

**Accelerated Sarcopenia
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1. Introduction and Purpose:

Accelerated Sarcopenia. Sarcopenia is a complex geriatric syndrome characterized by the loss of skeletal muscle mass, strength and function with aging¹⁵. It is a major contributor to the cycle of frailty³⁸ and increases the risk of falls, physical dependence, disability and mortality in older adults^{15,68,82}. While the loss of muscle mass and function occurs gradually with healthy aging, diseases or other insults and injuries can accelerate sarcopenia and lead to catastrophic declines in mobility and independence⁷⁴.

Type 2 Diabetes Chronically Accelerates Sarcopenia. Chronic diseases are associated with acceleration of sarcopenia. Among these, Type 2 Diabetes Mellitus (T2DM) has been reported in longitudinal studies to hasten the loss of muscle mass and strength in older persons, even after adjustment for comorbidities, glucose control, disease length and other factors^{58,72-74}. Insulin resistance may play a role in this process⁶². However, *we do not know which specific mechanisms are involved in the acceleration of sarcopenia by T2DM*.

Knowledge Gap. Conditions that accelerate sarcopenia, such as T2DM, increase the risk of disability and dependency in older adults. However, the fundamental mechanisms by which sarcopenia accelerators induce loss of muscle mass and function are unknown. As a consequence of this knowledge gap, and in the absence of specific therapeutic targets, it is not possible to develop innovative and effective strategies for prevention and treatment of accelerated sarcopenia in older patients.

Significance. The proposed research is expected to spur a vertical advancement in our knowledge of the mechanisms that underlie accelerated sarcopenia in older adults. *This work is significant because it will lead to the identification of novel, specific targets for the development of evidence-based interventions to prevent and treat accelerated sarcopenia and maintain independence in older adults at high risk of physical dependence.*

Benefit. Mobility disability and physical dependence are major concerns for older adults, and important contributors to rising healthcare costs in the geriatric population⁵³. In the year 2010, 26.9% of the U.S. adults 65 and older had diabetes¹¹. A decline in muscle mass and muscle quality in this patient population is associated with reduced mobility⁹⁰ and increased risk of functional disability⁹¹. Hospitalization and bed rest for any cause are major events that interact with chronic conditions, further reducing function and independence in older adults^{14,47}. By identifying the fundamental mechanisms that accelerate sarcopenia in these disabling conditions, we will be able to develop novel treatments for maintenance of independence. Thus, our research has the potential to positively impact not only healthcare outcomes but also costs over the long term.

2. Background:

Low muscle mass is a basic component of sarcopenia¹⁵. The association between muscle mass and function is not linear in healthy older adults³⁵. However, conditions that accelerate sarcopenia decrease muscle mass at the same rate as strength, or faster³. A major determinant of skeletal muscle mass is contractile protein content, which is regulated by the balance between protein synthesis and proteolysis⁹⁹. Incorporation of dietary amino acids into newly synthesized proteins is a fundamental process for maintenance and growth of skeletal muscle, because it allows for replacement of the essential amino acids released by proteolysis and lost with oxidation or transport to other tissues⁹². The stimulatory effect of an amino acid/protein meal on skeletal muscle protein synthesis is large and rapid^{48,84}, and driven by essential amino acids, particularly leucine, which directly stimulate muscle protein synthesis by activating translation initiation via the mTORC1 pathway^{18,5}. To activate mTORC1 signaling and protein synthesis, amino acids must first enter the muscle cell via amino acid transport. Amino acid transport is an active process that is emerging as a key regulator of skeletal muscle protein homeostasis and anabolism⁵⁵. Amino acid transport is controlled by several factors^{1,55}. We have reported that dietary amino acid intake^{27,96}, insulin⁸⁶ and resistance exercise²⁶ can independently activate amino acid transporter expression and function in humans, and this effect is associated with increased muscle protein synthesis¹⁹.

There is growing evidence from our group^{43,75,87} and others⁸⁰ that healthy aging *per se* induces a mild but significant reduction in the response of muscle proteins to anabolic stimulation. This phenomenon has been termed “anabolic resistance” and may account for the slow loss of muscle mass with advancing age. It is characterized by a reduced anabolic response to low amino acid doses^{17,56,57,84,94} and insulin⁴³. This is a true “resistance” because the maximal anabolic response to high doses of amino acids^{83,93,96} or insulin⁴³ is preserved in healthy older adults and comparable to that observed in younger individuals.

Our preliminary data suggest that a global, underlying mechanism of accelerated sarcopenia is an increase in anabolic resistance associated with a reduction in amino acid transport. With this application we will examine how amino acid transport is impacted by conditions accelerating sarcopenia and identify factors that control it.

We propose that resistance exercise will improve the response of muscle protein anabolism to amino acids in T2DM by increasing mTORC1 signaling and amino acid transporter expression. Upstream regulatory pathways will be explored also in this aim to identify promising therapeutic targets for interventions.

Innovation

The research we propose is innovative, in our opinion, for at least three major reasons:

1. Conceptual Innovation. Our proposed studies will explore for the first time the potential global role of amino acid transport in conditions that accelerate sarcopenia chronically and acutely. By using a well described and common model of accelerated sarcopenia, T2DM, this research may also rapidly yield practical innovation that could be rapidly translated in the clinical setting.

2. Methodological Innovation. Innovation also stems from our novel integrative and interdisciplinary translational approach that combines state of the art and novel methodologies to determine amino acid transporter expression and function and their upstream regulation. These methodologies include: a 3-pool model tracer methodology to measure amino acid trafficking across vascular and intracellular compartments²⁰; molecular techniques to measure regulation of muscle protein synthesis and amino acid transport^{26,27}; and near infrared spectroscopy (NIRS) to determine the role of endothelial function and microvascular muscle perfusion^{86,87}. Exercise is a well-established means to induce muscle growth. Resistance exercise also stimulates amino acid transport and transporter expression independent of changes in amino acid availability²⁶. In this application, we will *innovatively use exercise as a tool to modify the response of amino acid transport and transporter expression* with the purpose of testing our hypotheses and identifying new therapeutic targets.

3. Discovery of Novel Molecular Targets. Our secondary outcomes will include exploration of the role of novel regulatory proteins in relation to the changes in amino acid transport associated with T2DM. The basic assumption is that chronic and acute sarcopenia accelerators may reduce amino acid transport with different mechanisms. Thus, we will determine the potential role of newly identified amino acid sensing pathways such as the Sestrins-GATOR1/2 and the lysosomal vATPase/SLC38A9/Ragulator complex in the activation, or lack of thereof, of mTORC1 signaling and amino acid transporter expression and function.

Our approach — unique and substantially different from the status quo — is expected to unravel the key underlying mechanisms responsible for accelerated sarcopenia due to T2DM. The knowledge generated by our studies is needed to develop future evidence-based clinical trials of interventions.

3. Concise Summary of Project:

Subjects. We will test our central hypothesis in community dwelling, independent older adults with non-insulin dependent T2DM. To account for screening failures and drop-outs/withdrawals we plan to enroll about 30 subjects in order to have 15 subjects completing the study. Since data previously gathered on healthy subjects will serve as controls for T2DM subjects, who are expected to be overweight or obese, we will limit inclusion to subjects with BMI 23-35 kg/m². Those who have a BMI ≥ 35 kg/m² will be considered for inclusion on a case-to-case basis, depending on whether or not

thigh adiposity would impair muscle biopsy. Morbidly obese, frail or exercise trained subjects will be excluded, as these conditions can significantly impact muscle protein metabolism^{13,79,89}, confounding the results. Subjects will be weight stable, as active weight loss in obese older adults can temporarily improve the response of muscle protein synthesis to anabolic stimuli⁸⁸. Subjects will be recruited from the UTMB Pepper Center Volunteer Registry, the Diabetes Clinic for Seniors directed by Dr. Volpi, and the UTMB Stark Diabetes Clinic.

Screening: After carefully explaining procedures and risks, informed consent will be obtained from all subjects. Eligibility will be assessed with clinical history, physical exam, questionnaires, functional testing, and a number of laboratory tests including blood count, metabolic panel, hemoglobin A1c, uric acid levels, lipid profile, thyroid function, coagulation panel, urinalysis, urine toxicology drug screening, hepatitis B and C screening, HIV test, DEXA scan, and a stress test. An OGTT will also be performed, and may involve the use of tracer methodology (U-C¹³ glucose) to better describe glucose kinetics. Habitual physical activity will be measured using both StepWatch Activity Monitors (SAM) and pedometers for approximately 1 week during the screening process.

4. Study Design. Subjects will undergo a two acute experiments, one prior to and one immediately following the completion of 13 weeks' (1 run-in week and 12 weeks progressive resistance exercise) training, in which we will test the effects of 7 g of EAA in older T2DM patients.

1) Run-In: The day prior to the experiment we will:

- Admit subjects to the ITS-CRC.
- Draw safety labs (blood count, metabolic panel, coagulation panel, urinalysis) and other individualized clinical labs necessary to follow up on safety parameters.
- Provide standardized meals
- Hold vasodilators, monitor blood pressure, and manage hypertension with other medications if necessary^{43,75,87}.

2) Metabolic Experiment. The experimental procedure is designed to measure skeletal muscle amino acid transporter expression and function, whole body and muscle amino acid kinetics, muscle protein synthesis and proteolysis. The subjects will be studied in the post-absorptive state, after an overnight stay in the ITS-CRC. After admission, subjects will be given dinner at approximately 18:00 and a snack at approximately 21:00, after which they will be allowed only water ad libitum. At approximately 05:00 we will insert: an antecubital forearm venous catheter for systemic infusions; at approximately 07:00 we will insert a retrograde venous catheter in the opposite hand placed in a heating pad for arterialized blood draws; at approximately 09:00 we will attempt to place a pediatric central catheter in the femoral vein for blood draws. Subsequently, we will start a primed-continuous infusion of L-[ring-¹³C₆]-phenylalanine, L-[5,5,5-²H₃]-leucine, and [1-¹³C]-glycine to measure muscle protein turnover as well as the amino acid transport rates through LAT1 (Phe, Leu), SNAT2 (Gly), and PAT1 (Gly)^{55,93}. To reduce risks we will use arterialized blood samples, rather than arterial blood, to measure arterial amino acid concentrations and enrichments. This method has been previously validated for amino acid and other metabolite concentrations⁸, and we have preliminary data indicating that it is also valid for amino acid enrichments. After an approximately 4-hr baseline period, subjects will ingest the amino acid mixture, as a bolus. The mixture will contain EAA in the pattern found in animal protein²² and will be enriched with L-[ring-¹³C₆]-phenylalanine, L-[5,5,5-²H₃]-leucine to maintain the isotopic steady state²² (labeled glycine will not be added because glycine is not included in the oral mixture). Large vessel blood flow will be measured by Doppler^{86,87}. Microvascular nutritive flow is emerging as a potential regulator of muscle amino acid and protein metabolism⁸⁵⁻⁸⁷. Thus, whenever possible we will perform a Flow Mediated Dilation (FMD) test to detect endothelial function using Doppler Ultrasound and Near Infrared Spectroscopy. Frequent blood samples will be collected during the last hour of the basal period and the first and third hour of the amino acid period to measure amino acid concentrations and enrichments, glucose, insulin, cytokines and endothelin-1. T2DM is characterized by hyperglycemia, hyperinsulinemia, inflammation and endothelial dysfunction, which may have an impact on muscle protein metabolism^{69,86}. Additional serum samples will also be collected and stored for future analyses (e.g. other hormonal assays, proteomics, metabolomics). Up to two muscle biopsies will be taken in each period to measure pertinent parameters including amino

acid and protein kinetic parameters, cross-sectional area, cell signaling, novel regulatory pathways, and gene expression (see Analytical Methods). A portion of the biopsies will be collected for immunohistochemistry. A third incision may be made should the need arise. At the end of the experiment catheters will be removed and subjects will be given a meal.

3) **Resistance Training:** This experiment will involve measurement of the chronic effect of resistance exercise on the response of muscle amino acid transport and protein anabolism to approximately 7 g of EAA in older adults with T2DM. Within one week after the first metabolic experiment, subjects will initiate progressive, supervised resistance exercise training, 3 times per week for 12 wks. Prior to initiating the 12-week program, a run-in week will be completed, for a total of 13 weeks overall (*Table 1*). During the intervention, we will monitor physical activity with SAMs in all subjects to determine if exercise reduces habitual physical activity. If possible, we will also use continuous glucose monitors to assess changes in blood glucose control throughout the intervention.

At the end of the 12-week period, and approximately 48 hours after the last bout of exercise, we will repeat oral glucose tolerance testing. The Metabolic Experiment will be repeated the following day, to measure the effect of training on amino acid transport and muscle protein anabolism.

5. Sub-Study Procedures:

Specimens obtained during the study will be stored in the principal investigator's laboratory and may be used in future research. For example, from time to time, new laboratory tests and procedures are developed that could be used to better understand the human body during metabolism studies.

The extra muscle tissue/blood left over after the initial research study tests are done will be stored and used in future research. The samples will be used to look at muscle growth related pathways. DNA

will not be stored and no genetic testing will be done. Any future research will not involve additional specimens being taken or require any extra involvement from subjects. We will not need to contact the subjects again. The specimens will be labeled with a code number and will not directly identify any of the subjects who partake in the study.

Table 1. Progressive resistance exercise training protocol.

Resistance Exercise Training						
	Free Weights	Machine	Run-In Week	Weeks 1-4	Weeks 5-8	Weeks 9-12
Bicep curl	X	X				
Triceps Extension		X				
Chest press		X				
Lat pulldown		X	2 x 15 @ 60% 1-RM	3 x 15 @ 60% 1-RM	3 x 12 @ 65% 1-RM	3 x 10 @ 70% 1-RM
Shoulder press		X				
Leg press		X				
Leg extension		X				
Leg curl		X				
Seated calf raise		X				
crunches	-	-				

6. Criteria for Inclusion of Subjects:

- Age 60-85 yr
- Body mass index: 23-35 kg/m²

- Or BMI>35 if thigh adiposity does not impair muscle biopsy
- Stable body weight for at least 3 months
- T2DM
- Able to provide informed consent

7. Criteria for Exclusion of Subjects:

- Insulin therapy that cannot be converted to non-insulin therapies
- Significant diabetes complications
- A1c > 8%. Those excluded based on A1c will be eligible for re-evaluation.
- Impairment in Activities of Daily Living
- Score <25 on the 30-item Mini Mental State Examination
- Severe depression as determined by Geriatric Depression Scale >5
- ≥ 2 falls in the past 12 months
- Weight loss ≥ 10% bodyweight in the past 12 months
- Moderate or High activity level as indicated by the International Physical Activity Questionnaire Short Form
- Significant Cardiac abnormalities considered exclusionary by the study physician (e.g., CHF, CAD)
- Vascular disease
- Use of anticoagulant therapy. (e.g., warfarin, heparin, Pradaxa)
- Uncontrolled blood pressure: systolic >160 mmHg or diastolic > 100 mmHg
- History of stroke with motor disability
- Significant Chronic kidney disease
- Anemia
- Liver disease (AST/ALT 2.5 times above the normal limit)
- Significant Respiratory disease (acute upper respiratory infection, chronic lung disease)
- Active cancer or infection
- Recent (within 3 months), chronic treatment with anabolic steroids, systemic corticosteroids or estrogen.
- HIV or active hepatitis B or C
- Alcohol (CAGE questionnaire) or drug abuse (urine screening)
- Inability to perform OGTT (i.e., bad veins)
- History of mastectomy with axillary lymph node dissection
- Any other condition or event considered exclusionary by the PI and faculty physician

8. Sources of Research Material:

Participants will provide verbal and written data specifically for research purposes. The study physician will have access to UTMB EPIC for subjects Measurement equipment (DEXA, stress test, food diary, timers for functional tests) will capture numerical and qualitative data. The SAM device will collect data on steps. Study data will also be securely stored on REDCAP. Only trained study staff will have access to study data. These data will be confidential.

9. Recruitment Methods and Consenting Process:

Subjects will be recruited through an institutional review-board approved advertisement that will be posted at UTMB, in local newspapers, and at social centers such as the library and recreation centers. We will also utilize university announcements, flyers, patient letters, and word of mouth. After responding to ads, subjects will be pre-screened and asked to come in for a screening visit at the ITS-CRC. If they qualify, they will be offered an opportunity to participate in the study. This will happen on a "first come first served" basis. We will study a normal healthy population between the ages of 60-85 representative of the ethnic makeup of the United States. Prior to screening at the ITS-CRC, subjects will be asked to provide informed consent. During the consenting process, each subject will have a private room. The door will be closed during the consenting process. We will take

all measures to ensure that subject privacy is maintained. The details of the experimental procedures, including risks and benefits, will be explained by a research team member at the time of initial screening at the ITS-CRC. At this time, the subject will be given ample time to read the consent form, ask questions and discuss any concerns. After the subject has been satisfied, the subject will be given an opportunity to sign the consent form. At every study encounter, each subject will be asked if they are still willing to participate in the study. Each subject will be asked if they have any questions, concerns, complaints, or comments. If the subjects have any of those listed, they will be appropriately addressed by the PI.

10. Potential Risks:

Possible risks and discomforts a subject might be exposed to during the course of the study include:

Psychological Stress: Some of the questions asked as part of the study may make the subject feel uncomfortable.

Positive screening tests: One or more of the screening tests might be positive for a medical condition. This information might be upsetting for the subject. The PI and study physician, Dr. Elena Volpi, will provide counselling to the subject in the event that a screening test is positive for a medical condition or a blood level is abnormal. The likelihood of this is low.

Fasting: Risks of fasting include hypoglycemia, dizziness, nausea, and light headedness. Risk is low.

Blood draws: The subjects will undergo blood draws during the course of the study. The total amount of blood drawn will be approximately 170 ml for each of the two metabolic study days. The risks are pain, bruising and infection. People with borderline anemia might become more anemic with the repeated blood draws. Fainting is also a risk as some people faint at the sight of needles and/or blood. The risk of this event is low.

Anemia: Repeated blood draws or excessive sampling volume increases the risk of anemia. This is unlikely as the blood collected is less than a blood donation.

Muscle biopsies: The risks of this procedure are pain, bleeding, bruising, infection, and a little scar at the site of the incision. The risks are low. The biopsy scar is in the shape and size of this " _ " Some people may also faint at the sight of needles and/or blood. Up to 50% of subjects experience some soreness at the site of the biopsy up to 48 hours after it is done.

Stable Isotopes Infusion: The only possible risks are sepsis (blood infection) and fever, but the risk of these is so small that the precise number is unknown. The isotopes are tested in order to assure their sterility (absence of microorganisms, such as bacteria, that may cause infections and fever). Antibiotics can treat these infections should they occur.

DEXA Scan: Each DEXA scan will expose the subject to a small amount of radiation. Exposure to radiation can increase the risk of cancer and birth defects. The amount of radiation that a subject will be exposed to is so small that the exact risk is unknown.

Femoral Venous Catheter: We will insert only a pediatric catheter in the femoral vein. The risks are the following: (A) Trauma to structures adjacent to the targeted vein (hematoma, nerve injury) is the most commonly recognized complication/error during the cannulation process (1.9-3.6% incidence during central venous cannulation are generally reported). Traumas rarely are serious, but the experience of the operator and the use of ultra-sound assistance can bring the risk to virtually 0%. (B) Complications related to the presence of the catheters, i.e., infection, thrombosis and

thromboembolism. For patients with central venous catheters (CVC), an infection rate of 1.4 per 1,000 catheters is reported. However, most patients with CVC are immunosuppressed because of their underlying disease, and therefore have a higher risk of infections than our research volunteers. Infection in the immediate post-insertion period (3-5 days) is commonly due to infection of the subcutaneous track caused by intraoperative contamination and as such are more easily diagnosed and treated than the feared systemic infections more prone to appear later. Catheter-related thrombosis occurs in 3.7-10% of patients. This complication is more likely to occur in patients with specific thromboembolic risk. Factors that contribute to thrombosis in patients are hypercoagulability (malignant tumors), venous irritation caused by chemotherapeutic agents and hyperalimentation fluids. The risk is moderate.

Stress Test/Exercise: Risks may include muscle tightness, soreness, fatigue and rarely a muscle strain or tear. In older individuals, there is a small risk of myocardial infarction during maximal exercise tests. This is generally assumed to be approximately 1 in 10,000. Stress testing with ECG monitoring will be done at screening in the older subjects under the supervision of a MD to exclude subjects with asymptomatic heart disease. The exercise training sessions will be performed at the UTMB Center for Recovery, Physical Activity & Nutrition under the supervision of a CPR trained and BCLS licensed physical therapist or certified trainer. The exercise facility is equipped with emergency defibrillators. Muscle soreness or cramps, the most likely side effects of stress and muscle performance testing, can be reduced or avoided with an adequate warm-up and ramped up exercise training, both of which are integrated into our research protocol.

Strength Testing: The subject will experience soreness and tenderness in the muscles for 2 to 3 days following exercise. This is normal and is one of the ways in which the human body responds to exercise in order for the muscle to grow. A cardiovascular event such as a heart attack is a very rare potential risk.

Weight Training: It is rare but also possible that subjects may experience a major injury to their back, ligaments, joints, muscles, or tendons during exercise. This is a potential risk with any physical activity.

Near Infrared Spectrometry (NIRS): No additional protection is necessary. The effect of heat due to near infrared light absorption is low – less than 0.5°C.

Confidentiality: There is a general risk of disclosure of personal sensitive data in a clinical investigation. The risk is minimal.

11. Subject Safety and Data Monitoring:

Data Safety and Monitoring Plan. The ultimate responsibility for data and safety monitoring rests with Dr. Volpi. The risk level associated with this study is estimated to be moderate.

Our data safety and monitoring plan will include:

1. Review during of screening results and any other available data at the weekly investigators' meeting. A signed eligibility form will be kept on file for each subject.
2. A quarterly review of data safety and interim data analysis performed by the PI.
3. An annual review performed by the UTMB IRB.
4. Semi-annual patient safety review by a Medical Monitor, Mukaila Raji, MD, Chief of the Division of Geriatric Medicine, UTMB, will be initiated after completion of the first 10 subjects or after the occurrence of the first SAE whichever comes first. Minutes of the semi-annual Medical Monitor review will be kept on file and submitted to UTMB's IRB in a Miscellaneous form within a two week period.

Careful monitoring of the recruitment, enrollment, retention, adverse events, and study procedures will help protect the safety of study subjects, the quality of data, and the integrity of the study. As part of the safety plan for this study, Dr. Volpi, who is ultimately responsible for the medical

management of the subjects, will review each subject's records to ensure that appropriate mechanisms to protect the safety of study participants are being followed; that protocol requirements are being adhered to; and that data are accurate, complete, and secure. Subject records include consent forms, case report forms, flow of data forms, laboratory specimen records, inclusion/exclusion forms, adverse event logs, and medical charts.

Non-serious, anticipated adverse events (AE) include hematoma, pain, infection, syncope, and loss of subject confidentiality. If an anticipated adverse event occurs, Dr. Volpi will be immediately notified and a note will be entered into the subject's CRC chart. Dr. Volpi will be responsible for evaluating each AE and determining attribution as well as the impact of the AE on the risk/benefit ratio.

All unanticipated, serious, fatal and/or life-threatening adverse events will be reported to the IRB within 24h of occurrence. The IRB and the PI will be responsible for determining whether modifications to the protocol and consent form are required. If a determination is made that participants are found to be exposed to excessive risks in relation to anticipated benefits, the study will be immediately suspended. Studies will not resume until modifications are made that are deemed to result in an acceptable risk/benefit ratio by the PI and the IRB. Aggregate reports of adverse events will be prepared as mandated and forwarded to the IRB and ITS-CRC for review.

Plan for Adverse Event Reporting. The PI will be responsible for evaluating each adverse event and determining attribution as well as the impact on the risk/benefit ratio, using the following Grading and Attribution criteria:

Mild – transient laboratory test alterations that do not suggest injury; discomfort noted but no disruption of daily activities; no therapy, or only symptomatic therapy required.

Moderate – Laboratory test alterations indicating injury without long-term risk, discomfort sufficient to modify normal daily activity, specific therapy required (i.e., more than symptomatic).

Serious – laboratory tests indicating a serious health threat or permanent injury, incapacity, inability to work, or to perform normal daily activity, hospitalization required or prolonged, emergency treatment required, life-threatening events, death.

The attribution scale assesses the relation of the event to the study procedures. Dr. Volpi will judge whether an adverse event is: 1) *not related*; 2) *possibly related*; 3) *probably related*; or 4) *definitely related to the study interventions*.

Data Safety and Monitoring Board. We will consult our IRB at the time of protocol submission to determine if we need to establish a DSMB. Should a DSMB be required, we will establish it including physician-investigators from the Texas Regional CTSA Consortium and other Claude D. Pepper Centers.

12. Procedures to Maintain Confidentiality:

All study data will be collected in REDCap by the research team. Paper data flow sheets or case report forms will be stored in locked file cabinets in a secure area of the PI's recruitment office. Interim data will be reviewed by the PI. Any appearance of a Conflict of Interest by the research team will be avoided by the periodic monitoring by UTMB's Conflict of Interest Committee. The subjects' right to confidentiality will be protected at all times and no subject will be identified by name in any publication that results from this research.

13. Potential Benefits:

The only potential benefits for the participation in this protocol are related to the knowledge of the results of the screening tests that will be released to the volunteer, and the availability of nutritional counseling at the conclusion of the study. T2DM patients who participate in progressive resistance exercise training may experience an improvement in their blood glucose control, strength and endurance with training. No other benefits are expected for the subjects. The knowledge collected from the present studies may be of benefit to society, as these studies will help develop the scientific basis for designing treatments for reducing loss in muscle mass and function in older subjects with T2DM.

Importance of the Knowledge to be Gained. These studies will allow us to identify the biological mechanisms by T2DM accelerate sarcopenia in older individuals. Since T2DM is quite common in older adults and increases the risk of physical dependence, the knowledge to be gained will be very important to develop interventions to maintain independence in older adults.

REFERENCES CITED

1. Adams CM: Role of the transcription factor ATF4 in the anabolic actions of insulin and the anti-anabolic actions of glucocorticoids. *J Biol Chem* 282:16744-16753, 2007
2. Albert JD, Legaspi A, Horowitz GD, Tracey KJ, Brennan MF, Lowry SF: Extremity amino acid metabolism during starvation and intravenous refeeding in humans. *Am J Physiol* 251:E604-E610, 1986
3. Alley DE, Koster A, Mackey D, Cawthon P, Ferrucci L, Simonsick EM, Yu B, Hardy S, Goodpaster B, Sarkisian C, Houston DK, Kritchevsky SB, Cummings S, Lee JS, Tylavsky FA, Newman A, Harris T: Hospitalization and change in body composition and strength in a population-based cohort of older persons. *J Am Geriatr Soc* 58:2085-2091, 2010
4. American Diabetes Association: Standards of medical care in diabetes--2013. *Diabetes Care* 36 Suppl 1:S11-S66, 2013
5. Bell JA, Volpi E, Fujita S, Cadenas JG, Sheffield-Moore M, Rasmussen BB: Skeletal muscle protein anabolic response to increased energy and insulin is preserved in poorly controlled type 2 diabetes. *J Nutr* 136:1249-1255, 2006
6. Bickel CS, Cross JM, Bamman MM: Exercise dosing to retain resistance training adaptations in young and older adults. *Med Sci Sports Exerc* 43:1177-1187, 2011
7. Biolo G, Declan Fleming RY, Wolfe RR: Physiologic hyperinsulinemia stimulates protein synthesis and enhances transport of selected amino acids in human skeletal muscle. *J Clin Invest* 95:811-819, 1995
8. Brooks DC, Black PR, Arcangeli MA, Aoki tT, Wilmore DW: The heated dorsal hand vein: an alternative arterial sampling site. *JPEN J Parenter Enteral Nutr* 13:102-105, 1989
9. Castaneda C, Layne JE, Munoz-Orians L, Gordon PL, Walsmith J, Foldvari M, Roubenoff R, Tucker KL, Nelson ME: A randomized controlled trial of resistance exercise training to improve glycemic control in older adults with type 2 diabetes. *Diabetes Care* 25:2335-2341, 2002
10. Cauza E, Hanusch-Enserer U, Strasser B, Ludvik B, Metz-Schimmerl S, Pacini G, Wagner O, Georg P, Prager R, Kostner K, Dunky A, Haber P: The relative benefits of endurance and strength training on the metabolic factors and muscle function of people with type 2 diabetes mellitus. *Arch Phys Med Rehabil* 86:1527-1533, 2005
11. Centers for Disease Control and Prevention.: National diabetes fact sheet: national estimates and general information on diabetes and prediabetes in the United States, 2011. [article online], 2012. Available from http://www.cdc.gov/diabetes/pubs/pdf/ndfs_2011.pdf

12. Chantranupong L, Wolfson RL, Orozco JM, Saxton RA, Scaria SM, Bar-Peled L, Spooner E, Isasa M, Gygi SP, Sabatini DM: The Sestrins interact with GATOR2 to negatively regulate the amino-acid-sensing pathway upstream of mTORC1. *Cell Rep* 9:1-8, 2014
13. Chevalier S, Gougeon R, Nayar K, Morais JA: Frailty amplifies the effects of aging on protein metabolism: role of protein intake. *Am J Clin Nutr* 78:422-429, 2003
14. Covinsky KE, Palmer RM, Fortinsky RH, Counsell SR, Stewart AL, Kresevic D, Burant CJ, Landefeld CS: Loss of independence in activities of daily living in older adults hospitalized with medical illnesses: increased vulnerability with age. *J Am Geriatr Soc* 51:451-458, 2003
15. Cruz-Jentoft AJ, Baeyens JP, Bauer JM, Boirie Y, Cederholm T, Landi F, Martin FC, Michel JP, Rolland Y, Schneider SM, Topinkova E, Vandewoude M, Zamboni M: Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age Ageing* 39:412-423, 2010
16. Cusi K, Maezono K, Osman A, Pendergrass M, Patti ME, Pratipanawatr T, DeFronzo RA, Kahn CR, Mandarino LJ: Insulin resistance differentially affects the PI 3-kinase- and MAP kinase-mediated signaling in human muscle. *J Clin Invest* 105:311-320, 2000
17. Cuthbertson D, Smith K, Babraj J, Leese G, Waddell T, Atherton P, Wackerhage H, Taylor PM, Rennie MJ: Anabolic signaling deficits underlie amino acid resistance of wasting, aging muscle. *FASEB Journal* 19:422-424, 2005
18. Dickinson JM, Fry CS, Drummond MJ, Gundermann DM, Walker DK, Glynn EL, Timmerman KL, Dhanani S, Volpi E, Rasmussen BB: Mammalian target of rapamycin complex 1 activation is required for the stimulation of human skeletal muscle protein synthesis by essential amino acids. *J Nutr* 141:856-862, 2011
19. Dickinson JM, Volpi E, Rasmussen BB: Exercise and nutrition to target protein synthesis impairments in aging skeletal muscle. *Exerc Sport Sci Rev* 41:216-223, 2013
20. Dillon EL, Casperson SL, Durham WJ, Randolph KM, Urban RJ, Volpi E, Ahmad M, Kinsky MP, Sheffield-Moore M: Muscle protein metabolism responds similarly to exogenous amino acids in healthy younger and older adults during NO-induced hyperemia. *Am J Physiol Regul Integr Comp Physiol* 301:R1408-R1417, 2011
21. Dreyer HC, Fujita S, Glynn EL, Drummond MJ, Volpi E, Rasmussen BB: Resistance exercise increases leg muscle protein synthesis and mTOR signalling independent of sex. *Acta Physiol (Oxf)* 2010
22. Drummond MJ, Dickinson JM, Fry CS, Walker DK, Gundermann DM, Reidy PT, Timmerman KL, Markofski MM, Paddon-Jones D, Rasmussen BB, Volpi E: Bed rest impairs skeletal muscle amino acid transporter expression, mTORC1 signaling, and protein synthesis in response to essential amino acids in older adults. *Am J Physiol Endocrinol Metab* 302:E1113-E1122, 2012
23. Drummond MJ, Dreyer HC, Fry CS, Glynn EL, Rasmussen BB: Nutritional and contractile regulation of human skeletal muscle protein synthesis and mTORC1 signaling. *J Appl Physiol* 106:1374-1384, 2009
24. Drummond MJ, Dreyer HC, Pennings B, Fry CS, Dhanani S, Dillon EL, Sheffield-Moore M, Volpi E, Rasmussen BB: Skeletal muscle protein anabolic response to resistance exercise and essential amino acids is delayed with aging. *J Appl Physiol* 104:1452-1461, 2008

25. Drummond MJ, Fry CS, Glynn EL, Dreyer HC, Dhanani S, Timmerman KL, Volpi E, Rasmussen BB: Rapamycin administration in humans blocks the contraction-induced increase in skeletal muscle protein synthesis. *J Physiol* 587:1535-1546, 2009
26. Drummond MJ, Fry CS, Glynn EL, Timmerman KL, Dickinson JM, Walker DK, Gundersmann DM, Volpi E, Rasmussen BB: Skeletal muscle amino acid transporter expression is increased in young and older adults following resistance exercise. *J Appl Physiol* 111:135-142, 2011
27. Drummond MJ, Glynn EL, Fry CS, Timmerman KL, Volpi E, Rasmussen BB: An increase in essential amino acid availability upregulates amino acid transporter expression in human skeletal muscle. *Am J Physiol Endocrinol Metab* 298:E1011-E1018, 2010
28. Drummond MJ, McCarthy JJ, Fry CS, Esser KA, Rasmussen BB: Aging differentially affects human skeletal muscle microRNA expression at rest and after an anabolic stimulus of resistance exercise and essential amino acids. *Am J Physiol Endocrinol Metab* 295:E1333-E1340, 2008
29. Drummond MJ, Timmerman KL, Markofski MM, Walker DK, Dickinson JM, Jamaluddin M, Brasier AR, Rasmussen BB, Volpi E: Short-term bed rest increases TLR4 and IL-6 expression in skeletal muscle of older adults. *Am J Physiol Regul Integr Comp Physiol* 305:R216-R223, 2013
30. Ernst E, Schmidt-Pauly E, Muhlig P, Matrai A: Blood viscosity in patients with bone fractures and long term bedrest. *Br J Surg* 74:301-302, 1987
31. Ferrando AA, Lane HW, Stuart CA, Davis-Street J, Wolfe RR: Prolonged bed rest decreases skeletal muscle and whole body protein synthesis. *Am J Physiol* 270:E627-33, 1996
32. Ferrando AA, Paddon-Jones D, Hays NP, Kortebein P, Ronsen O, Williams RH, McComb A, Symons TB, Wolfe RR, Evans W: EAA supplementation to increase nitrogen intake improves muscle function during bed rest in the elderly. *Clin Nutr* 29:18-23, 2010
33. Ferrara CM, Goldberg AP, Ortmeyer HK, Ryan AS: Effects of aerobic and resistive exercise training on glucose disposal and skeletal muscle metabolism in older men. *J Gerontol A Biol Sci Med Sci* 61:480-487, 2006
34. Fiatarone MA, O'Neill EF, Ryan ND, Clements KM, Solares GR, Nelson, ME, Roberts SB, Kehayias JJ, Lipsitz LA, Evans WJ: Exercise training and nutritional supplementation for physical frailty in very elderly people. *N Engl J Med* 330:1769-1775, 1994
35. Fielding RA, Vellas B, Evans WJ, Bhasin S, Morley JE, Newman AB, Abellan van KG, Andrieu S, Bauer J, Breuille D, Cederholm T, Chandler J, De MC, Donini L, Harris T, Kannt A, Keime GF, Onder G, Papanicolaou D, Rolland Y, Rooks D, Sieber C, Souhami E, Verlaan S, Zamboni M: Sarcopenia: an undiagnosed condition in older adults. Current consensus definition: prevalence, etiology, and consequences. International working group on sarcopenia. *J Am Med Dir Assoc* 12:249-256, 2011
36. Fisher SR, Goodwin JS, Protas EJ, Kuo YF, Graham JE, Ottenbacher KJ, Ostir GV: Ambulatory activity of older adults hospitalized with acute medical illness. *J Am Geriatr Soc* 59:91-95, 2011
37. Food and Nutrition Board, Institute of Medicine: Protein and Amino Acids. In *Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acid, Cholesterol, Protein, and Amino*

Acids (Macronutrients). Institute of Medicine, Ed. Washington, The National Academies Press, 2005, p. 589-768

38. Fried LP, Hadley EC, Walston JD, Newman AB, Guralnik JM, Studenski S, Harris TB, Ershler WB, Ferrucci L: From bedside to bench: research agenda for frailty. *Sci Aging Knowledge Environ* 2005:e24, 2005
39. Fry CS, Drummond MJ, Glynn EL, Dickinson JM, Gundermann DM, Timmerman KL, Walker DK, Dhanani S, Volpi E, Rasmussen BB: Aging impairs contraction-induced human skeletal muscle mTORC1 signaling and protein synthesis. *Skelet Muscle* 1:11, 2011
40. Fry CS, Drummond MJ, Glynn EL, Dickinson JM, Gundermann DM, Timmerman KL, Walker DK, Volpi E, Rasmussen BB: Skeletal muscle autophagy and protein breakdown following resistance exercise are similar in younger and older adults. *J Gerontol A Biol Sci Med Sci* 68:599-607, 2013
41. Fry CS, Glynn EL, Drummond MJ, Timmerman KL, Fujita S, Abe T, Dhanani S, Volpi E, Rasmussen BB: Blood flow restriction exercise stimulates mTORC1 signaling and muscle protein synthesis in older men. *J Appl Physiol* 108:1199-1209, 2010
42. Fujita S, Dreyer HC, Drummond MJ, Glynn EL, Cadenas JG, Yoshizawa F, Volpi E, Rasmussen BB: Nutrient signalling in the regulation of human muscle protein synthesis. *J Physiol* 582:813-823, 2007
43. Fujita S, Glynn EL, Timmerman KL, Rasmussen BB, Volpi E: Supraphysiological hyperinsulinaemia is necessary to stimulate skeletal muscle protein anabolism in older adults: evidence of a true age-related insulin resistance of muscle protein metabolism. *Diabetologia* 52:1889-1898, 2009
44. Fujita S, Rasmussen BB, Bell JA, Cadenas JG, Volpi E: Basal muscle intracellular amino acid kinetics in women and men. *Am J Physiol Endocrinol Metab* 292:E77-E83, 2007
45. Fujita S, Rasmussen BB, Cadenas JG, Grady JJ, Volpi E: Effect of insulin on human skeletal muscle protein synthesis is modulated by insulin-induced changes in muscle blood flow and amino acid availability. *Am J Physiol Endocrinol Metab* 291:E745-E754, 2006
46. Fujita S, Rasmussen BB, Cadenas JG, Grady JJ, Volpi E: The effect of insulin on human skeletal muscle protein synthesis is modulated by insulin-induced changes in muscle blood flow and amino acid availability. *Am J Physiol Endocrinol Metab* 291:E745-E754, 2006
47. Gill TM, Allore H, Guo Z: The deleterious effects of bed rest among community-living older persons. *J Gerontol A Biol Sci Med Sci* 59:755-761, 2004
48. Glynn EL, Fry CS, Drummond MJ, Timmerman KL, Dhanani S, Volpi E, Rasmussen BB: Excess leucine intake enhances muscle anabolic signaling but not net protein anabolism in young men and women. *J Nutr* 140:1970-1976, 2010
49. Gougeon R, Morais JA, Chevalier S, Pereira S, Lamarche M, Marliss EB: Determinants of whole-body protein metabolism in subjects with and without type 2 diabetes. *Diabetes Care* 31:128-133, 2008
50. Gougeon R, Pencharz PB, Marliss EB: Effect of NIDDM on the kinetics of whole-body protein metabolism. *Diabetes* 43:318-328, 1994

51. Graham JE, Fisher SR, Berges IM, Kuo YF, Ostir GV: Walking speed threshold for classifying walking independence in hospitalized older adults. *Phys Ther* 90:1591-1597, 2010
52. Halvatsiotis P, Short KR, Bigelow M, Nair KS: Synthesis rate of muscle proteins, muscle functions, and amino acid kinetics in type 2 diabetes. *Diabetes* 51:2395-2404, 2002
53. Hardy SE, Kang Y, Studenski SA, Degenholtz HB: Ability to walk 1/4 mile predicts subsequent disability, mortality, and health care costs. *J Gen Intern Med* 26:130-135, 2011
54. Highstead RG, Tipton KD, Creson DL, Wolfe RR, Ferrando AA: Incidence of associated events during the performance of invasive procedures in healthy human volunteers. *J Appl Physiol* 98:1202-1206, 2005
55. Hundal HS, Taylor PM: Amino acid transceptors: gate keepers of nutrient exchange and regulators of nutrient signaling. *Am J Physiol Endocrinol Metab* 296:E603-E613, 2009
56. Katsanos CS, Kobayashi H, Sheffield-Moore M, Aarsland A, Wolfe RR: Aging is associated with diminished accretion of muscle proteins after the ingestion of a small bolus of essential amino acids. *Am J Clin Nutr* 82:1065-1073, 2005
57. Katsanos CS, Kobayashi H, Sheffield-Moore M, Aarsland A, Wolfe RR: A high proportion of leucine is required for optimal stimulation of the rate of muscle protein synthesis by essential amino acids in the elderly. *Am J Physiol Endocrinol Metab* 291:E381-E387, 2006
58. Kim TN, Park MS, Yang SJ, Yoo HJ, Kang HJ, Song W, Seo JA, Kim SG, Kim NH, Baik SH, Choi DS, Choi KM: Prevalence and determinant factors of sarcopenia in patients with type 2 diabetes: the Korean Sarcopenic Obesity Study (KSOS). *Diabetes Care* 33:1497-1499, 2010
59. Kimball SR, Jefferson LS: Regulation of global and specific mRNA translation by oral administration of branched-chain amino acids. *Biochem Biophys Res Commun* 313:423-427, 2004
60. Kortebein P, Ferrando A, Lombeida J, Wolfe R, Evans WJ: Effect of 10 days of bed rest on skeletal muscle in healthy older adults. *JAMA* 297:1772-1774, 2007
61. Ledgerwood AM, Saxe JM, Lucas CE: Venous access devices: a review. *Surgery Annual* 25 Pt 2:45-57, 1993
62. Lee CG, Boyko EJ, Barrett-Connor E, Milkovic I, Hoffman AR, Everson-Rose SA, Lewis CE, Cawthon PM, Strotmeyer ES, Orwoll ES: Insulin sensitizers may attenuate lean mass loss in older men with diabetes. *Diabetes Care* 34:2381-2386, 2011
63. Manders RJ, Koopman R, Beelen M, Gijssen AP, Wodzig WK, Saris WH, van Loon LJ: The muscle protein synthetic response to carbohydrate and protein ingestion is not impaired in men with longstanding type 2 diabetes. *J Nutr* 138:1079-1085, 2008
64. Markofski MM, Dickinson JM, Drummond MJ, Fry CS, Fujita S, Gundersmann DM, Glynn EL, Jennings K, Paddon-Jones D, Reidy PT, Sheffield-Moore M, Timmerman KL, Rasmussen BB, Volpi E: Effect of age on basal muscle protein synthesis and mTORC1 signaling in a large cohort of young and older men and women. *Exp Gerontol* 65:1-7, 2015
65. Mayhew DL, Kim JS, Cross JM, Ferrando AA, Bamman MM: Translational signaling responses preceding resistance training-mediated myofiber hypertrophy in young and old humans. *J Appl Physiol* (1985) 107:1655-1662, 2009

66. Miller S, Chinkes D, MacLean DA, Gore D, Wolfe RR: In vivo muscle amino acid transport involves two distinct processes. *American Journal of Physiology - Endocrinology & Metabolism* 287(1):E136-41, 2004
67. Moller-Loswick AC, Zachrisson H, Hyltander A, Korner U, Matthews DE, Lundholm K: Insulin selectively attenuates breakdown of nonmyofibrillar proteins in peripheral tissues of normal men. *Am J Physiol* 266:E645-E652, 1994
68. Newman AB, Kupelian V, Visser M, Simonsick EM, Goodpaster BH, Kritchevsky SB, Tylavsky FA, Rubin SM, Harris TB: Strength, but not muscle mass, is associated with mortality in the health, aging and body composition study cohort. *J Gerontol A Biol Sci Med Sci* 61:72-77, 2006
69. Okada S, Hiuge A, Makino H, Nagumo A, Takaki H, Konishi H, Goto Y, Yoshimasa Y, Miyamoto Y: Effect of exercise intervention on endothelial function and incidence of cardiovascular disease in patients with type 2 diabetes. *J Atheroscler Thromb* 17:828-833, 2010
70. Paddon-Jones D, Sheffield-Moore M, Urban RJ, Sanford AP, Aarsland A, Wolfe RR, Ferrando AA: Essential amino acid and carbohydrate supplementation ameliorates muscle protein loss in humans during 28 days bedrest. *J Clin Endocrinol Metab* 89:4351-4358, 2004
71. Paddon-Jones D, Sheffield-Moore M, Zhang XJ, Volpi E, Wolf SE, Aarsland A, Ferrando AA, Wolfe RR: Amino acid ingestion improves muscle protein synthesis in the young and elderly. *American Journal of Physiology - Endocrinology & Metabolism* 286(3):E321-8, 2004
72. Park SW, Goodpaster BH, Lee JS, Kuller LH, Boudreau R, de RN, Harris TB, Kritchevsky S, Tylavsky FA, Nevitt M, Cho YW, Newman AB: Excessive loss of skeletal muscle mass in older adults with type 2 diabetes. *Diabetes Care* 32:1993-1997, 2009
73. Park SW, Goodpaster BH, Strotmeyer ES, de RN, Harris TB, Schwartz AV, Tylavsky FA, Newman AB: Decreased muscle strength and quality in older adults with type 2 diabetes: the health, aging, and body composition study. *Diabetes* 55:1813-1818, 2006
74. Park SW, Goodpaster BH, Strotmeyer ES, Kuller LH, Broudeau R, Kammerer C, de RN, Harris TB, Schwartz AV, Tylavsky FA, Cho YW, Newman AB: Accelerated loss of skeletal muscle strength in older adults with type 2 diabetes: the health, aging, and body composition study. *Diabetes Care* 30:1507-1512, 2007
75. Rasmussen BB, Fujita S, Wolfe RR, Mittendorfer B, Roy M, Rowe VL, Volpi E: Insulin resistance of muscle protein metabolism in aging. *FASEB Journal* 20:768-769, 2006
76. Rogers FB, Cipolle MD, Velmahos G, Rozycki G, Luchette FA: Practice management guidelines for the prevention of venous thromboembolism in trauma patients: the EAST practice management guidelines work group. *J Trauma* 53:142-164, 2002
77. Sakowski C, Starc V, Smith SM, Schlegel TT: Sedentary long-duration head-down bed rest and ECG repolarization heterogeneity. *Aviat Space Environ Med* 82:416-423, 2011
78. Shah KB, Rao TL, Laughlin S, El-Etr AA: A review of pulmonary artery catheterization in 6,245 patients. *Anesthesiology* 61:271-275, 1984
79. Smith GI, Atherton P, Villareal DT, Frimel TN, Rankin D, Rennie MJ, Mittendorfer B: Differences in muscle protein synthesis and anabolic signaling in the postabsorptive state and in response to food in 65-80 year old men and women. *PLoS ONE* 3:e1875, 2008

80. Smith GI, Reeds DN, Hall AM, Chambers KT, Finck BN, Mittendorfer B: Sexually dimorphic effect of aging on skeletal muscle protein synthesis. *Biol Sex Differ* 3:11, 2012
81. Staten MA, Matthews DE, Bier DM: Leucine metabolism in type II diabetes mellitus. *Diabetes* 35:1249-1253, 1986
82. Studenski S, Perera S, Patel K, Rosano C, Faulkner K, Inzitari M, Brach J, Chandler J, Cawthon P, Connor EB, Nevitt M, Visser M, Kritchevsky S, Badinelli S, Harris T, Newman AB, Cauley J, Ferrucci L, Guralnik J: Gait speed and survival in older adults. *JAMA* 305:50-58, 2011
83. Symons TB, Sheffield-Moore M, Mamerow MM, Wolfe RR, Paddon-Jones D: The anabolic response to resistance exercise and a protein-rich meal is not diminished by age. *J Nutr Health Aging* 15:376-381, 2011
84. Symons TB, Sheffield-Moore M, Wolfe RR, Paddon-Jones D: A moderate serving of high-quality protein maximally stimulates skeletal muscle protein synthesis in young and elderly subjects. *J Am Diet Assoc* 109:1582-1586, 2009
85. Timmerman KL, Dhanani S, Glynn EL, Fry CS, Drummond MJ, Jennings K, Rasmussen BB, Volpi E: A moderate acute increase in physical activity enhances nutritive flow and the muscle protein anabolic response to mixed nutrient intake in older adults. *Am J Clin Nutr* 95:1403-1412, 2012
86. Timmerman KL, Lee JL, Dreyer HC, Dhanani S, Glynn EL, Fry CS, Drummond MJ, Sheffield-Moore M, Rasmussen BB, Volpi E: Insulin stimulates human skeletal muscle protein synthesis via an indirect mechanism involving endothelial-dependent vasodilation and mammalian target of rapamycin complex 1 signaling. *J Clin Endocrinol Metab* 95:3848-3857, 2010
87. Timmerman KL, Lee JL, Fujita S, Dhanani S, Dreyer HC, Fry CS, Drummond MJ, Sheffield-Moore M, Rasmussen BB, Volpi E: Pharmacological vasodilation improves insulin-stimulated muscle protein anabolism but not glucose utilization in older adults. *Diabetes* 59:2764-2771, 2010
88. Villareal DT, Smith GI, Shah K, Mittendorfer B: Effect of Weight Loss on the Rate of Muscle Protein Synthesis During Fasted and Fed Conditions in Obese Older Adults. *Obesity (Silver Spring)* 2011
89. Villareal DT, Smith GI, Sinacore DR, Shah K, Mittendorfer B: Regular multicomponent exercise increases physical fitness and muscle protein anabolism in frail, obese, older adults. *Obesity (Silver Spring)* 19:312-318, 2011
90. Volpato S, Bianchi L, Lauretani F, Lauretani F, Bandinelli S, Guralnik JM, Zuliani G, Ferrucci L: Role of Muscle Mass and Muscle Quality in the Association Between Diabetes and Gait Speed. *Diabetes Care* 2012
91. Volpato S, Maraldi C, Fellin R: Type 2 diabetes and risk for functional decline and disability in older persons. *Curr Diabetes Rev* 6:134-143, 2010
92. Volpi E, Campbell WW, Dwyer JT, Johnson MA, Jensen GL, Morley JE, Wolfe RR: Is the optimal level of protein intake for older adults greater than the recommended dietary allowance? *J Gerontol A Biol Sci Med Sci* 68:677-681, 2013
93. Volpi E, Ferrando AA, Yeckel CW, Tipton KD, Wolfe RR: Exogenous amino acids stimulate net muscle protein synthesis in the elderly. *J Clin Invest* 101:2000-2007, 1998

94. Volpi E, Kobayashi H, Mittendorfer B, Sheffield-Moore M, Wolfe RR: Essential amino acids are primarily responsible for the amino acid stimulation of muscle protein anabolism in healthy elderly adults. *Am J Clin Nutr* 78:250-258, 2003
95. Volpi E, Mittendorfer B, Rasmussen BB, Wolfe RR: The response of muscle protein anabolism to combined hyperaminoacidemia and glucose-induced hyperinsulinemia is impaired in the elderly. *J Clin Endocrinol Metab* 85:4481-4490, 2000
96. Volpi E, Mittendorfer B, Wolf SE, Wolfe RR: Oral amino acids stimulate muscle protein anabolism in the elderly despite higher first pass splanchnic extraction. *Am J Physiol Endocrinol Metab* 277:E513-E520, 1999
97. Walker DK, Drummond MJ, Dickinson JM, Borack MS, Jennings K, Volpi E, Rasmussen BB: Insulin increases mRNA abundance of the amino acid transporter SLC7A5/LAT1 via an mTORC1-dependent mechanism in skeletal muscle cells. *Physiological Reports* 2:e00238, 2014
98. Walker DK, Fry CS, Drummond MJ, Dickinson JM, Timmerman KL, Gundermann DM, Jennings K, Volpi E, Rasmussen BB: PAX7+ satellite cells in young and older adults following resistance exercise. *Muscle Nerve* 46:51-59, 2012
99. Welle S: *Human Protein Metabolism*. New York, NY, Springer, 1999
100. Welle S, Nair KS: Failure of glyburide and insulin treatment to decrease leucine flux in obese type II diabetic patients. *Int J Obes* 14:701-710, 1990
101. Wojtaszewski JF, Hansen BF, Gade, Kiens B, Markuns JF, Goodyear LJ, Richter EA: Insulin signaling and insulin sensitivity after exercise in human skeletal muscle. *Diabetes* 49:325-331, 2000
102. Yarasheski KE, Pak-Loduca J, Hosten DL, Obert KA, Brown, MB, Sinacore DR: Resistance exercise training increases mixed muscle protein synthesis rate in frail women and men ≥ 76 yr old. *Am J Physiol Endocrinol Metab* 277:E118-E125, 1999