31(7):419-26. Epub 1993/07/01. PubMed PMID: 8399781.
39. Horst RL, Littledike ET, Gray RW, Napoli JL. Impaired 24,25-dihydroxyvitamin D production in anephric human and pig. *The Journal of clinical investigation.* 1981;67(1):274-80. Epub 1981/01/01. doi: 10.1172/JCI110023. PubMed PMID: 6969731; PubMed Central PMCID: PMC371597.
40. Zerwekh JE, McPhaul JJ, Jr., Parker TF, Pak CY. Extra-renal production of 24,25-dihydroxyvitamin D in chronic renal failure during 25 hydroxyvitamin D3 therapy. *Kidney international.* 1983;23(2):401-6. Epub 1983/02/01. PubMed PMID: 6601738.
41. Lambert PW, Stern PH, Avioli RC, Brackett NC, Turner RT, Greene A, Fu IY, Bell NH. Evidence for extrarenal production of 1 alpha ,25-dihydroxyvitamin D in man. *The Journal of clinical investigation.* 1982;69(3):722-5. Epub 1982/03/01. PubMed PMID: 6895901; PubMed Central PMCID: PMC371031.
42. Shimada T, Hasegawa H, Yamazaki Y, Muto T, Hino R, Takeuchi Y, Fujita T, Nakahara K, Fukumoto S, Yamashita T. FGF-23 is a potent regulator of vitamin D metabolism and phosphate homeostasis. *J Bone Miner Res.* 2004;19(3):429-35. doi: 10.1359/JBMR.0301264. PubMed PMID: 15040831.
43. Laha TJ, Strathmann FG, Wang Z, de Boer IH, Thummel KE, Hoofnagle AN. Characterizing Antibody Cross-Reactivity for Immunoaffinity Purification of Analytes prior to Multiplexed Liquid Chromatography-Tandem Mass Spectrometry. *Clinical Chemistry.* 2012. Epub 2012/09/13. doi: 10.1373/clinchem.2012.185827. PubMed PMID: 22968104.
44. Haddad JG, Min C, Mendelsohn M, Slatopolsky E, Hahn TJ. Competitive protein-binding radioassay of 24,25-dihydroxyvitamin D in sera from normal and anephric subjects. *Arch Biochem Biophys.* 1977;182(2):390-5. doi: 0003-9861(77)90519-7 [pii]. PubMed PMID: 900941.
45. Horst RL, Littledike ET, Gray RW, Napoli JL. Impaired 24,25-dihydroxyvitamin D production in anephric human and pig. *J Clin Invest.* 1981;67(1):274-80. doi: 10.1172/JCI110023. PubMed PMID: 6969731; PubMed Central PMCID: PMC371597.
46. Zerwekh JE, McPhaul JJ, Parker TF, Pak CY. Extra-renal production of 24,25-dihydroxyvitamin D in chronic renal failure during 25 hydroxyvitamin D3 therapy. *Kidney Int.* 1983;23(2):401-6. PubMed PMID: 6601738.
47. Weisman Y, Eisenberg Z, Leib L, Harell A, Shasha SM, Edelstein S. Serum concentrations of 24,25-dihydroxy vitamin D in different degrees of chronic renal failure. *Br Med J.* 1980;281(6242):712-3. PubMed PMID: 7427411; PubMed Central PMCID: PMC3713984.

48. Koenig KG, Lindberg JS, Zerwekh JE, Padalino PK, Cushner HM, Copley JB. Free and total 1,25-dihydroxyvitamin D levels in subjects with renal disease. *Kidney Int.* 1992;41(1):161-5. PubMed PMID: 1593853.
49. Gray RW, Weber HP, Dominguez JH, Lemann J, Jr. The metabolism of vitamin D3 and 25-hydroxyvitamin D3 in normal and anephric humans. *The Journal of clinical endocrinology and metabolism.* 1974;39(6):1045-56. Epub 1974/12/01. PubMed PMID: 4372244.
50. Wang Z, Lin YS, Dickmann LJ, Poulton EJ, Eaton DL, Lampe JW, Shen DD, Davis CL, Shuhart MC, Thummel KE. Enhancement of hepatic 4-hydroxylation of 25-hydroxyvitamin D3 through CYP3A4 induction in vitro and in vivo: implications for drug-induced osteomalacia. *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research.* 2013;28(5):1101-16. Epub 2012/12/06. doi: 10.1002/jbmr.1839. PubMed PMID: 23212742; PubMed Central PMCID: PMC3609874.
51. Wang Z, Schuetz EG, Xu Y, Thummel KE. Interplay between vitamin D and the drug metabolizing enzyme CYP3A4. *The Journal of steroid biochemistry and molecular biology.* 2013;136:54-8. Epub 2012/09/19. doi: 10.1016/j.jsbmb.2012.09.012. PubMed PMID: 22985909; PubMed Central PMCID: PMC3549031.
52. Bell NH, Greene A, Epstein S, Oexmann MJ, Shaw S, Shary J. Evidence for alteration of the vitamin D-endocrine system in black s. *J Clin Invest.* 1985;76(2):470-3. Epub 1985/08/01. doi: 10.1172/JCI111995. PubMed PMID: 3839801.
53. Laha TJ, Strathmann FG, Wang Z, de Boer IH, Thummel KE, Hoofnagle AN. Characterizing antibody cross-reactivity for immunoaffinity purification of analytes prior to multiplexed liquid chromatography-tandem mass spectrometry. *Clin Chem.* 2012;58(12):1711-6. doi: 10.1373/clinchem.2012.185827. PubMed PMID: 22968104; PubMed Central PMCID: PMC3731945.
54. Strathmann FG, Laha TJ, Hoofnagle AN. Quantification of 1alpha,25-dihydroxy vitamin D by immunoextraction and liquid chromatography-tandem mass spectrometry. *Clinical Chemistry.* 2011;57(9):1279-85. Epub 2011/07/20. doi: 10.1373/clinchem.2010.161174. PubMed PMID: 21768219; PubMed Central PMCID: PMC3261234.
55. Bosworth CR, Levin G, Robinson-Cohen C, Hoofnagle AN, Ruzinski J, Young B, Schwartz SM, Himmelfarb J, Kestenbaum B, de Boer IH. The serum 24,25-dihydroxyvitamin D concentration, a marker of vitamin D catabolism, is reduced in chronic kidney disease. *Kidney international.* 2012;82(6):693-700. Epub 2012/06/01. doi: 10.1038/ki.2012.193. PubMed PMID: 22648296; PubMed Central PMCID: PMC3434313.
56. de Boer IH, Sachs MC, Chonchol M, Himmelfarb J, Hoofnagle AN, Ix JH, Kremersdorf RA, Lin YS, Mehrotra R, Robinson-Cohen C, Siscovick DS, Steffes MW, Thummel KE, Tracy RP, Wang Z, Kestenbaum B. Estimated GFR and circulating 24,25-dihydroxyvitamin D3 concentration: a participant-level analysis of 5 cohort studies and clinical trials. *Am J Kidney Dis.* 2014;64(2):187-97. doi: 10.1053/j.ajkd.2014.02.015. PubMed PMID: 24703961; PubMed Central PMCID: PMC4111986.
57. Wang Z, Senn T, Kalthorn T, Zheng XE, Zheng S, Davis CL, Hebert MF, Lin YS, Thummel KE. Simultaneous measurement of plasma vitamin D(3) metabolites, including 4beta,25-dihydroxyvitamin D(3), using liquid chromatography-tandem mass spectrometry. *Anal Biochem.* 2011;418(1):126-33. Epub 2011/07/26. doi: 10.1016/j.ab.2011.06.043. PubMed PMID: 21784054; PubMed Central PMCID: PMC3164754.

APPENDIX 1. SCHEDULE OF STUDY PROCEDURES

Procedure	VISIT 1 SCREENING	VISIT 2 BASELINE (DAY 0)	VISIT 3 DAY 1	VISIT 4 DAY 4	VISIT 5 DAY 7	VISIT 6 DAY 14	VISIT 7 DAY 21	VISIT 8 DAY 28	VISIT 9 DAY 42	VISIT 10 DAY 56
Informed Consent	X									
Focused Medical History	X									
Medical History		X								
Physical Exam and Vital Signs	X	X								
Medications Inventory	X	X								X
Blood Samples*	X	X	X	X	X	X	X	X	X	X
Urine collections	X									
24-hour urine collections		X	X							
DNA collection		X								
Physical activity and food frequency questionnaires		X								
Accelerometer			X		X					
DXA		X								
Study Drug Administration		X								

*Blood Draws Detailed in Appendix 2.

APPENDIX 2. DRAFT BLOOD DRAW TYPES AND VOLUMES

Blood sample	VISIT 1 SCREENING	VISIT 2 BASELINE (DAY 0)	VISIT 3 DAY 1	VISIT 4 DAY 4	VISIT 5 DAY 7	VISIT 6 DAY 14	VISIT 7 DAY 21	VISIT 8 DAY 28	VISIT 9 DAY 42	VISIT 10 DAY 56	TOTAL VOLUME (ML)
Clinical lab: CBC (3 mL EDTA-plasma [purple top])	X										3
Clinical lab: chemistries (4 mL serum [lime green top, PST])	X	X			X						12
Clinical lab: total 25(OH)D (4 mL serum [lime green top, PST])	X										4
Clinical lab: PTH*		X			X						0*
Vitamin D metabolites (10 mL serum [red top])		X**	X	X	X	X	X	X	X	X	114
Stored blood (10 mL EDTA-plasma [purple top])		X									10
DNA***		X									0**
Total volume (mL)	11	48	10	10	14	10	10	10	10	10	143

Notes: * PTH measured with chemistries, requires no additional volume; ** 14 mL serum prior to injection of D6-25(OH)D₃ plus two 10 mL serum samples after injection of D6-25(OH)D₃;

***DNA extracted from EDTA plasma vacutainer, requires no additional volume