

Title: Novel Blood Test to Predict Safe Foods for Infants and Toddlers With Food Protein-induced Enterocolitis Syndrome (FPIES)

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1 **A NOVEL BLOOD TEST TO PREDICT SAFE (NON-TRIGGER) FOODS FOR INFANTS AND TODDLERS**
2 **WITH FOOD PROTEIN-INDUCED ENTEROCOLITIS SYNDROME (FPIES).**

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14 **VERSION 2; ***.**
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17 **I. PROTECTION OF HUMAN SUBJECTS**
18

19 **1. Risks to Human Subjects**
20

21 **A. Human Subjects Involvement, Characteristics, and Design**
22

23 Describe and justify proposed human involvement
24

25 The **objective** of this project is to validate a blood test that can distinguish safe from trigger foods for
26 protein-induced enterocolitis syndrome (FPIES).
27

28 **Background:** FPIES is an allergic disease that leads to repetitive profuse vomiting in response to solid food.
29 In severe cases, when patients fail to thrive, FPIES requires nasogastric tube installment or total parenteral
30 nutrition (TPN). The major challenge in managing FPIES is the identification of safe (non-trigger) foods in a
31 timely manner, while avoiding repetitive allergic reactions and recovery periods. This study proposes a
32 solution to the problem by developing a new blood assay that screens a large number of foods (more than
33 20) in a culture plate. Its purpose is to rapidly predict what the safe foods are. The outcome will overcome
34 the time-lapse required to identify such safe foods, which could normally extend past a year or longer, by
35 which time the child may have failed to thrive or developed food aversions due to association with negative
36 experiences.
37

38 FPIES leads to repetitive hospital and ER visits with 2,064 encounters occurring at the University of
39 Michigan Health System (UMHS) in the past year (from 11/01/17 until 11/01/18; data obtained using
40 Datadirect for cohort discovery results; FPIES ICD10 K52.2). The latter number of encounters only
41 comprises newly diagnosed patients.
42

43 In this study, FPIES patients will be recruited from the UMHS patient cohort. From 11/01/17 – 11/01/18, this
44 cohort comprised 223 newly diagnosed patients at UMHS. Two sub-groups will be identified as follows: *i)*
45 children exhibiting FPIES reactions to 2-3 foods, *ii)* children exhibiting FPIES reactions to 4 or more foods.
46 The reason for separating patients into these two groups is due the existence of two separate groups [one
47 with 2-3 food triggers, and another with more than 4 food triggers] as previously described by the UMHS
48 Allergists who care for FPIES patients (Volertas S., Hudson E., McMorris G., and G. Sanders. *Annals of*
49 *Allergy, Asthma & Immunology* 121:S119 2018).

50
51 The plan is to enroll up to 20 patients with FPIES: up to 10 exhibiting reactions to 2-3 foods, and up to 10
52 exhibiting FPIES reactions to 4 or more foods. Recruitment will take place at the University of Michigan
53 Allergy Clinics, and University of Michigan Pediatric outpatient clinics.
54

55 **Hypotheses:**

- 56 1. Our blood test that we developed will distinguish safe from trigger foods for FPIES patients.
57 2. The mechanism of FPIES involves innate immunity and the identified heatmap pattern for trigger foods
58 will remain after foods are tolerated.

59 **Specific aims:**

- 60 1. Assess the assay's precision by its predictive accuracy in distinguishing safe foods from trigger foods.
61 2. Evaluate the reliability of the test when using a panel of standardized, frozen foods vs freshly prepared
62 foods
63 3. Demonstrate persistence of the immune reaction to trigger foods by comparing results of the test after
64 trigger foods are tolerated to the initial results

65 **Exploratory aim:**

- 66 1. Gain further insight into the pathophysiology of FPIES through single cell RNASeq

67 **Statistical Design:** A random-effects logit model will be used to model the binary outcome (safe or trigger
68 food) as a function of the 9 biomarker measurements in the assay. The random effect in the logit model will
69 take into consideration of the correlated data measured within the same subject. A cluster Receiver
70 operating characteristic (ROC) curve analysis will be used to assess the precision of the assay. Specifically,
71 we will compute the area under the cluster ROC curve (AUC), along with a 95% confidence interval (CI).
72 The assay is predictive if the lower limit of the 95% CI is above 0.5, which is the null value indicating no
73 predictive ability.

74 **Brief description of the proposed study**

75 FPIES is a severe food allergy to a wide variety of foods in babies and toddlers. Usually, the symptoms
76 resolve between the ages of 3 to 5 years old. However, the children are at risk of failing to thrive, and
77 requiring a nasogastric tube or TPN. The objective of the study is to validate a newly developed blood assay
78 that will rapidly predict safe (non-trigger) foods for the affected children. In our preliminary data (Case Study
79 HUM00155216), the blood assay safely predicted more than 10 consecutive safe foods without triggering
80 an allergic response. Prior to the assay, allergic reactions and ER visits were repeatedly occurring.

81 **Subject Recruitment:** Subjects will be recruited through the University of Michigan Allergy Clinics, and
82 University of Michigan Pediatric outpatient clinics:

- 83 1. Patients may be referred by the clinician caring for the patient.
- 84 2. Pre-screening by reviewing FPIES visits in the past 2 years will be performed. Data Direct, EMERSE
85 and billing codes may be used to identify potential subjects. The patient's primary allergist or
86 pediatrician will be informed about study team's contact with the identified patients.
 - 87 o Patients will then be contacted by email and/or telephone by the clinical members of the
88 study team to explain the study and determine whether they are interested in participating.
89 Opt out information will be provided in the email and by phone contact.
- 90 3. Patients may self-refer:
 - 91 o The study will be listed in UMHealthResearch.org
 - 92 o Flyers describing the study, with study team contact information, will be posted in the
93 participating clinics.

94 **Inclusion Criteria:**

- 95 1. Patients aged 1 month – 7 years old with a physician diagnosis of FPIES:
 - 96 a. Group 1: Documented reactions to 2-3 trigger foods with recurrent delayed vomiting.
 - 97 b. Group 2: Documented reactions to 4 or more trigger foods with recurrent delayed vomiting.

98 **Exclusion Criteria:**

- 99 1. Patients without a physician diagnosis of FPIES.
- 100 2. Patients who are currently on medications that suppress the immune system.

- 109 3. Patients who do not have at least 2 trigger foods identified.
110 4. Patients who have a history of an organic GI disease (e.g., inflammatory bowel disease, celiac
111 disease, biliary disorders, bowel resection), cardiac, pulmonary, neurologic, renal, endocrine, or
112 gynecological pathology
113 5. Lack of parental or guardian informed consent.

114
115 Study Design

- 118 1. Potential participants will be contacted by email and / or telephone after pre-screening of physician-
119 diagnosed FPIES patients who have visited the UM allergy clinics within the past three years, or who
120 are currently in contact with the UM allergy clinic nutritionist. An option for the patients to opt out will
121 be provided. The patient's primary allergist or pediatrician will be informed about study team's
122 contact with the identified patients. Patients may be referred by their primary allergist during a clinic
123 visit.
124 2. If a parent/guardian expresses interest in the study, they will be sent/given a copy of the informed
125 consent to review.
126 3. An appointment will be scheduled with a study team member at the UM allergy clinics.
127 4. The protocol will be discussed face-to-face with parent/guardian with an opportunity to ask questions
128 and review the informed consent document. After written informed consent is signed by one
129 parent/guardian, study procedures can begin. Parent/guardian will be given a copy of the signed
130 informed consent and it will be imaged into the chart.
131 5. Parent/Guardian will be asked to fill out Questionnaire One at the time of entry into the study.
132 6. Parent/Guardian will be instructed regarding scheduling the blood draw for analysis:
133 a. Only known "safe foods" (foods that do not induce repetitive vomiting) are to be consumed
134 for one week before blood draw.
135 b. Blood draw will be performed at the MLabs Blood Draw Stations at Michigan Medicine
136 outpatient clinics.
137 7. Patient will have 6 ml of blood drawn for analysis:
138 a. 5.5 ml of blood for testing of potential food reactions
139 b. 0.5 ml of blood for RNA/DNA analysis, which will be stored.
140 8. After blood draw, diet will be managed per the primary clinician and nutritionist.
141 a. Parents will be given Questionnaire Two to fill out detailed information about all foods
142 introduced until the results of the assay are received.
143 b. During that time, food introduction will continue as directed by primary allergist and/or
144 nutritionist
145 9. Blood will be analyzed by the PI in a research lab at the University of Michigan to determine
146 differences in the PI's assay between potentially tolerated and not tolerated foods (see description of
147 laboratory test lines 216-232)
148 10. After identification of potentially "safe foods" to be introduced in the future, the information will be
149 given to the clinical study team and child's clinician, who will contact parents by phone or email
150 (parental preference).
151 a. Parents will be asked to detail current nutritional status, including foods tolerated or not
152 tolerated since the initial blood draw. Results will be recorded by the study team and the
153 parents will be asked to return Questionnaire Two.
154 b. After discussion within the clinical team (including clinical research team and child's
155 clinicians), parents will be instructed to introduce new foods identified as potentially safe into
156 the diet, one at a time for seven days.
157 c. Results of these trials will be recorded on Questionnaire Three and returned to the study
158 team 3 month intervals.

159 11. A second blood draw will be performed, at least 2 weeks after the initial blood draw. 6 cc will be
160 drawn:

- 161 a. To explore the consistency of a standard panel of foods vs individually prepared foods
- 162 b. To perform additional analyses including single cell RNASeq (scRNA-seq) on WBCs
- 163 following treatment with foods.

164 12. Subjects will be followed at 3 month intervals with review of tolerated foods, number and types of
165 known triggers assessed. When a subject is tolerating two or more triggers foods for 3 or more
166 months:

- 167 a. Questionnaire Four will be given to fill out and return
- 168 b. Subject will have 6 mL drawn for analysis
 - 169 i. 5.5 mL to rerun the initial assays
 - 170 ii. 0.5 mL for RNASeq on WBCs following treatment with foods.

171 Study Grid

	Baseline visit - Day 0	Lab Day 0- 14	Lab >2 weeks after initial blood draw	At home 1-3 months	3 up to 36 months	At home every 3 months up to 36 months	> 2 trigger foods tolerated
Consent	X						
Questionnaire 1	X						
Blood Draw		X	x				X
Phone/email/in-person to inform of potentially tolerated foods.				X			
Questionnaire 2				X			
Testing Foods Found as tolerable					X		
Questionnaire 3						X	
Questionnaire 4						X	X

172
173 B. Sources of Materials

174 The research material obtained from the human subjects

175 1. 6ml of blood in a K2 EDTA (lavender top) blood tube withdrawn by specialized phlebotomists in
176 the blood draw unit (0.5 ml of this will be used to isolate DNA for potential future sequencing under a
177 separate IRB). White blood cells (WBCs) will be isolated from 5.5 ml of blood using ACK buffer lysis of red
178 blood cells, within 1 hour of blood collection, as performed previously for the Case Study (HUM00155216).
179 Blood will be kept on ice in the K2 EDTA tubes during the waiting period (< 1 hour). WBCs will be plated in
180 24 well plates, and exposed to different food homogenates for 3 hours. Total RNA will be extracted, and RT-
181 qPCR for our gene panel will be used to generate the heatmap.

182 A second blood draw may be requested from a limited number of patients, at least 2-4 weeks after
183 the initial blood draw, for scRNA-seq. In this case, the blood will be prepared in the same manner as for the
184 initial draw above, but instead of extracting RNA, samples will be submitted to the UofM Advanced
185 Genomics Core for them to perform scRNA-seq on the samples. The data will be analyzed using the cloupe
186 software by 10x Genomics.

187 A third blood draw may occur (or second blood draw if scRNA were not performed for that patient)
188 after the patient recovers from several triggers. The patient will be screened at 3 month intervals, so the
189 third (or second if no scRNA-seq) will be performed for this purpose at least 3 months after the initial blood
190 draw. In this case, the cells will be prepared in the same manner as the initial blood draw, but treated with
191

193 only a limited number of safe, trigger foods, and outgrown trigger foods.

194 2. At the termination of the protocol samples and data will cease to be accessed, or otherwise
195 accessed again after IRB approval transferred to another existing protocol. There are no plans to destroy
196 records or samples.

197 **Protection of Information:**

198 All data will be kept on REDCap, a password-protected, HIPAA compliant, web-based application
199 developed by Vanderbilt University to capture data for clinical research and create databases and projects.
200 The databases use instruments such as surveys and forms as research capture tools. Projects are self-
201 sufficient and secure databases that can be used for normal data entry or for surveys across multiple
202 distinct time points. Only study team members will have access to this data.

- 203 1. A master list that includes identifiable data will be kept in a REDCap "Study Housekeeping"
204 database that includes subject number, patient name, parent names, MRN, contact information
205 including address, email, phone number, documentation of blood draw, samples saved and filling
206 out of questionnaires.
- 207 2. All data variables will be kept in a separate "Research Data" REDCap database. Subjects will be
208 identified only by subject number in this database. Data will include:
- 209 a. Results of blood tests
210 b. Questionnaire answers
211 c. Age
212 d. Number of food reactions
213 e. Identified trigger foods
214 f. Identified tolerated foods.
- 215 3. Data downloaded for analysis will have only subject number included.
- 216 4. Stored RNA, DNA or cDNA samples will contain only the subject number without personal
217 identifiers.
- 218 5. Parents/guardians may request that stored blood samples be destroyed at the end of the study.

219 **C. Potential Risks**

220 **Describe potential risks**

221 The risk in this study arises due to incorrect predictions by the test. The current test being validated is not
222 an FDA-approved or cleared test. However, current standard of care recommends which foods to trial in a
223 random fashion, and incorrect predictions frequently occur leading to reactions. Hence this study will not
224 pose an added risk to the current standard of care. The indication of what to trial will be communicated to
225 the patients by the allergist and nutritionist as currently occurs in the standard of care. However, the only
226 difference is that the allergist and nutritionist will in this study use this assay for indication regarding what
227 foods to trial. If a food is falsely identified as safe by the assay but triggers a reaction, then these reactions
228 will be managed by the nutritionist and allergist as per current standard of care for failed trials.

229 Blood draw from infants and toddlers is a safe procedure with minimal risks. Trained and experienced
230 phlebotomists at UMHS perform blood draws on infants and toddlers routinely. The procedure may cause
231 anxiety for the infant/toddler. The test can result in a small bruise or mild soreness for the child at the site of
232 blood draw. The bruise can last for a few days.

233 A risk of breach of confidentiality always exists in all studies.

234 **Describe alternate treatments and procedures**

235 The parent/child can proceed with introduction of new foods per guidance of the nutritionist. This is based
236 on historically better tolerated foods in the general FPIES population and may or may not apply to the
237 patient.

245 **2. Adequacy of protection against risks**

246 **A. Recruitment and Informed Consent**

249 Plan for recruitment

250 All FPIES patients will be recruited from the University of Michigan Allergy Clinics, and University of
251 Michigan Pediatric outpatient clinics. The patients will be identified by pre-screening of the UM FPIES
252 patient cohort. Pre-screening will be performed by members of the Division of Allergy and Immunology at
253 Michigan Medicine. Patients will not be contacted by the study team until pre-approval is obtained from the
254 primary allergist or pediatric clinician. Suitable FPIES patients who fulfill the inclusion and exclusion criteria
255 will be contacted by email and / or phone. An option to opt out will be provided. If patients are interested in
256 participating, a meeting with the study team will be scheduled at the UM allergy clinics. The patients will be
257 presented with the consent form by the study team member to consider consenting for the study and the
258 blood draw procedure. Patients will be told that their decision whether to participate in the study will not
259 affect their clinical care.

261 Describe the circumstances of consent

262 A study coordinator or clinical study team member familiar with the study protocol will obtain informed
263 consent for all participants. Consent/assent will be obtained prior to any study measures/questionnaires.
264 Each participant will receive a verbal and written explanation of the purposes, procedures, risks, and
265 potential benefits of the study in language appropriate for the individual.

266 **B. Protections Against Risk**

268 Planned procedures for protecting against risk

- 271 1. The risk of a false result will be managed by the allergist and nutritionist as per current standard of care in
272 managing reactions to failed trials.
- 274 2. The risk from blood draw is minimal and routinely performed by the phlebotomists at UMHS.
- 276 3. All data variables will be kept in a separate "Research Data" REDCap database. Subjects will be
277 identified only by subject number in this database.
- 279 4. A separate "Housekeeping" REDCap database will be used to link the patient identifiable information and
280 the study number.

282 **3. Potential benefit of the proposed research to the research subjects and others**

284 Potential benefits

285 There may be direct benefit to the patient by the potential of identifying tolerated foods, thus avoiding the
286 adverse reactions of frequent emesis and potential dehydration, as well as the trauma parents encounter
287 when giving their infant a food that causes such reactions. There may be benefit to the larger community of
288 FPIES patients if this protocol indicates the new assay can identify tolerated vs non-tolerated foods.

290 Discuss why risks to subjects are reasonable in relation to benefits

291 Patients already encounter false predictions and failed trials with allergic reactions regularly. The indication
292 from the assay is expected to predict safe foods at a higher rate than the guesswork utilized in the current
293 standard of care.

295 **4. Importance of the knowledge to be gained**

297 Discuss importance of the knowledge to be gained

298 Validation of this test has the potential to modify patient care strategy for patients with FPIES. As such,
299 random guessing of safe foods would be replaced by a scientific "guide" to what might be hypoallergenic to
300 the body's immune cells using this test.

301
302 Discuss why the risks are reasonable in relation to the importance of the knowledge to be gained
303 Currently, trialing food is random. Obtaining a guide for what might be a safe food, by utilizing our proposed
304 assay, would be invaluable. Incorrect predictions cannot pose more risk than random selection of foods to
305 trial. Hence, there is only benefit to be gained if the test is predictive or partially predictive.
306

307 **II. INCLUSION OF WOMEN AND MINORITIES** 308

309 *Inclusion of Women*

310 This study will include children of both genders.
311

312 *Inclusion of Minorities*

313 The disease is overwhelmingly predominant in White Caucasian children (Based on personal communication with
314 allergists caring for FPIES patients at UofM clinics, and the following published abstract: R. Tarrant and A. and Byrne. Clinical
315 Presentation and Food Allergens Associated with Food Protein-Induced Enterocolitis Syndrome – a frequently misdiagnosed rare
316 form of gastrointestinal food hypersensitivity. *European Society of Pediatric Gastroenterology, Hepatology and Nutrition* 62(1); DOI:
317 10.13140/RG.2.1.1621.2087). Therefore, for the purpose of feasibility of meeting the required proposed patient
318 numbers for statistical analyses, we have chosen a majority of White Caucasian subjects for this initial
319 study. If patients from other ethnicities – who do not usually have FPIES enroll – they will not be excluded
320 from the study.
321

322 *Inclusion of Children*

323 Children with FPIES will be included in this study.
324

325 **III. PLANNED ENROLLMENT TABLE**

Total Planned
Enrollment: 20

TARGETED/PLANNED ENROLLMENT: Number of Subjects			
Ethnic Category	Females	Males	Total
Hispanic or Latino	1	1	2
Not Hispanic or Latino	9	9	18
Ethnic Category: Total of All Subjects *	10	10	20
Racial Categories			
American Indian/Alaska Native			
Asian	1	1	2
Native Hawaiian or Other Pacific Islander			
Black or African American	1	1	2
White	8	8	16
Racial Categories: Total of All Subjects *	10	10	20