

**Impact of DNA fragmentation in sperm on the pregnancy outcome after
intra-uterine insemination in a spontaneous cycle**

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Introduction

Infertility affects about 10% of all couples and is defined by a failure to achieve a clinical pregnancy within a year of regular unprotected sexual intercourse. (Boivin 2007, Zegers-Hochschild 2009).

Male infertility accounts for nearly 30% of diagnoses, whilst an estimated 40% of male factor infertility remains unexplained (de Kretser 1997) and most of male factor infertility cannot be treated adequately.

The current diagnosis of male infertility is based on semen criteria updated by the WHO in 2010. These guidelines would consider a man normal with reference values of greater than 15 million sperm, greater than 4% normal morphology, and 32% progressive motility. By doing so more men are considered fertile while there may be an unnoticed rise in the number of infertile men meantime being assigned to the ‘unexplained’ category. Furthermore , although the WHO classification suggests accuracy through a methodology harmonized across laboratories (Wang 2014), the relevance for the choice of treatment and the predictive value for an infertile couple embarking on medically assisted reproduction technologies (MAR) is questionable. (Hamilton 2015)

In other words, there is a strong clinical need to distinguish fertile from infertile men through new sperm function testing and to be able to select both the patient population who will benefit from MAR as well as the type of treatment.

Numerous studies utilizing different techniques for assessing sperm DNA fragmentation support the existence of a significant association between sperm DNA damage and pregnancy outcomes (Collins et al., 2008). Moreover, a significant number of subfertile men have abnormal DNA integrity despite normal semen parameters (Kodama et al., 1997; Spano et al., 2000; Zini et al., 2001). The level of DNA fragmentation correlates negatively with pregnancy and delivery in both natural and assisted conceptions (Evenson & Wixon 2006; Zhang et al. 2008; Zini 2011).

Our laboratory has set up a program where direct DNA fragmentation testing with terminal deoxyuridine nick end labeling (TUNEL) assay (Mitchell 2011) on fresh samples was established both on the total and vital fraction, before and after density gradient. The data results have been submitted and will be available soon (Punjabi 2017).

In this prospective cohort study we aim to investigate the role of sperm DNA fragmentation analysis in selecting the patient who will benefit from intra-uterine insemination (IUI) therapy since IUI is still considered the first step in MAR and is performed at a large scale in Belgium and worldwide.

The overall success rate of IUI remains controversial. On average, reported clinical pregnancy rates are only 5–13% per IUI cycle (Bensdorp et al., 2007; Goverde et al., 2000; Guzick et al., 1999; Nyboe Andersen et al., 2009; Steures et al., 2004, 2007; The ESHRE Capri Workshop Group, 2009; Tummon et al., 1997; Verhulst et al., 2006).

A Cochrane analysis pointed to the position of IUI in male subfertility treatment as being one of the most frequently used treatments (Veltman-Verhulst et al., 2016) although its efficacy in the setting of mild male/unexplained infertility is yet to be proven. The authors concluded that there was insufficient evidence of effectiveness to recommend or advise against IUI.

In addition, the literature pertaining IUI consists of a broad range of indications for therapy, ranging from but not limited to: unexplained fertility, mild male infertility and mild endometriosis. Respectively, each of these diagnoses also have slightly different definitions from one study to another, thus making a univocal recommendation concerning IUI very challenging.

Our goal would be to better select patients for IUI, widely considered less invasive and expensive therapy than others, as about 50% of the patients entering an IUI program will not become pregnant after 4-6 IUI cycles and thus in selected cases (Van Hoof et al., 2009) progression to more advanced fertility therapy could be beneficial.

Study Objectives

The objective is to test the hypothesis that DNA fragmentation testing can play a diagnostic and thus pivotal role in selecting the patient who will benefit from intra-uterine insemination (IUI) therapy leading to a higher clinical pregnancy and live birth rate (primary outcomes) compared to the standard semen criteria.

Additionally, DNA fragmentation will be measured both at the time of the diagnostic work-up as at the time of insemination with evaluation in the neat and washed specimen, both in the total and vital fraction. We aim to assess whether density gradient retrieves high quality motile sperm with little DNA damage and where DNA fragmentation testing best fits in the clinical patient pathway (secondary outcomes).

Study Endpoints

Primary outcomes:

- DNA fragmentation as a predictor of clinical pregnancy and live birth rate

Secondary outcomes:

- DNA fragmentation in the total and vital fraction before and after density gradient in the diagnostic sample (pre-IUI)
- DNA fragmentation in the total and vital fraction before and after density gradient in the therapeutic sample (IUI sample)
- DNA fragmentation in relation to the cumulative clinical pregnancy and live birth rate

Study Phase & Design

Prospective interventional study
Duration: 24 months

Source population: couples with unexplained, mild male infertility, mild endometriosis defined by revised American Society for Reproductive Medicine stage I and II

Single center, university setting, no commercial affiliation

Study Population

Inclusion criteria:

Couples seeking fertility treatment after at least 12 months of unprotected intercourse are eligible. All couples underwent basic fertility investigations which included semen analysis, evaluation of menstrual cycle, and tubal patency testing (Chlamydia antibody test, hysterosalpingography/HyfoSy or laparoscopy).

All participating couples should provide written informed consent

Inclusion criteria are

- Unexplained infertility
 - o We classify couples as having unexplained subfertility when the fertility investigations showed at least one patent fallopian tube, a normal ovulatory

- menstrual cycle (26-32), and a normal semen analysis (pre-wash total motile sperm count above 10 million).
- Mild male infertility defined as one or more abnormal semen parameters according to the WHO 2010 criteria on at least two examinations; diagnosis is determined by most abnormal result
- Women: age between 18 and 40 years, BMI between 18,5-30, presence of at least one patent tube on hysterosalpingography/Hysterosalpingo foam sonography (Hyfosalpingo) and/or laparoscopy, normal uterine cavity (ultrasound, hysterosalpingography or laparoscopy), regular menstrual cycle (24-38 days, FIGO recommendations 2011)
- Men: age between 18 and 65 years, BMI between 18,5-30, normal semen analysis or mild male subfertility
- Exclusion criteria: double sided tubal disease, severe endometriosis (classified as revised American Society for Reproductive Medicine stage III or IV), premature ovarian failure, and known endocrine disorders (such as Cushing's syndrome or adrenal hyperplasia), azoö- or necrozoöspermia

Statistics

- Sample size: 120 patients
A power analysis was performed to estimate the sample size needed for ROC curve analysis. Since there is scarce literature for sperm DNA fragmentation on an IUI sample both before and after gradient, we cannot rely on reference values to estimate the area under the curve (AUC) or distribution of cases versus controls (kappa). In function of a significance of 0,05 (type I error) and power of 0,8 (1-type II error), a kappa varying between 1:1 and 1:3 and AUC varying between 0,7-0,75 we estimate a minimum of 38 patients and a maximum of 130 patients should be included.
- Diagnostic parameters: sensitivity, specificity, area under receiver operator curve, predictive value, likelihood ratio, odds ratio
- Null hypothesis:
 - 1) There is no correlation between DNA fragmentation and clinical pregnancy/live birth after IUI
 - 2) Density gradient isn't capable of selecting motile sperm with low DNA fragmentation for insemination

Clinical protocol

- IUI in a spontaneous cycle
- Basal ultrasound and hormonal analysis on cycle day 2 or 3 (estradiol (E2), progesterone (P), luteinizing hormone (LH) as well as follicle stimulating hormone (FSH))
- Cycle monitoring from D10-12 with ultrasound and/or hormonal analysis (E2, P, LH)
- 250 microgram hCG or 5000 IU is administered to induce ovulation when a maximum of 2 dominant follicles are present. Either Pregnyl, Choragon, Ovitrelle will be used
- IUI is planned between 32-40 hours after HCG administration or \pm 24-26 hours after detection of spontaneous LH surge
- Both ultrasound and hormonal analysis (E2, P, LH) are done on the day of planning of IUI (day of hCG administration or day of LH surge)
- Sperm preparation is performed according to local validated procedures in accordance with the 2010 WHO criteria. DNA fragmentation analysis by TUNEL assay will be carried out both before and after density gradient.
- Patients are advised to rest in the supine position during 15 minutes immediately after IUI according to evidence from a RCT (Custers et al. 2009)
- Detection of pregnancy
 - β HCG testing is performed 15-20 days after insemination
 - A second β HCG testing is performed 7 days later to confirm a positive evolution of pregnancy
 - In case of a positive evolution of β HCG an ultrasound is performed around 5-6 weeks after IUI
 - Follow-up of pregnancy until birth through staff, telephone or e-mail contact until birth

- Items to be registered
 - Number of eligible patients (patients who fulfilled inclusion criteria and to whom the study was proposed)
 - Number of drop outs between inclusion and end of treatment
 - Number of spontaneous pregnancies between inclusion and end of treatment
 - Number of live births between inclusion and end of treatment

 - Hunault score and modified Hunault-score according to Bendsorp et al.
 - All Belrap items
 - Semen analysis methodology checklist according to Björndahl et al. (Appendix)
 - Patient population characteristics:
 - Infertility: duration, primary versus secondary, female/male/mixed/unexplained
 - Cause of female infertility: Ovulatory disorder, Anatomical infertility, Endometriosis (revised American Society for Reproductive Medicine classification) or Adenomyosis
 - Possible cause of male infertility: Varicocele, Cryptorchidism, Male accessory gland infections, Germ cell malignancies and testicular microcalcifications, Disorders of ejaculation, Idiopathic. Primary spermatid failure, genetic disorders of infertility, obstructive azoospermia, hypogonadism were not reported since azoospermia is considered an exclusion criterium.

- Data analysis and Publication policy
Data analysis and preparation for publication will be done at the University Hospital of Antwerp / University of Antwerp.

References

Bendsorp AJ, Cohlen BJ, Heineman MJ, Vandekerckhove P. Intra-uterine insemination for male subfertility. Cochrane Database of Systematic Reviews 2007, Issue 3. Art. No.: CD000360.

Boivin, J, Bunting, L, Collins, JA and Nygren KG. International Estimates of Infertility Prevalence and Treatment-Seeking: Potential Need and Demand for Infertility Medical Care. *Hum Reprod.* 2007; 22: 1506-1512.

Collins JA, Barnhart KT, Schlegel PN.. Do sperm DNA integrity tests predict pregnancy with in vitro fertilization? *Fertil Steril.* 2008; 89: pp. 823-831

Custers Inge M, Flierman Paul A, Maas Pettie, Cox Tessa, Van Dessel Thierry J H M, Gerards Mariette H et al. Immobilisation versus immediate mobilisation after intrauterine insemination: randomised controlled trial. *BMJ* 2009; 339 :b4080

de Kretser DM. Male infertility. *Lancet.* 1997; 15: 349(9054):787-790.

Evenson D, Wixon R. Meta-analysis of sperm DNA fragmentation using the sperm chromatin structure assay. *Reprod Biomed Online.* 2006;12(4):466-472.

Goverde AJ, McDonnell J, Vermeiden JP, Schats R, Rutten FF, Schoemaker J. Intrauterine insemination or in-vitro fertilisation in idiopathic subfertility and male subfertility: a randomised trial and cost-effectiveness analysis. *Lancet* 2000;355:13–18.

Guzick DS, Carson SA, Coutifaris C, Overstreet JW, Factor-Litvak P, Steinkampf MP, Hill JA, Mastroianni L, Buster JE, Nakajima ST et al. Efficacy of superovulation and intrauterine insemination in the treatment of infertility. *N Engl J Med* 1999;340:177–183

Hamilton JA, Cissen M, Brandes M, Smeenk JM, de Bruin JP, Kremer JA, Nelen WL, Hamilton CJ. (2015) Total motile sperm count: a better indicator for the severity of male factor infertility than the WHO sperm classification system. *Hum Reprod.* 2015;30(5):1110-21

Kodama H, Yamaguchi R, Fukuda J, Kasai H and Tanaka T . Increased oxidative deoxyribonucleic acid damage in the spermatozoa of infertile male patients. *Fertil Steril.* 2007; 68(3):519–524.

Mitchell LA, De Iuliis GN, Aitken RJ. *Int J Androl.* 2011;34(1):2-13. The TUNEL assay consistently underestimates DNA damage in human spermatozoa and is influenced by DNA compaction and cell vitality: development of an improved methodology.

Spano M, Bonde J, Hjollund HI et al. Sperm chromatin damage impairs human fertility. *Fertil Steril.* 2000; 73: 43–50.

Steures P, van der Steeg JW, Mol BW, Eijkemans MJC, van der Veen F, Habbema JDF, Hompes PGA, Bossuyt PMM, Verhoeve HR, van Kasteren YM et al. Prediction of an ongoing pregnancy after intrauterine insemination. *Fertil Steril* 2004;82:45–51.

The ESHRE Capri Workshop Group, *Hum Reprod Update* 2009 15:265-277

Tummon IS, Asher LJ, Martin JS, Tulandi T. Randomized controlled trial of superovulation and insemination for infertility associated with minimal or mild endometriosis. *Fertil Steril* 1997;68(1):8-12

Van Hoof K, Elseviers M, De Pauw I, Eestermans W, De Neubourg D. Cumulative ongoing pregnancy rates after IUI and investigation of factors affecting drop-out. *ESHRE campus symposium: Artificial insemination: an update.* Genk. 13-15/12/2009

Veltman-Verhulst SM, Hughes E, Ayeleke RO, Cohlen BJ. Intra-uterineinsemination for unexplained subfertility. *Cochrane Database Syst Rev* 2016;9:CD001838.

Verhulst SM, Cohlen BJ, Hughes E, te Velde E, Heineman MJ. Intra-uterine insemination for unexplained subfertility. *Cochrane Database Syst Rev* 2006;Art No.: CD001838.

Wang C, Swerdloff RS. Limitations of Semen Analysis as a Test of Male Fertility and Anticipated Needs from Newer Tests. *Fertil Steril.* 2014;102(6):1502-1507.

World Health Organization. WHO laboratory manual for the examination of human semen and sperm-cervical mucus interaction. 5th ed. World Health Organization, Geneva; 2010.

Zegers-Hochschild, F. et al. International Committee for Monitoring Assisted Reproductive Technology (ICMART) and the World Health Organization (WHO) revised glossary of ART terminology. *Fertil Steril* 2009;92(5): 1520 - 1524

Zhang Y, Wang H, Wang L, Zhou Z, Sha J, Mao Y, et al. The clinical significance of sperm DNA damage detection combined with routine semen testing in assisted reproduction. *Mol Med Rep.* 2008;1:617–624.

Zini, A., Bielecki, R., Phang, D. and Zenzes, M.T. Correlations between two markers of sperm DNA integrity, DNA denaturation and DNA fragmentation, in fertile and infertile men. *Fertil. Steril* 2001; 75(4): 674-677.

Zini A. Are sperm chromatin and DNA defects relevant in the clinic? *Syst Biol Reprod Med.* 2011;57(1-2):78-85.