iNUDGE: INtegration of liqUiD biopsy based next generation Gene sEquencing in newly diagnosed NSCLC – A stepped wedge cluster randomized clinical trial

Principal Investigator	Charu Aggarwal, MD, MPH 10-137, South Pavilion
	3400 Civic Center Boulevard
	Philadelphia, PA 19104 Tel: 215-662-6318 Fax: 215-349-5326
Co-investigators	<u>Penn Medicine</u> Christopher D' Avella, MD E. Paul Wileyto, PhD Katharine Rendle, PhD, MPH Melina Marmarelis, MD, MSCE Ramy Sedhom, MD Samuel Kerr, MD Shayma Kazmi, MD
Funding Agency: IRB Protocol Number: UPCC Protocol Number: Version Date:	Eli Lilly 27522 852795 4/8/2025

Table of Contents

1.	BACKGROUND AND RATIONALE	3
2.	OBJECTIVES	4
2.1	Primary Objectives	4
2.2	Primary Outcomes	4
2.3	Secondary Outcomes	4
3. STI	UDY POPULATION	4
3.1	Target Population	4
3.2	Inclusion Criteria	5
3.3	Exclusion Criteria	5
3.4	Vulnerable Populations	5
4.	STUDY DESIGN	5
4.1	Preliminary Studies	5
4.2	Overview	5
4.3	Study Duration and Timeline	9
4.4	Study Setting	10
5.	STUDY PROCEDURES	10
5.1	Recruitment and Retention	10
5.2	Informed Consent	11
5.3	Sources of Materials	12
6.	STATISTICAL DESIGN AND POWER	12
6.1	Sample Size	12
6.2	Analysis Plan	13
7.	RESOURCES NECESSARY FOR HUMAN RESEARCH PROTECTION	14
8.	STUDY TEAM	15
9.	REFERENCES	16
10.	APPENDICES	18

1. BACKGROUND AND RATIONALE

The development of targeted therapies has changed the treatment paradigm for non-small cell lung cancer (NSCLC). With the growing number of FDA approved targeted therapies, current NCCN quidelines recommend comprehensive molecular genotyping, defined as detection of mutations in seven genes (EGFR, ALK, BRAF, ROS1, MET, RET, and NTRK) prior to first line (1L) therapy for all newly diagnosed patients with metastatic non-squamous (mNSg) NSCLC to enable the delivery of personalized therapy.^{1, 2} Furthermore, the emergence of immunecheckpoint inhibitors has amplified the importance of molecular genotyping in the care of these patients because patients with actionable genomic alterations rarely respond to immunotherapy. even in the presence of high PD-L1 expression and should be preferentially treated with targeted therapy.³ In addition, there is a growing body of evidence that introduction of targeted tyrosine kinase inhibitors after immunotherapy may be associated with higher rates of immune related adverse events, even after discontinuation of immunotherapy.⁴ Additionally, in previous studies, amongst patients with a mutation in a NCCN-listed gene, exposure to targeted therapy has been shown to be associated with improved overall survival.⁵ Given these considerations. upfront tumor genotyping is now considered an essential step in guiding treatment decisions for all patients with mNSq NSCLC, prior to 1L therapy.

Despite the critical importance of molecular testing in patients with advanced NSCLC, numerous barriers impede timely completion of testing prior to initiation of 1L systemic therapy.⁶⁻⁸ Common issues include insufficient tissue for testing, lack of infrastructure for obtaining and sending biopsy samples for testing, and unacceptably long turnaround times for results.⁹ These issues have created a critical need for additional convenient, and minimally invasive options for tumor genotyping.^{10, 11} We and others have previously demonstrated that the incorporation of concurrent plasma based next-generation gene sequencing (NGS), ordered at the same time as tissue NGS, improves detection of clinically actionable mutations in patients with advanced NSCLC.¹¹

At our institution, we piloted a behavioral economics (BE) informed "nudge" strategy to guide physicians' clinical practice to include concurrent use of plasma and tissue-based NGS at initial diagnosis. This real-world cohort study was conducted at the Abramson Cancer Center and 2 community sites within UPHS. Across the 3 practice sites, a provider team-focused Electronic Health Record (EHR) -based "nudge intervention" was designed to order plasma-based NGS at the time of new patient consultation. Eligible patients for the nudge were identified using an EHR based checklist, that included 3 criteria: i. new diagnosis, ii. treatment naïve, iii. mNSg NSCLC. Results from the intervention period (4/2021-12/2021) were compared to baseline data from similar patients treated at our institution between 01/2019 and 03/2021. Of the 526 patients with mNSq NSCLC that were included in the analysis: 381 were included in the pre-intervention cohort and 145 in the post-intervention cohort. After implementation of the EHR-based nudge, we observed that a higher proportion of patients underwent concurrent tissue + plasma testing in the post intervention cohort compared to pre-intervention 90.3% (131/145) vs. 68.8% (262/381), p<0.00001. Additionally, by virtue of having robust tissue + plasma testing performed, there were improved rates of comprehensive molecular genotyping in the postintervention cohort compared to pre-intervention, 98.6% (143/145) vs. 87.1% (332/381), p=0.00007. A greater proportion of patients had comprehensive genotyping available prior to 1st-line therapy in the post-intervention vs. pre-intervention cohort (86.2% vs. 76.3%, p=0.013).

These findings demonstrated that behavioral, EHR-based nudges are feasible and can promote guideline concordant diagnostic testing at both community and academic sites. The overarching goal of this current trial is to expand the application of the BE informed nudges, which includes a Best Practice Advisory (BPA) and Electronic Decision Support Tool (e-CDS) approach, which

has been operationalized within Epic, the EHR used at UPHS, to six satellite hospitals. Our central hypothesis is that this approach will dramatically increase adoption of comprehensive molecular testing and enhance the delivery of molecularly informed first-line therapy in patients with newly diagnosed metastatic non-squamous NSCLC. Molecular testing will be defined as i) comprehensive: *EGFR, ALK, BRAF, ROS1, MET, RET, KRAS, Her2* and *NTRK* testing, ii) incomplete: <6 genes tested, and iii) no testing performed. Clinically actionable mutations will be defined as an alteration in one of the seven genes on the comprehensive gene list with an FDA approved targeted therapy in the 1L setting, plus *KRAS* G12C, *EGFR* exon 20 insertion, and *ErbB2* mutations. Molecularly informed first line therapy will be defined as one that is informed by results of NGS, obtained by plasma, tissue or both.

2. OBJECTIVES

2.1 Primary Objectives

<u>Objective 1:</u> In a stepped wedge cluster randomized trial of patients with newly diagnosed metastatic NSCLC, test the effectiveness multicomponent BE informed EHR nudge intervention to increase timely receipt of comprehensive molecular test results before 1L therapy by integrating concurrent tissue and plasma-based molecular testing into the workup of newly diagnosed patients. Molecular testing will be defined as i) comprehensive: *EGFR, ALK, BRAF, ROS1, MET, RET, KRAS, Her2* and *NTRK* testing, ii) incomplete: <6 genes tested, and iii) no testing performed. Clinically actionable mutations will be defined as an alteration in one of the seven genes on the comprehensive gene list with an FDA approved targeted therapy in the 1L setting, plus *KRAS* G12C, *EGFR* exon 20 insertion, and *ErbB2* mutations. Molecularly informed first line therapy will be defined as one that is informed by results of NGS, obtained by plasma, tissue or both.

<u>Objective 2</u>: Evaluate contextual mechanisms contributing to the adoption, reach, and effectiveness of EHR nudge interventions with a lens for health equity.

2.2 Primary Outcomes

<u>Objective 1:</u> Availability of comprehensive molecular test results (as defined above) prior to first line therapy for patients with mNSq NSCLC.

2.3 Secondary Outcomes

<u>Objective 1:</u> 1) successful EHR-based nudge and e-CDS delivery, 2) turnaround time of delivery of provider focused alerts after receipt of plasma genotyping results, 3) completion of comprehensive molecular testing (tissue and/or plasma testing), 4) reasons for failure to complete comprehensive molecular testing (QNS or other), 5) time to molecularly-informed treatment initiation, 6) type of therapy received (targeted therapy, chemo-immunotherapy, immunotherapy, clinical trial or none) and 7) overall survival.

<u>Objective 2:</u> Individual and contextual factors shaping adoption (provider-level ordering of molecular testing), reach (patient-level completion of molecular testing), and effectiveness (receipt of molecular testing), of interventions, guided by RE-AIM with Equity Extension Framework¹².

3. STUDY POPULATION

3.1 Target Population

This stepped wedge cluster randomized trial will be conducted across newly diagnosed patients with mNSq NSCLC treated at Penn Medicine that comprise 3 clusters (sites): 1) Lancaster General Health(LGH), 2) Penn – New Jersey (Princeton Medical Center (PMPH), Penn

Medicine at Cherry Hill (PMCH), Penn Medicine at Washington Township (PMWT), and Penn Medicine Voorhees (PMV)), and 3) Penn Presbyterian Medical Center (PPMC).

3.2 Inclusion Criteria

Objective 1 and Objective 2 - Patients

- a) Patients with histological, or cytological diagnosis of mNSq NSCLC who have not yet received systemic treatment for metastatic disease.
- b) Patients must have completed at least one medical oncology visit at one of the participating sites: LGH, PMPH, PMCH, PMWT, PMV, PPMC for mNSq NSCLC.

Objective 2 – Clinicians

 All study site personnel involved in molecular testing, including but not limited to medical oncologists, advanced practice providers, registered nurses, phlebotomy and laboratory technician staff, and front desk staff

3.3 Exclusion Criteria

a) Incomplete staging information.

3.4 Vulnerable Populations

a) Children, pregnant women, fetuses, neonates, or prisoners are not included in this research study.

4. STUDY DESIGN

4.1 Preliminary Studies

We have conducted two prior studies that inform the design of this trial. In the first, we evaluated the impact of plasma-based molecular testing in addition to tissue testing on the detection of actionable mutations in patients with metastatic NSCLC. In 229 patients who underwent concurrent plasma and tissue molecular testing, tissue alone detected targetable mutations in 21% of patients, whereas addition of plasma testing increased targetable mutation detection to 36%.¹¹ Thus, plasma-based testing increased the rate of detection of therapeutically targetable alterations in metastatic NSCLC when used concurrently with tissue testing. In a second study, initiation of plasma-based testing, based on a BE informed EHR based nudge at time of new patient evaluation increased the proportion of patients undergoing concurrent tissue + plasma NGS testing to 90.3% (131/145) vs. 68.8% (262/381), p<0.00001(Aggarwal C et al, ASCO Quality Care Symposium, 2022). Thus, the proposed stepped wedge cluster randomized trial will examine effectiveness of this approach in a larger proportion of patients, reduce disparities in molecular testing, and target testing more optimally.

4.2 Overview

<u>Objective 1:</u> In a stepped wedge cluster randomized trial of patients with newly diagnosed mNSq NSCLC, test the effectiveness of a multicomponent BE informed EHR nudge intervention (as defined below) to increase timely receipt of comprehensive molecular test results before 1L therapy by integration of concurrent tissue and plasma molecular testing.

The design of this trial will include 3 clusters, representing the 6 community hospitals. There will be an initial period in which no clusters are exposed to the intervention. Subsequently, at regular intervals (the "steps") one cluster (or a group of clusters) will be randomized to cross from the control to the intervention under evaluation. This process will continue until all clusters have

crossed over to be exposed to the intervention. At the end of the study there will be a period when all clusters are exposed. Data collection will continue throughout the study, so that each cluster will contribute observations under both control and intervention observation periods.

Interventions:

An EHR-based nudge intervention that allows for default placement of a plasma based molecular genotyping order at time of the first new patient visit will be implemented (Figure 1). Additionally, results detected on the default plasma NGS order will be conveyed to providers in the form of an electronic clinical decision support (e-CDS) notification (Figure 3). These interventions will be considered the multicomponent nudge intervention and will be tailored to the organizational needs of each cluster.





- The EHR-based nudge intervention will fire at the time of the first telephone encounter with a new patient coordinator (NPC) based on a set of pre-populated molecular questions (Supplemental eFigure 1) for all patients with a new diagnosis of mNSq NSCLC.
- 2. The EHR-based nudge intervention will appear when the visit is opened within the electronic medical record by a provider and will allow default placement of a plasmabased NGS order. Thoracic oncology providers can opt out of this order if they feel it is not appropriate for the patient or because other molecular testing has already been initiated/completed. Large gene panel (>50 genes) based plasma will be used, and be based on site preference.

Standardization of the process will include availability of plasma kits at each of the sites LGH, PMPH, PMCH, PMWT, PMV, PPMC and clinical labs, and communication of the ordering process will be conveyed with the respective medical support teams (APP, RN) (**Figure 2**). Sites will be encouraged to choose one plasma-based assay to be used at their site in order to streamline order design.

Figure 2. Current and proposed future workflow for plasma-NGS ordering.



As part of the downstream EHR-based nudge intervention workflow, an electronic clinical decision support (e-CDS) system for alterations detected on plasma genotyping will be created and implemented into the EHR as a "Research (non-chargeable) Encounter" to alert the provider team caring for the patient (**Supplemental eFigure 2**). This support program will be created to notify clinicians of targetable mutations, as well as absence of mutations detected on plasma testing as a means of improving the timely delivery of molecularly informed therapy and alerting providers to available clinical trials.

Figure 3: Screening of plasma NGS reports and creation of e-CDS.



- 1. The study team will review plasma NGS reports for therapeutically targetable alterations (based on NCCN recommended biomarkers including *EGFR*, *ALK*, *ROS1*, *RET*, *MET*, *BRAF*, *KRAS*, ErbB2 and *NTRK*) (**Table 1**). When identified, the research coordinator(s) will alert the internal review team (Drs. Aggarwal and Marmarelis).
- 2. If deemed appropriate the research coordinator(s) will assemble and send an EHRbased reflex alert with the information included in **Figure 3** to the patient's oncologist and associated APP. This alert will be created by the internal review team and will include information about the possible therapeutic options for this alteration including available clinical trials.
- 3. To ensure that providers are reaching the most appropriate molecularly informed treatment decision, e-CDS alerts will be sent to providers even for mutations deemed not "therapeutically targetable" (i.e., *STK11, TP53*, etc.).
- 4. Provider response to the e-CDS program, plan to prescribe targeted therapy as well as prescription of targeted therapy in response to a molecular alteration will be monitored to

determine provider engagement. Reasons for not prescribing targeted therapy will also be recorded, if available.

Table 1.

Gene	Mutations	Targeted Therapies
EGFR	Exon 19 del/ Exon 21 L858R	Osimertinib
EGFR	Exon 18del/ins, E709A, G719A, G719C, G719R, G719S, Exon19del, <i>Exon20Ins</i> , T790M, S768I, <u>C797S</u> , L858R, L861Q	Afatinib, Erlotinib, Gefitinib, Osimertinib, Dacomitinib, Amivantamab
ALK	EML-ALK fusion , <u>F1174L</u> , <u>G1123S, G1202R</u> , <u>I1171S</u> , <u>I1171T</u> , <u>L1196Q</u>	Alectinib, Brigatinib, Ceritinib, Crizotinib, Lorlatinib
ROS1	Fusions	Brigatinib, Ceritinib, Crizotinib, Entrectinib, Lorlatinib
RET	RET-KIF5B Fusion RET fusions with CCDC6, NCOA, TRIM33, CUX1, KIAA1217, FRMD4A, KIAA1468	Pralsetinib, Selpercatinib
MET	Exon 14 skipping mutation	Crizotinib, Capmatinib, Cabozantinib
BRAF	V600E , <u>V600</u>	Dabrafenib, Dabrafenib/Trametinib, Vemurafenib
KRAS	G12C	Clinical Trials
ErbB2	Exon20Ins	Trastuzumab-deruxtecan, Ado-trastuzumab, emtansine, Afatinib, Lapatinib, Neratinib
NTRK	NTRK 1,2,3 fusions	Larotrectinib, Entrectinib

<u>Objective 2</u>: Evaluate contextual mechanisms contributing to the adoption, reach, and effectiveness of EHR nudge interventions, with a lens for health equity.

Using rigorous approaches proven successful in our prior work¹³, we will recruit patient and clinician participants from each site to complete semi-structured interviews and structured questionnaires (**Supplemental eFigure 3**). The goal of this objective is to understand contextual mechanisms (e.g., patient, clinician, clinic, structural factors) shaping adoption, reach, and effectiveness of each intervention and identify how response may differ by key patient characteristics. These data will be analyzed using convergent mixed methods analysis, which is employs the simultaneous collection and analysis of both quantitative and qualitative data to gain a comprehensive understanding of the multi-level factors shaping trial outcomes.

4.3 Study Duration and Timeline

The study duration will be approximately 34 months.

Trial Timeline												
Project Timeline		Year 1 Year 2 Year 3										
	Q1	Q2	Q3	Q4	Q1	Q2	Q3	Q4	Q1	Q2	Q3	Q4
Protocol Development, IRB	•											
Objective1: Deliver			•	•	•	•	٠	•	•	•		
intervention												
Objective 1: Endpoint			•	•	•	•	٠	•	•	•		
ascertainment												
Objective 1: Endpoint analysis											•	٠
Objective 2: Recruit & conduct				٠	•	•	٠	٠	•	•	•	
patient interviews												
Objective 2: Recruit & conduct										•	•	•
clinician interviews												
Objective 2: Mixed methods										•	•	•
coding & analysis												
Baseline Analysis and			•		•		•		•			
disseminate results												
Submit manuscripts &									•	•	•	•
disseminate overall results												

<u>Objective 1:</u> Following an observation period, in month 6, we will initiate the stepped wedge cluster randomized trial. Each cluster will include a 5-month observation period, and a 1-month washout. The active enrollment period will be 24 months, with a 6-month lookback for baseline period observation. Longitudinal data capture to collect secondary outcome information for patients will be completed up to 90 days after study completion. Thus, collection and verification of all study endpoints will be completed by the end of year 3.



		20	21			20	22			20	23			20	24			20	25		Retrospective Data	
	Q1	Q2	Q3	Q4	. ,	shout Period rvention Active																
New Jersey Cluster	-	-	-	-	-	-	-	-	-	-		-			-	-	-	-	-			dy Close
Lancaster																						
Presbyterian																						

<u>Objective 2:</u> For patient interviews, we will use rolling enrollment to capture variation in processes and effectiveness over time. Patient interview recruitment and data collection is estimated to begin during the first month of intervention observation (i.e., second month of after implementation to account for washout period) and continue until saturation if reached. Clinician recruitment and data collection will start after active trial observation is completed at the specific site. Mixed methods analysis (including transcription and coding) will run throughout year 3.

4.4 Study Setting

This study will occur within the University of Pennsylvania Health System.

5. STUDY PROCEDURES

5.1 Recruitment and Retention

<u>Objective 1:</u> A waiver of informed consent is requested for the stepped wedge cluster randomized trial. The study evaluates molecular testing rates before and after intervention at each site. Molecular testing at the initial diagnosis of mNSq NSCLC is standard, therefore, physicians and their patients will not be consented as this is the standard of practice.

<u>Objective 2:</u> This portion of the study will utilize prospective consent with waiver of written documentation. An estimated 30-40 patients and 10-20 clinicians will be interviewed (sample size dependent upon when data saturation is reached). Interview participants will also complete a structured questionnaire at the time of the interview.

Patients: A sample of patients will be invited to participate in the semi-structured interview and survey via email, phone call, and/or letter within approximately six weeks after their first medical oncology visit with a Penn Medicine provider. Participants who have completed molecular testing will be invited in purposively selected batches of approximately 5 per month (to enhance capture over time) until we reach saturation, which we estimate to be approximately 30-40 patients. We will oversample for Black patients (at least 50% of the sample at each site) to understand effectiveness by race. We will stratify by study site (5 -10 patients at each site) as well as the presence of mutations (including actionable mutations, non-clinically actionable mutations, and absence of mutations) in order to understand how molecular testing influences treatment pathways for each group.

Clinicians: Recruitment outreach of clinicians (i.e., procedures to invite for participation) will be similar to that of patients, except we will wait until end of active observation at each site to avoid potential contamination. Clinicians will be purposively sampled by adoption (e.g., low or high levels of ordering for molecular testing) and clinical role (e.g., oncologists, nurses, clinical leads, new patient coordinators) to enhance variation. Recruitment will continue until we reach

saturation, which we estimate to be approximately 2-3 clinicians per site, and approximately 10-20 clinicians total.

Data Collection: For patients, email and letter invitations will be followed by phone calls from research staff to assess interest and schedule interviews with all interested participants. Clinician interviews will also be contacted for invitation and scheduling by letter, email and/or phone. Interviews will be conducted by a trained member of the research team and overseen by Dr. Rendle (Co-I), who has extensive experience in qualitative research. Interviews will be conducted in-person, by phone or using a HIPAA-compliant video platform, depending on participant preference. Structured questionnaire data will be collected via REDCap, a HIPAA compliant survey platform, or verbally administered if the participant cannot access this platform.

5.2 Informed Consent

<u>Objective 1:</u> This study will employ a waiver of consent mechanism.

Objective 2: Potential interview participants will be initially contacted by study team members by patient portal, email, phone, and/or letter (depending on what is available for a specific participant) and given the option to decline further contact from the team. If the participant has not opted out within two weeks, they will be contacted by phone to assess interest in participating in the study. If the participant agrees to participate, they will be scheduled to have an interview via telephone or in person (based on preference and study procedures at the time). For this portion of the study, a waiver of documentation of written informed consent will be used because the risk to the individual is minimal, a signed informed consent form (ICF) could identify that an individual participated in the study and obtaining a signed paper form would significantly decrease the likelihood of proceeding towards an interview. Prior to the start of the interview, research staff will review study purpose, procedures, and the rights of the participant. They will also provide an information statement to participants via email prior to the scheduled interview. Research staff will state that participation is voluntary and ask the participant's permission to record their interview. They will describe the transcription and de-identification process, and they will ask permission to proceed with the interview. All participants will be free to withdraw participation at any time, and study enrollment will not impact employment or care at Penn Medicine. An amended patient full interview guide and guestionnaire are available in the appendices. The full interview guide for clinicians will be submitted for IRB review prior to commencement of interviews.

Measures and Outcomes

<u>Primary outcome</u>: The primary endpoint is receipt of comprehensive molecular test results prior to 1L therapy for patients with mNSq NSCLC. This outcome encompasses successful completion of concurrent tissue and plasma based molecular testing and the ability of the patient and oncology care team to have all necessary information to collaboratively arrive at the optimal treatment approach. We anticipate that approximately 80% of patients in the interventional arm will have molecular test results available prior to initiation of first line therapy. The primary outcome will be assessed by review of clinician documentation (e.g., progress notes) within the electronic medical record (EHR). Baseline data will be collected from all 3 clusters.. Molecular testing rates will be assessed, proportion of patients that undergo complete molecular genotyping prior to start of 1L therapy for mNSq NSCLC will be tabulated (Comprehensive testing will be defined as testing of all NCCN recommended biomarkers). Proportion of patients receiving targeted therapies when therapeutically targetable alterations are detected (**Table 1**) will be tabulated on a quarterly basis. <u>Secondary outcomes</u> include: 1) successful EHR based nudge delivery, 2) turnaround time of delivery of provider focused alerts after receipt of plasma genotyping results, 3) completion of comprehensive molecular testing (tissue and/or plasma testing), 4) reasons for failure to complete comprehensive molecular testing (QNS or other), 5) time to molecularly-informed treatment initiation, 6) type of therapy received (targeted therapy, chemo-immunotherapy, immunotherapy, clinical trial or none) and 7) overall survival at 1 year, and 2 years. We will also compare our primary and secondary outcomes from our enrollment sites to baseline data collected from a non-study site (i.e., PCAM), to explore contemporaneous academic benchmark for molecular testing outside of the study.

<u>Objective 2:</u> We will use structured and validated measures and develop a semi-structured interview guide. The interview guide will be developed using RE-AIM with Equity Extension Framework, a widely used implementation science framework that measures key implementation outcomes (reach, effectiveness, adoption, implementation, and maintenance) and monitors how these outcomes vary by key determinants of health (equity). We will also draw upon the Consolidated Framework for Implementation success. For patients, we will also assess factors that we hypothesize will impact completion of patient testing (e.g., patient knowledge of molecular testing, perceived importance of molecular testing, barriers to testing) and sociodemographics (e.g., self-reported race/ethnicity, health literacy, insurance, medical mistrust) using structured and validated items when available. An amended patient full interview guide and questionnaire are available in the appendices. The full interview guide for clinicians will be submitted for IRB review prior to commencement of interviews.

5.3 Sources of Materials

<u>Objective 1:</u> Electronic health record (EHR) data will be used to collect the primary endpoint as well as covariates required for statistical analysis, including any data received from FlatIron, Guardant360, and Care Everywhere related to treatment and/or testing that has been integrated as clinical data into the PennEHR.

<u>Objective 2:</u> Semi-structured interviews and structured questionnaire data will be collected and analyzed in conjunction with quantitative measures conducted.

6. STATISTICAL DESIGN AND POWER

6.1 Sample Size

<u>Objective 1:</u> We have calculated sample size based on estimates of completion of comprehensive molecular testing prior to initiation of first line therapy. Based on our prior studies, we anticipate that the baseline rate of comprehensive molecular testing prior to first line therapy is 60%. In this stepped wedge cluster randomized trial, we wish to detect an absolute increase of 20% in our primary outcome for patients in the intervention arm.

A sample of 3 clusters in a complete stepped-wedge cluster-randomized design with 4 time periods (including the baseline), 3 steps, 1 cluster(s) switching from control to treatment at each step, and an average of 120 subjects per cluster with an average of 30 subjects per cluster per time period (for a total sample size of 360 subjects) achieves 80% power to detect a difference between proportions of 0.21701. The treatment proportion is assumed to be 0.81701 under the alternative hypothesis. The control proportion is 0.6. The test statistic used is the two-sided Wald Z-Test. The ICC is 0, and the significance level of the test is 0.05.

<u>Objective 2:</u> Proposed sample size (30-40 patients and 10-20 clinicians) is based on the estimated number of interviews needed to reach data saturation (within each group, by site, and by presence of mutations) to support mixed methods evaluation. However, interviews will continue until saturation is achieved.

6.2 Analysis Plan

Objective 1 Primary Analyses Outcome Measures: Relative and absolute change in availability of molecular testing prior to IL therapy. The change will be calculated from baseline pre-intervention period to intervention periods in all the intervention arms. Comprehensive molecular testing will be defined as comprehensive if results for EGFR, *ALK, BRAF, ROS1, MET, RET,* and *NTRK* testing are available from plasma, tissue, or both.

Primary Analyses Statistical Plan

The primary outcome is binary and will be analyzed using logistic regression, fitted using Generalized Estimating Equations (GEE). The model will include a time varying covariate to represent pre-treatment, washout, and treatment, within each randomized cluster, and an ordinal categorical variable to represent time. The GEE model will adjust variances for correlation within institution (cluster). The primary hypothesis will be tested using the z-score corresponding to the difference between treatment and pretreatment proportions (after adjustment for time effects).

Objective 1 Secondary Analyses Outcome Measures:

- 1. <u>Successful EHR based nudge delivery</u>:
 - a. Amongst eligible patients (see eligibility above), calculate the proportion of patients who received any part of the multicomponent intervention (yes/no). Applicable for the patients enrolled in the time periods following randomization.
 - b. We will also explore the proportion of patients that received the full intervention (BPA + e-CDS) in contrast to a portion of the intervention to assess fidelity.
- 2. Turnaround time of delivery of provider focused alerts:
 - a. Reported as number of days, median. Applicable for the patients enrolled in the time periods following randomization.
- 3. Completion of comprehensive molecular testing:
 - a. Amongst eligible patients, relative and absolute change in completion of comprehensive molecular testing will be tabulated, regardless of timing of 1L therapy.
 - b. Relative and absolute change in completion of comprehensive testing by tissue and plasma, plasma alone, or tissue alone will be tabulated.
- 4. <u>Reasons for failure to complete comprehensive molecular testing:</u>
 - a. Summarize reasons for failure of completion of testing
 - i. Tissue related (QNS)
 - ii. Patient related factors (unable to biopsy, patient declined biopsy etc)
 - iii. Assay related factors (plasma assay does not detect mutations)
 - iv. Other
- 5. <u>Time to molecularly informed treatment initiation:</u>
 - a. Amongst eligible patients, relative and absolute change in time to start 1L therapy.

- i. Calculated as time to therapy from the date of diagnosis of Stage IV disease (date of biopsy)
- ii. Calculated as time to therapy from the date of first new patient visit with medical oncology
- 6. <u>Type of therapy received</u>:
 - a. Targeted therapy
 - b. Chemo-immunotherapy
 - c. Immunotherapy
 - d. Clinical trial or n
 - e. None
- 7. Overall Survival:
 - a. Time from initial diagnosis to date of death or last follow up.
 - b. 1 year and 2-year overall survival rates will be calculated for the intervention group and compared to baseline.

Secondary Analyses Statistical Plan

Secondary outcomes will be summarized by time and treatment condition, as proportions, means, medians, as appropriate with two-sided 95% CIs. Successful nudge delivery, completion of testing, reasons for failure to complete testing, and type of therapy received will be tabulated and summarized as proportions. Turnaround time of delivery for provider focused alerts will be treated as time to event and summarized as median time with 95% CI. Time to treatment initiation and Overall Survival will be summarized as time to event using Kaplan Meier methods, with the effect of intervention estimated as the hazard ratio (with two-sided 95% CI).

For a complete list of clinical and interventional related variables that will be collected throughout the course of this study please refer to **Supplemental eTable 1** below.

Objective 2 Analyses Plan

We will use convergent mixed-methods analysis to explore the multilevel factors shaping the effectiveness of our EHR-based nudge intervention. The constant comparative method, guided by grounded theory, will be used to inductively explore emergent themes and deductively identify a priori domains of interest within and across interviews, guided by the RE-AIM with Equity Extension Framework and the Consolidated Framework for Implementation Research (CFIR). Two trained coders will first independently read each transcript to identify themes within each domain. We then will use this list to develop a coding dictionary and apply it to subset of the data. We will measure inter-rater reliability to document and improve coding consistency. Once high reliability is achieved (e.g., kappa > 0.8), we will apply the full coding dictionary to the interview data using Atlas.ti (computer-assisted qualitative analytic software) and produce thematic reports summarizing our findings by each domain and sub-theme. Data from structured questionnaires will be analyzed descriptively. Qualitative and quantitative data will be analyzed and triangulated using a convergent mixed-methods approach (QUAL + QUAN).

7. RESOURCES NECESSARY FOR HUMAN RESEARCH PROTECTION

Adequate facilities are available within Penn Medicine's Clinical Practice Network. Members of the research team, listed in HSERA, will be overseen by the PI and include appropriate personnel to successfully implement this pilot project. All personnel will complete required training before being granted access to any identifying information. Training includes information on confidentiality through the Collaborative IRB Training Initiative (CITI) courses. All personnel

will also be trained in procedures for reporting unintentional breaches in confidentiality to the PI. All personnel will be aware that violations of participant's confidentiality, either unintentional or deliberate, may result in termination of hire.

Protection of Human Subjects

Computer-based files will only be made available to personnel involved in the study through the use of access privileges and passwords. Wherever feasible, identifiers will be removed from study-related information. Precautions are already in place to ensure the data are secure by using passwords and HIPAA-compliant encryption. Data on physicians and patients will be obtained from EHR and Penn Data Store. Any information that is obtained will be used only for research purposes and to inform the interventions described above. Information on individual patients will only be disclosed within the study team. All study staff will be reminded of the confidential nature of the data collected and contained in these databases. Data will be stored, managed, and analyzed on a secure, encrypted server behind the University of Pennsylvania Health System (UPHS) firewall. Data access will be password protected. Whenever possible, data will be de-identified for analysis.

8. STUDY TEAM

Our interdisciplinary team includes investigators with world known experts in thoracic oncology, and clinical implementation of molecular testing, implementation science, behavioral economics, EHR-based strategies, and mixed-methods research. At the University of Pennsylvania, the work will be led by Charu Aggarwal, MD, MPH, Leslye M. Heisler Associate Professor of Lung Cancer Excellence and Associate Director, Penn Center for Precision Medicine . Other key investigators include Melina E. Marmarelis, MD, a medical oncologist with clinical and research expertise in lung cancer. Other co-investigators include E. Paul Wilyeto, PhD (biostatistician), and Katharine Rendle, PhD, MSW, MPH who bring statistical and implementation science expertise to the team. Local team leaders will be Chris D. Avella, MD at Penn Presbyterian Medical Center, Ramy Sedhom, MD at Penn-Princeton, Shayma Kazmi, MD at Penn-Cherry Hill and Penn-Washington Township, and Penn Voorhees, and Samuel Kerr, MD at Penn-LGH. The study team will include Peter Gabriel, MD, Chief Oncology Informatics Officer at the Abramson Cancer Center, Meagan Hume, MDP, MPH, Senior Innovation Manager at PC3I, Anthony Martella, BA, Innovation Manager at PC3I, , , and, Clinical Research Coordinator B, Clinical Research Unit. Weilu Song, MS, MPH, Statistical Analyst, Dept. of Family Medicine and Community Health, Chelsea Saia, MPH, Senior Health Data Manager, Dept. of Family Medicine & Community Health, Jocelyn Wainwright, MS, Associate Director of Research Operations, PC3I, Alex Watts, MS, Statistical Analyst at Penn CCEB, Jillian Kalman, BA, Clinical Research Coordinator, Dept. of Family Medicine and Community Health, Anne Montgomery, PhD, MSc, Qualitative Research Investigator, Dept. of Family Medicine and Community Health, Xiaoke Wang, MD, Clinical Research Coordinator B, Willdragon Wang, Clinical Research Coordinator B, Busra Karatas, Clinical Research Unit, Penn Medicine, and Naomi Yu, Research Assistant.

9. REFERENCES

1. Aggarwal C, Rolfo CD, Oxnard GR, Gray JE, Sholl LM, Gandara DR. Strategies for the successful implementation of plasma-based NSCLC genotyping in clinical practice. Nat Rev Clin Oncol 2020.

2. Rolfo C, Mack P, Scagliotti GV, et al. Liquid Biopsy for Advanced NSCLC: A Consensus Statement From the International Association for the Study of Lung Cancer. J Thorac Oncol 2021;16(10): 1647-62.

3. Lisberg A, Cummings A, Goldman JW, et al. A Phase II Study of Pembrolizumab in EGFR-Mutant, PD-L1+, Tyrosine Kinase Inhibitor Naive Patients With Advanced NSCLC. J Thorac Oncol 2018;13(8): 1138-45.

4. Schoenfeld AJ, Arbour KC, Rizvi H, et al. Severe immune-related adverse events are common with sequential PD-(L)1 blockade and osimertinib. Ann Oncol 2019;30(5): 839-44.

5. Singal G, Miller PG, Agarwala V, et al. Association of Patient Characteristics and Tumor Genomics With Clinical Outcomes Among Patients With Non-Small Cell Lung Cancer Using a Clinicogenomic Database. JAMA 2019;321(14): 1391-99.

6. Robert NJ, Espirito JL, Chen L, et al. Biomarker testing and tissue journey among patients with metastatic non-small cell lung cancer receiving first-line therapy in The US Oncology Network. Lung Cancer 2022;166: 197-204.

7. Bruno DS, et al. Racial disparities in biomarker testing and clinical trial enrollment in nonsmall cell lung cancer (NSCLC). . Journal of Clinical Oncology 2021;39(no. 15_suppl (May 20, 2021)): 9005-05.

8. Presley CJ, Tang D, Soulos PR, et al. Association of Broad-Based Genomic Sequencing With Survival Among Patients With Advanced Non-Small Cell Lung Cancer in the Community Oncology Setting. JAMA 2018;320(5): 469-77.

9. Thompson JC, Yee SS, Troxel AB, et al. Detection of Therapeutically Targetable Driver and Resistance Mutations in Lung Cancer Patients by Next-Generation Sequencing of Cell-Free Circulating Tumor DNA. Clin Cancer Res 2016;22(23): 5772-82.

10. Leighl NB, Page RD, Raymond VM, et al. Clinical Utility of Comprehensive Cell-free DNA Analysis to Identify Genomic Biomarkers in Patients with Newly Diagnosed Metastatic Non-small Cell Lung Cancer. Clin Cancer Res 2019;25(15): 4691-700.

11. Aggarwal C, Thompson JC, Black TA, et al. Clinical Implications of Plasma-Based Genotyping With the Delivery of Personalized Therapy in Metastatic Non-Small Cell Lung Cancer. JAMA Oncol 2019;5(2): 173-80.

12. Shelton RC, Chambers DA, Glasgow RE. An Extension of RE-AIM to Enhance Sustainability: Addressing Dynamic Context and Promoting Health Equity Over Time. Front Public Health 2020;8: 134.

13. Rendle KA, Abramson CM, Garrett SB, Halley MC, Dohan D. Beyond exploratory: a tailored framework for designing and assessing qualitative health research. BMJ Open 2019;9(8): e030123.

14. Glaser BG, Strauss AL. *The Discovery of Grounded Theory: Strategies for Qualitative Research*. Aldine Pub Co; 1967.

15. Creswell JW, Plano Clark VL. *Designing and Conducting Mixed Methods Research*. 3rd ed. SAGE Publications, Inc; 2018.

16. Rihoux B, Ragin C. Configurational Comparative Methods. SAGE Publications; 2008.

17. Kane H, Lewis MA, Williams PA, Kahwati LC. Using qualitative comparative analysis to understand and quantify translation and implementation. *Transl Behav Med*. 2014;4(2):201-208. doi:10.1007/s13142-014-0251-6

18. Weiner BJ, Jacobs SR, Minasian LM, Good MJ. Organizational designs for achieving high treatment trial enrollment: a fuzzy-set analysis of the community clinical oncology program. *J Oncol Pract*. 2012;8(5):287-291. doi:10.1200/JOP.2011.000507

19. Baumgartner M, Ambühl M. Causal modeling with multi-value and fuzzy-set Coincidence Analysis. *Polit Sci Res Methods*. 2020;8(3). doi:10.1017/psrm.2018.45

20. Ragin CC. *Redesigning Social Inquiry: Fuzzy Sets and Beyond*. University of Chicago; 2008.

21. Whitaker RG, Sperber N, Baumgartner M, et al. Coincidence analysis: a new method for causal inference in implementation science. *Implement Sci*. 2020;15(1):108-108. doi:10.1186/s13012-020-01070-3

22. Thiem A. Using Qualitative Comparative Analysis for Identifying Causal Chains in Configurational Data: A Methodological Commentary on Baumgartner and Epple (2014). *Sociol Methods Res.* 2015;44(4):723-736. doi:10.1177/0049124115589032

23. Baumgartner M, Thiem A. Often Trusted but Never (Properly) Tested: Evaluating Qualitative Comparative Analysis. *Sociol Methods Res*. 2020;49(2). doi:10.1177/0049124117701487

24. Baumgartner M, Ambühl M. Cna: An R Package for Configurational Causal Inference and Modeling.

10. APPENDICES

Supplemental eTable 1: Clinical variables and tumor characteristics

Histology i.e. adenocarcinoma, poorly differentiated etc. Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any	Variable	Description
Study Patient ID Unique identifier generated by study team Name First Last MRN Medical record number DOB date of birth, MM/DD/YYYY DOD date of death, MM/DD/YYYY DOC date of death, MM/DD/YYYY Vital status Alive, deceased ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Concologist MD name (First Last) Date of 1% MedOnc encounter MM/DD/YYYY Stong status Current, former, never Molecular testing Spot, NGS, FISH, IHC, etc. Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes <	Demographics	•
MRN Medical record number DOB date of birth, MM/DD/YYYY DOD date of death, MM/DD/YYYY DOD date of death, MM/DD/YYYY Vital status Alive, deceased ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MW/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tissue Testing MM/DD/YYYY Type of Tissue Testing Spt, NGS, FISH, IHC, etc. Type of Tissue Testing Spt, NGS, FISH, IHC, etc. Gene panel size		Unique identifier generated by study team
DOB date of birth, MM/DD/YYYY DOD date of death, MM/DD/YYYY Vital status Alive, deceased ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tissue Testing MM/DD/YYYY Spot, NGS, FISH, IHC, etc. yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	Name	First Last
DOD date of death, MM/DD/YYYY Vital status Alive, deceased ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tissue Testing MM/DD/YYYY Smoking status Current, former, never Molecular testing Spot, NGS, FISH, IHC, etc. Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	MRN	Medical record number
Vital status Alive, deceased ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of 1st MedOnc encounter MM/DD/YYYY Smoking status Current, former, never Molecular testing Spot, NGS, FISH, IHC, etc. Type of Tissue Testing Spot, NGS, FISH, IHC, etc. Tissue testing successful? cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	DOB	date of birth, MM/DD/YYYY
ECOG date (@ time of 1st MedOnc visit) MM/DD/YYYY ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tist MedOnc encounter MM/DD/YYYY Smoking status Current, former, never Molecular testing Spot, NGS, FISH, IHC, etc. Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	DOD	date of death, MM/DD/YYYY
ECOG value 0-1, >2 Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of 1st MedOnc encounter MM/DD/YYYY Smoking status Current, former, never Molecular testing MM/DD/YYYY Type of Tissue Testing Spot, NGS, FISH, IHC, etc. Tissue testing successful? yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	Vital status	Alive, deceased
Days difference (ECOG date – Dx date) # (days) American Indian or Alaskan Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tissue Testing MM/DD/YYYY Spot, NGS, FISH, IHC, etc. yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) Tissue Testing Spot, NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	ECOG date (@ time of 1 st MedOnc visit)	MM/DD/YYYY
American Indian or Alaskan Native, Asian, Black or Race African American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, Unknown Ethnic group Hispanic or Latino, Non-Hispanic or Latino, Declined Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of Tissue Testing MM/DD/YYYY Smoking status Current, former, never Molecular testing Spot, NGS, FISH, IHC, etc. Yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) tissue testing successful? Tissue testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	ECOG value	0-1, >2
RaceAfrican American, Native Hawaiian or Other Pacific Islander, White, Other, Declined, UnknownEthnic groupHispanic or Latino, Non-Hispanic or Latino, DeclinedSexFemale, male, other (if available)Date of Diagnosisdate of clinical or pathologic diagnosis, MM/DD/YYYYAge at Diagnosis# (year)Histologyi.e. adenocarcinoma, poorly differentiated etc.TNM StageStage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1)Primary site of treatmentPPMC, PMC, LGH, PMCH, PMWT, OHCIMedical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Days difference (ECOG date – Dx date)	# (days)
Sex Female, male, other (if available) Date of Diagnosis date of clinical or pathologic diagnosis, MM/DD/YYYY Age at Diagnosis # (year) Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of 1st MedOnc encounter MM/DD/YYYY Smoking status Current, former, never Molecular testing MM/DD/YYYY Type of Tissue Testing Spot, NGS, FISH, IHC, etc. Tissue testing successful? yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	Race	African American, Native Hawaiian or Other Pacific
Date of Diagnosisdate of clinical or pathologic diagnosis, MM/DD/YYYYAge at Diagnosis# (year)Histologyi.e. adenocarcinoma, poorly differentiated etc.TNM StageStage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1)Primary site of treatmentPPMC, PMC, LGH, PMCH, PMWT, OHCIMedical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Ethnic group	Hispanic or Latino, Non-Hispanic or Latino, Declined
Age at Diagnosis# (year)Histologyi.e. adenocarcinoma, poorly differentiated etc.TNM StageStage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1)Primary site of treatmentPPMC, PMC, LGH, PMCH, PMWT, OHCIMedical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Sex	Female, male, other (if available)
Histology i.e. adenocarcinoma, poorly differentiated etc. TNM Stage Stage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1) Primary site of treatment PPMC, PMC, LGH, PMCH, PMWT, OHCI Medical Oncologist MD name (First Last) Date of 1st MedOnc encounter MM/DD/YYYY Smoking status Current, former, never Molecular testing MM/DD/YYYY Type of Tissue Testing Spot, NGS, FISH, IHC, etc. Tissue testing successful? yes/no (QNS – quality/quantity not sufficient, cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	Date of Diagnosis	date of clinical or pathologic diagnosis, MM/DD/YYYY
TNM StageStage IV (T4, N0, M0 or any T, N1, M0 or any T, any N, M1)Primary site of treatmentPPMC, PMC, LGH, PMCH, PMWT, OHCIMedical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Age at Diagnosis	# (year)
TNM StageN, M1)Primary site of treatmentPPMC, PMC, LGH, PMCH, PMWT, OHCIMedical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Histology	i.e. adenocarcinoma, poorly differentiated etc.
Medical OncologistMD name (First Last)Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingMM/DD/YYYYDate of Tissue TestingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	TNM Stage	
Date of 1st MedOnc encounterMM/DD/YYYYSmoking statusCurrent, former, neverMolecular testingCurrent, former, neverMolecular testingMM/DD/YYYYDate of Tissue TestingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Primary site of treatment	PPMC, PMC, LGH, PMCH, PMWT, OHCI
Smoking statusCurrent, former, neverMolecular testingMM/DD/YYYYDate of Tissue TestingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Medical Oncologist	MD name (First Last)
Molecular testingMM/DD/YYYYDate of Tissue TestingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Date of 1 st MedOnc encounter	MM/DD/YYYY
Date of Tissue TestingMM/DD/YYYYType of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Smoking status	Current, former, never
Type of Tissue TestingSpot, NGS, FISH, IHC, etc.Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Molecular testing	
Tissue testing successful?yes/no (QNS – quality/quantity not sufficient, cancelled, etc.)Tissue Testing PlatformCPD NGS/FTP, CARIS, GenPath OnkoSight, etc.Gene panel size>50 vs. <50 genes	Date of Tissue Testing	MM/DD/YYYY
Tissue testing succession? cancelled, etc.) Tissue Testing Platform CPD NGS/FTP, CARIS, GenPath OnkoSight, etc. Gene panel size >50 vs. <50 genes	Type of Tissue Testing	
Gene panel size >50 vs. <50 genes	Tissue testing successful?	
Tissue NCCN markers*EGFR, ALK, BRAF, ROS1, MET, RET, ErbB2, NTRK.Tissue additional markersKRAS, ErbB2, BRCA1 (yes/no and specific alteration)Date of Plasma TestingMM/DD/YYYYPlasma Testing PlatformGuardant360®, Foundation etc.	Tissue Testing Platform	CPD NGS/FTP, CARIS, GenPath OnkoSight, etc.
Tissue NCCN markers NTRK. Tissue additional markers KRAS, ErbB2, BRCA1 (yes/no and specific alteration) Date of Plasma Testing MM/DD/YYYY Plasma Testing Platform Guardant360®, Foundation etc.	Gene panel size	
Inssue additional markers alteration) Date of Plasma Testing MM/DD/YYYY Plasma Testing Platform Guardant360®, Foundation etc.	Tissue NCCN markers*	NTRK.
Plasma Testing Platform Guardant360®, Foundation etc.	Tissue additional markers	
	Date of Plasma Testing	MM/DD/YYYY
Type of Plasma Testing Spot, NGS, FISH, IHC, etc.	Plasma Testing Platform	Guardant360®, Foundation etc.
	Type of Plasma Testing	Spot, NGS, FISH, IHC, etc.

Plasma testing successful?	yes/no (QNS – quality/quantity not sufficient,
Plasma NCCN markers*	cancelled, etc.) EGFR, ALK, BRAF, ROS1, MET, RET, ErbB2, NTRK.
Plasma additional markers	KRAS, ErbB2, BRCA1 (yes/no and specific alteration)
Gene panel size	>50 vs. <50 genes
Testing modality	T, P, or T+P
Treatment	
First line start date	MM/DD/YYYY
First line treatment regimen	systemic therapy regimen (IO, chemo-IO, TKI, etc.)
First line last treatment date	MM/DD/YYYY
Second line treatment start date	MM/DD/YYYY
Second line treatment regimen	systemic therapy regimen (IO, chemo-IO, TKI, etc.)
Second line last treatment date	MM/DD/YYYY
Radiation prior to first line?	yes/no
Type of radiation prior to first line	curative, palliative etc.
Date of completion of radiation	MM/DD/YYYY
Assessment	
NCCN markers tested prior to 1L start?	yes/no
Targeted Tx prescribed if targetable mutation detected? (If no, will record reasons why)	yes/no
Line of targeted TX prescribed?	1L, 2L, subsequent lines
Response to Reflex Alert Notification	
Alteration detected	yes/no
Alteration gene	EGFR L858R, KRAS G12C, KIF5B-RET Fusion, etc.
Date of plasma report	MM/DD/YYYY
Date reflex alert sent	MM/DD/YYYY
Turnaround time (TAT) of alert	# (days)
Name of MD receiving alert	First Last
Tx naïve at time of alert?	yes/no
Tx pre-reflex alert	systemic therapy regimen (IO, chemo-IO, TKI, etc.)
Tx post-reflex alert	systemic therapy regimen (IO, chemo-IO, TKI, etc.)
Change in Tx pre-vs. post reflex alert?	yes/no
Plan to prescribe a targeted Tx?	yes/no
Receipt of targeted Tx?	yes/no
Clinical team review requested	yes/no

NPC Data Entry	† 4
Time taken: 11/9/2022 📩 1547 🕐 🖁 Responsible	Show All Choices
Molecular Stage IV? Yes No Unknown	*
New Diagnosis? Yes No	
Histology?	
Adenocarcinoma Squamous Other Unknown	
Molecular testing related to this diagnosis? Yes No Unknown	
Here Close X Cancel	↑ Previous ↓ Next

Supplemental eFigure 1: Pre-populated molecular questions

Supplemental eFigure 2: e-CDS alert template

Research (Non-C	hargeable)		11/8/2022 Division of Hematology/Oncology
Scholes, Dylan			
Progress Notes EMR-based Clinical Decisi	on Support (e-CDS) Notification		oordinator) • Encounter Date: 11/8/2022 • Signed
	alteration was detected on ment options now or in the f	a recent liquid biopsy test that yo outure.	ou sent. Please see below for
EGFR p.E746_A750del (Exon 19 deletion)	1 st -line	≥ 2 nd -line	
NCCN recommended	Osimertinib Afatinib Gefitinib Dacomitinib Erlotinib + ramucirumab Erlotinib + bevacizumab	 Targeted therapies available depending on molecular resistance mechanism. 	
Clinical Trials at UPenn			
//This patient is enrolled in th funded through Loxo Oncolo	(Melissa.Volpe@pennmedicine.u	penn.edu) for inquiries about clinical tria based NGS in newly diagnosed NSC olecular profiling was sent at the time available in Epic/	CLC (UPCC: XXXX)
Charu Aggarwal, MD MPH Melina Marmarelis, MD MSC Justin Bekelman, MD Meagan Hume, MDP Anthony Martella, BA Dylan Scholes, BS	Æ		

Supplemental eFigure 3: Patient Interview Semi-Structured Questions

Supplemental eFigure 3: Patient Interview Semi-Structured Questions

iNUDGE Patient Interview Guide (Version Date: 2.7.2025): Semi-Structured Questions

OPENING SCRIPT

Hello, thank you again for agreeing to this interview today and for taking time out of your day to speak with me. The objective of this study is to better understand your experience receiving cancer treatment at [SITE] and in particular your thoughts and experiences related to biomarker testing as part of your lung cancer care. Our future goal is to try to identify ways we can better support patients like you and improve how we deliver care. As a patient who has been diagnosed with cancer, we consider you an expert in this project and value your input and experiences. There are no right or wrong answers.

Participation in this study is completely voluntary and you can withdraw at any time. Please know that everything you say today is confidential. We request that you allow us to audio-record the conversation as it will ensure we capture your thoughts and views completely. The things you share will not be connected to your name, and the recording will be destroyed after it is transcribed. Any identifying information, for example your name, will be removed from the transcript.

Do you have any questions about the study or what is required to participate? (Answer all questions).

1. Are you comfortable with me recording this conversation? (Pause for confirmation).

2. Do you agree to participate in this study? (Pause for confirmation)

If you need a break or want to stop at any point, please let me know.

TURN TAPE ON NOW

State Interviewer Name, Date, Participant ID into recorder

- 1. To help me understand a bit more about your experiences, I would like to start by asking you to tell me about where you are currently in your cancer treatment journey and how the journey has been thus far?
 - a. Are they any specific things that have helped you through this process, either from your care team, your family, friends, or any other sources of support?

Thank you for sharing your experiences – I recognize that this can be hard to describe and talk about. Now, I am going to shift to more specific questions about your care experiences.

- 2. Do you recall approximately which month/year you had your first visit with your medical oncologist for your lung cancer care? It is okay if you don't remember I know this time is challenging and overwhelming for most patients.
- 3. During your first visit (or first visits), can you describe to me how challenging (or not) it was to understand your diagnosis and treatment options?
 - a. What, if anything, helped you to understand?
 - b. What, if anything, helped you to remember information that was given to you during these first appointments?
 - c. What might have helped you better understand and remember information during these first appointments?

As part of lung cancer care, some patients are recommended to do molecular testing (sometimes called biomarker testing) to identify which treatments might be best for their specific cancer. Sometimes molecular testing is done even before you know you have cancer for sure. Molecular testing often involves giving your blood or another type of sample at a lab or in the clinic when you see your care team.

- 4. How familiar are you, if at all, with molecular testing or biomarker testing related to lung cancer care?
 - a. IF FAMILIAR (AT ALL), what are some things you've heard about it? Where and when did you learn those things?
 - b. IF NOT AT ALL [GO TO NEXT QUESTION BUT PREFACE WITH I KNOW I JUST ASKED THIS GENERALLY BUT TO CONFIRM, DO YOU RECALL IF YOU...].
- 5. How, if at all, did your cancer care team talk with you about molecular testing in relation to your lung cancer care? [IF NO DISCUSSION, SKIP TO Q6]
 - a. How, if at all, did your care team *discuss* how molecular testing might change what treatment options would be best for you?
 - b. Do you recall if they *recommended* that you complete molecular testing, at any point in your care?
 - c. Do you recall if *you asked questions* related to molecular testing?
 - a. [IF YES] How comfortable did you feel asking questions about molecular testing with your care team?
 - b. [IF YES] How well did your care team answer your questions? Did you leave with any unanswered questions or concerns?

Now I want to ask you a few more questions about your experience with molecular testing.

- 6. At any point in your lung cancer care, did you complete molecular testing [IF NO RECALL, SKIP TO Q9]
 - a. [IF YES]: Can you tell me a bit more about <u>when</u> and <u>where</u> you completed molecular testing?
 - b. How, if at all, were your results discussed with you? By whom?
 - c. Do you recall if your care team informed you that you had molecular changes (potentially called mutations) that could shape your care or cancer? If you feel comfortable, would you mind sharing what they told you?
- 7. *Barriers/Facilitators*: What, if anything, made it easier for you to complete (or not complete) molecular testing? What, if anything, made it harder for you to complete (or not complete) molecular testing?
 - a. How, if it all, did insurance coverage impact your decision to be tested? Did you have any problems with coverage or reimbursement?
- 8. How, if at all, has your decision to complete testing negatively impacted you? For example, unexpected costs, time away from work, anxiety, or other experiences.

Now I'd like to ask you specifically about discussions and decisions you had about your cancer treatment options. Depending on your experience and the type of cancer, your doctor may have presented different options – or not. I understand you may not remember specific details so it's absolutely okay to say, 'I do not remember'.

- 9. First, I want to ask you a general question about your preferences for how much you like to be involved in decisions about treatment, because every person is different. In general, do you prefer to make decisions about treatment yourself, or share decision-making with your care team, or leave all decisions to your care team?
 - a. Can you tell me why? Are there situations when you prefer your clinicians to be more involved? Less involved?
 - b. What things, if anything, make it easier for you to be involved? Harder?
- 10. What, if anything, do you recall about the early conversations you had with your care team about your treatment options, shortly after you were diagnosed?
 - a. How and what treatment options were presented to you?
 - b. What information did you receive about the benefits and risks of each option?
 - c. How, if at all, did your care team incorporate your preferences during the decision-making process?
 - d. Did your care team provide a treatment recommendation? If so, how was it presented?
 - e. How, if at all, did your care team discuss the possibility of participating in clinical trials?
 - f. [IF MT RECALL] How, if at all, did your care team incorporate your molecular testing into these discussions of treatment options?
 - g. In general, how do you feel about your oncology care team's role in your decisions about treatment? Too involved, not involved enough, or just right?

Now I'd like to ask you about your experience with treatment.

- 11. If you feel comfortable, can you tell me what types of treatment, if any, you have started for your lung cancer care?
- 12. [IF MT RECALL] Have you been prescribed any medications related to the results of your molecular testing?
 - a. If yes, have you started taking your medications? How has your experience been so far?

- b. How challenging is it to take your medication as prescribed?
- c. How important is it to you to take your medication as prescribed? Why?
- 13. *Barriers/facilitators*: What things have made receiving treatment harder? What things have made receiving treatment easier?
- 14. How does it feel to be on this treatment?
 - a. Have you experienced side-effects on treatment? How have you coped with these side effects?
 - b. What concerns or worries have you had during your treatment? What are your coping strategies?
 - c. How, if at all, has this treatment changed your daily life and quality of life?

Now I'd like to ask how you could be better supported. I recognize that some things are likely outside the scope of Penn Medicine, please answer these questions as if anything is possible.

- 15. What things, if any, would help to improve your quality of life today? Support your emotional or mental well-being? Make you feel better health-wise?
- 16. What can be done to improve your experience with your cancer care team?

Wrap up

- 17. Is there anything else that you would like to tell us about your experience with molecular testing specifically or your lung care experience in general?
- 18. To help me understand a bit more about your experiences, I would like to start by asking you to tell me about where you are currently in your cancer treatment journey and how the journey has been thus far?
 - b. Are they any specific things that have helped you through this process, either from your care team, your family, friends, or any other sources of support?

Thank you for sharing your experiences – I recognize that this can be hard to describe and talk about. Now, I am going to shift to more specific questions about your care experiences.

- 19. Do you recall approximately which month/year you had your first visit with your medical oncologist for your lung cancer care? It is okay if you don't remember I know this time is challenging and overwhelming for most patients.
- 20. During your first visit (or first visits), can you describe to me how challenging (or not) it was to understand your diagnosis and treatment options?
 - d. What, if anything, helped you to understand?
 - e. What, if anything, helped you to remember information that was given to you during these first appointments?
 - f. What might have helped you better understand and remember information during these first appointments?

As part of lung cancer care, some patients are recommended to do molecular testing (sometimes called biomarker testing) to identify which treatments might be best for their specific cancer. Sometimes molecular testing is done even before you know you have cancer for sure. Molecular testing often involves giving your blood or another type of sample at a lab or in the clinic when you see your care team.

- 21. How familiar are you, if at all, with molecular testing or biomarker testing related to lung cancer care?
 - a. IF FAMILIAR (AT ALL), what are some things you've heard about it? Where and when did you learn those things?
 - b. IF NOT AT ALL [GO TO NEXT QUESTION BUT PREFACE WITH I KNOW I JUST ASKED THIS GENERALLY BUT TO CONFIRM, DO YOU RECALL IF YOU...].
- 22. How, if at all, did your cancer care team talk with you about molecular testing in relation to your lung cancer care? [IF NO DISCUSSION, SKIP TO Q6]
 - b. How, if at all, did your care team *discuss* how molecular testing might change what treatment options would be best for you?
 - c. Do you recall if they *recommended* that you complete molecular testing, at any point in your care?
 - d. Do you recall if you asked questions related to molecular testing?
 - a. [IF YES] How comfortable did you feel asking questions about molecular testing with your care team?
 - b. [IF YES] How well did your care team answer your questions? Did you leave with any unanswered questions or concerns?

Now I want to ask you a few more questions about your experience with molecular testing.

- 23. At any point in your lung cancer care, did you complete molecular testing [IF NO RECALL, SKIP TO Q9]
 - a. [IF YES]: Can you tell me a bit more about <u>when</u> and <u>where</u> you completed molecular testing?
 - b. How, if at all, were your results discussed with you? By whom?
 - c. Do you recall if your care team informed you that you had molecular changes (potentially called mutations) that could shape your care or cancer? If you feel comfortable, would you mind sharing what they told you?
- 24. *Barriers/Facilitators*: What, if anything, made it easier for you to complete (or not complete) molecular testing? What, if anything, made it harder for you to complete (or not complete) molecular testing?
 - a. How, if it all, did insurance coverage impact your decision to be tested? Did you have any problems with coverage or reimbursement?
- 25. How, if at all, has your decision to complete testing negatively impacted you? For example, unexpected costs, time away from work, anxiety, or other experiences.

Now I'd like to ask you specifically about discussions and decisions you had about your cancer treatment options. Depending on your experience and the type of cancer, your doctor

may have presented different options – or not. I understand you may not remember specific details so it's absolutely okay to say 'I do not remember'.

- 26. First, I want to ask you a general question about your preferences for how much you like to be involved in decisions about treatment, because every person is different. In general, do you prefer to make decisions about treatment yourself, or share decision-making with your care team, or leave all decisions to your care team?
 - a. Can you tell me why? Are there situations when you prefer your clinicians to be more involved? Less involved?
 - b. What things, if anything, make it easier for you to be involved? Harder?
- 27. What, if anything, do you recall about the early conversations you had with your care team about your treatment options, shortly after you were diagnosed?
 - h. How and what treatment options were presented to you?
 - i. What information did you receive about the benefits and risks of each option?
 - j. How, if at all, did your care team incorporate your preferences during the decision-making process?
 - k. Did your care team provide a treatment recommendation? If so, how was it presented?
 - I. How, if at all, did your care team discuss the possibility of participating in clinical trials?
 - m. [IF MT RECALL] How, if at all, did your care team incorporate your molecular testing into these discussions of treatment options?
 - n. In general, how do you feel about your oncology care team's role in your decisions about treatment? Too involved, not involved enough, or just right?

Now I'd like to ask you about your experience with treatment.

- 28. If you feel comfortable, can you tell me what types of treatment, if any, you have started for your lung cancer care?
- 29. [IF MT RECALL] Have you been prescribed any medications related to the results of your molecular testing?
 - a. If yes, have you started taking your medications? How has your experience been so far?
 - b. How challenging is it to take your medication as prescribed?
 - c. How important is it to you to take your medication as prescribed? Why?
- 30. *Barriers/facilitators*: What things have made receiving treatment harder? What things have made receiving treatment easier?
- 31. How does it feel to be on this treatment?
 - a. Have you experienced side-effects on treatment? How have you coped with these side effects?
 - b. What concerns or worries have you had during your treatment? What are your coping strategies?
 - c. How, if at all, has this treatment changed your daily life and quality of life?

Now I'd like to ask how you could be better supported. I recognize that some things are likely outside the scope of Penn Medicine, please answer these questions as if anything is possible.

- 32. What things, if any, would help to improve your quality of life today? Support your emotional or mental well-being? Make you feel better health-wise?
- 33. What can be done to improve your experience with your cancer care team?

Wrap up

34. Is there anything else that you would like to tell us about your experience with molecular testing specifically or your lung care experience in general?