



Fondazione IRCCS Ca' Granda
Ospedale Maggiore Policlinico

Sistema Socio Sanitario



Regione
Lombardia

Department of Services and Preventive Medicine
UOC Transfusion Center - Director: Dr. Daniele Prati
Tel. 0255036595 - email: luca.valenti@unimi.it

**“Development of a neuronal microscope for cell characterization and manipulation”
(Neuronal microscopy for cell behavior visual examination and manipulation)
Acronym: REVEAL**

Protocol version number: see 1.0 Date: 09 December 2020

Promoter: IRCCS Ca' Granda Foundation Maggiore Policlinico Hospital, Via Sforza
28, 20122 Milan, Italy

Coordinating center: UOC Transfusion Center
IRCCS Ca' Granda Foundation Ospedale Maggiore Policlinico, Via Sforza 35, 20122 Milan, Italy

Principal Investigator: Prof. Luca Vittorio Valenti

STATEMENT OF CONFIDENTIALITY

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UOCTransfusion Center – Director: Dr. Daniele Prati
Tel. 0255036595–email:luca.valenti@unimi.it

FLOWCHART

	Screening	Intervention
Period	(-t1)	(t1)
<i>Enlistment</i>		
informedconsent	-	
inclusion/exclusioncriteria	-	
<i>Administrationoftheintervention</i>		
Biologicalsamplecollection		-

LISTOFABBREVIATIONS

ALS:Automated Lab Solutions GmbH

THEREIS:EthicsCommittee

CEA:Commissariatàl'énergieatomiqueetauxénergiesalternatives

CI:InformedConsent

CRF:CaseReportForm,datacollectionform**EC:**

EuropeanCommunity

ENS:EcoleNormaleSupérieuredeLyon**GCP:**GoodClinicalPractic

e, good clinical practice **HCC:**HepatocellularCarcinoma,

hepatocellularcarcinoma**LMU:**LudwigMaximilian University

of Munich **WUT:**PolitechnikaWarszawska





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UOCTransfusion Center – Director: Dr. Daniele Prati
Tel. 0255036595–email:luca.valenti@unimi.it

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1. INTRODUCTION

1.1 Background and rationale

Hepatocellular carcinoma (HCC) is the most common type of primary liver cancer and is a leading cause of cancer-related death worldwide. The prognosis of HCC remains poor, with a 5-years survival rate of 18% (1). Risk factors for HCC include viral infection, autoimmune hepatitis, chronic alcohol use or metabolic fatty liver disease, obesity, and diabetes mellitus (2). Furthermore, alterations and chronic inflammation of the microenvironment can facilitate the transformation of normal liver stem cells into precancerous tumor stem cells (3). All these underlying pathogenic stimuli can induce a spectrum of genetic and epigenetic modifications, which are involved in the cell cycle, cell growth and regulation of adhesion. Therefore, heterogeneity and tumor priming potential arise from a combination of both endogenous and exogenous factors (4). In particular, hepatocellular carcinoma is an extraordinarily heterogeneous disease due to the morphological and histological diversity of liver cell types, and such intra-tumoral heterogeneity represents a major challenge for tumor characterization and therapeutic management of patients with HCC.

Current *in vitro* models, based on conventional hepatoma and hepatocarcinoma cell lines, fail to recapitulate key characteristics of tumor tissues such as three-dimensional tissue architecture, cellular heterogeneity, and cell-cell interactions. Organoids, which are 3D cellular structures generated from both induced pluripotent stem cells and adult tissue-resident stem cells, have recently been exploited to overcome the limitations of 2D cell culture systems, emerging as powerful tools for studying human diseases (5, 6). This is possible as they are able to stably preserve the genetic information of the autologous tissue and mimic the pathological state of the tissue itself.

These structures will be the basis for developing a new and promising methodology of microscopy, called "neuronal", which is strongly based on neural networks capable of perception, interpretation, inference, prediction, decision, and action.

2. OBJECTIVE/HYPOTHESIS OF THE EXPERIMENT

The aim of the project, within which this clinical study is part, is to validate the ability of a "neuronal" microscope to decipher the molecular mechanisms at the origin of cancer, especially by addressing the problem of biological heterogeneity. Molecular characterization of tumor cell subtypes at the single-cell level has the potential to resolve the temporal order of events governing the developmental trajectories of liver cancer. Thanks to the neuronal microscope able to select individual cells, we intend to identify and isolate cells at different points of the trajectories that lead fr



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Tel. 0255036595-email:luca.valenti@unimi.it

omthenormalstatetoth

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Via Francesco Sforza, 28 - 20122 Milano
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tumor (“past, present and future”).

More specifically, the scientific project aims to develop a “cell-harvesting neuronal microscope” that will allow in combination with biological analyses (proteomics, genetics, etc.) to characterize the cellular heterogeneity known to cause liver cancer. The challenge is to develop a new molecular imaging technique which can potentially reveal new cellular mechanisms at the origin of a disease.

To demonstrate that, this neuronal microscopy can lead to understanding the cellular origin of a disease, it will be used to decipher, identify and predict the state of cells originating from liver tumors derived from patients and mouse models of this disease.

2.1 Primary objective

The primary objective of the study conducted at the Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico is to generate and characterize three-dimensional cell culture models, called organoids, of the main liver cell populations, in order to imitate the early stages of human carcinogenesis and then analyze the onset of HCC, to identify the specific mechanisms involved in hepatocellular carcinoma.

2.2 Secondary objectives

1. Liver organoids (normal, pre-tumor or tumor) are used to “train” the neuronal microscope in order to distinguish various cell subtypes within heterogeneous cultures and recognize pre-neoplastic cells able to initiate the transition from a normal state to a tumor state.

2. On this basis, a “single cell sampling” system will be developed that will allow us to detect and collect different subtypes of liver cells that are expected to become cancerous and to analyze them before the tumor manifests itself.

3. STUDY DESIGN

3.1 Study design

Interventional, with the collection of biological, non-pharmacological, monocentric material.



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RESPONSIBILITY (role of the promoter and collaborators)

Operational unit	Participant name	Role and functions in the firm
Department of Transfusion Medicine and Hematology, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico	Dr. Daniele Prati Prof. Luca Valenti (principal investigator)	Sample recruitment and characterization; Isolation and generation of organoids, spheroids and tissues; Characterization of organoids by gene and protein expression Data analysis

Internal collaborations:

Operational unit	Participant name	Role and functions in the firm
UOC General Surgery and Liver Transplants, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico	Prof. Giorgio Rossi	Reporting of eligible patients and sample characterization
UOC Scientific Direction Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico	Dr. Stefano Gatti	Reporting of eligible patients, sample characterization and support in cell isolation protocols

External collaborations:

Collaborating body	Role and functions in the project
Commissariat à l'énergie atomique et aux énergies alternatives, France (CEA)	Development of the 2D neuronal scanning lens-free microscope and prototyping of the 3D neuronal microscope; validation of the microscope prototype through the use of human and murine samples generated respectively at the Foundation and ENS Lyon
Iprasense, France	Technical development and implementation of the lens-free neuronal microscope
Ecole Normale Supérieure de Lyon, France (ENS)	Generation of mouse models of liver disease; processing and isolation of samples generated at the Foundation; sequencing and transcriptomic analysis of single cells to generate cell trajectories



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LudwigMaximilianUniversityofMunich,Germany(LMU)	Proteomicandmulti-omicanalysesofcellandorganoidclusters
AutomatedLabSolutionsGmbH,Germany(ALS)	Developmentandvalidationofthelens- freeneuronalmicroscope;creationoftheprototypefortheis- olationofsinglecells
PolitechnikaWarszawska,Poland(WUT)	Metrological evaluation and validation of the developed 2Dand 3D microscopes, through the design and production ofcalibrationsamplesusing3Dprintingtechnology

3.2 Inclusioncriteria

Adultpatientsundergoingsurgicalcholecystectomy,liverresectionforhepatocellularcarcinomaor whole liver explants, who have given consent to participate in the study, will be included.

3.3 Exclusioncriteria

Patientswhoarepositiveforchronicviralhepatitis(HCV–RNAandHBsAg)willbeexcluded.

4. PROCEDURESRELATINGTOTHESTUDY

4.1Intervention

IncollaborationwiththeLiverTransplantUnit of Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico,wastetissuesampleswillbecollectedfrompatientsundergoingcholecystectomy,liverresecti onforhepatocellularcarcinoma(bothintra–tumoralandextra– tumoral tissues)ortakenfromexplantedwholelivers.

Thiswillallowustoisolatecellsandgenerateorganoidsstartingfromthethreeconditionsofinteresttous:

- 1) normallivertissues(cholecystectomy);
- 2) diseasedliver(extra–tumoralparenchymaandliverexplants);
- 3) hepatocellularcarcinoma.

Weaimtocollectupto10independentsamplespertissuetype.

To validate the heterogeneity obtained in vitro using organoids, mono– and multi–lineage liverspheroidswillbegeneratedbyexploitingpreviouslyestablishedhepatomacelllines(11),namely:phenoty pically differenthepatomacelllines (HepG2andSNU398cells)



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and primary cell lines representative of tumor heterogeneity (HCC-1, HCC-2, HCC-3, generated within the study "Impact of the 148M variant of the PNPLA3 gene on the regulation of retinol metabolism and activation of human hepatic stellate cells", Opinion No. 29_2016 and 29_2016bis, proceedings 300/2016), cultured in 96-well plates.

To study the role of specific genetic pathways in regulating tumor heterogeneity, we will knock down genes of interest for HepG2 hepatoma cells. To regulate the expression of specific proteins in cellular models, we will use specific antisense oligonucleotides (morpholinos: MPO, GeneTools) that block protein translation, or MP O that does not recognize any mRNA sequence as a control.

The biopsy fragments or fragments obtained from surgical resections will be transported in Celsior perfusion solution at 4°C and will be processed at the Translational Medicine laboratory in the shortest possible time to isolate liver organoids. The tissues will be mechanically fragmented into small pieces, avoiding reducing them to single cells; this will increase the formation efficiency of the organoids. Then the fragments will be further subjected to enzymatic digestion with a solution containing Collagenase and DNase at 37°C. The cell clusters obtained from enzymatic digestion will be included in reduced growth factor Matrigel. Once the Matrigel has polymerized, the complete culture medium to

grow liver organoids will be added. Isolations obtained from normal tissues will be cultured in medium containing: Advanced DMEM/F12 supplemented with 1% N2 and 1% B27 (both from GIBCO), 1.25 mM N–

Acetylcysteine (Sigma), 10 nM Leu–

Gastrin (Sigma), 50 ng/ml EGF (Peprotech), 1 µg/ml RSPO1 (Peprotech), 100 ng/ml IGF10 (Peprotech), 25 ng/ml HGF (Peprotech), 10 mM Nicotinamide (Sigma), 5 µM A83.01 (Peprotech), and 10 µM Forskolin (Peprotech). In order to increase the isolation efficiency of the organoids for the first few days, 25 ng/ml Noggin (Peprotech), 100 ng/ml Wnt3a (Peprotech) and 10 µM Rock inhibitor Y27632 (Peprotech) will be added to the medium. Isolations obtained from tumor tissues will be divided and cultured in classical medium or

in tumoroid medium containing: Advanced DMEM/F12 supplemented with 1% N2 and 1% B27 (both from GIBCO), 1.25 mM N–Acetylcysteine (Sigma), 10 nM Leu–Gastrin (Sigma), 50 ng/ml EGF (Peprotech), 100 ng/ml IGF10 (Peprotech), 25 ng/ml HGF (Peprotech), 10 mM Nicotinamide (Sigma), 5 µM A83.01 (Peprotech), 10 µM Forskolin (Peprotech), and 3 nM dexamethasone (Sigma) (8).

The samples isolated at the Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico will be sent to the ENS in Lyon, that in collaboration with CEA will develop the transcriptomic analysis on both whole human organoids and isolated cells, using the neuronal microscope.

CEA, through the use of human and murine cell samples isolated at the Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico and ENS Lyon respectively, will develop the 2D and 3D imaging platforms. LMU in collaboration with CEA will develop proteomic analysis both on whole human organoids and on isolated cells using the neuronal microscope.





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5. ENDPOINTS

5.1 PrimaryEndpoint

Isolation of epithelial cells in order to generate models of major hepatocellular carcinoma cellpopulations in a three-dimensional culture, called liver organoids, and their molecularcharacterizationofthegeneratedmodels.

5.2SecondaryEndpoints

- Knowledgeofthebehavioralandmorphologicaldifferencesbetweenphysiologicalandtumorphepatocytes;
- Knowledgeofthemolecularmechanismsunderlyingcellarlevelobservationsexploiting“omics”technologiesandCRISPR–Cas9geneticengineering.

6. DURATION / TIMELINE OF THE STUDY

Month and year of study start: 01/2021

Monthand year of enrollment closure: 01/2024

Month and yearofstudyend:01/2025

The study will have a duration of 48 months. In the first three years we expect to enroll approximately 30 subjects inaccordancewiththeinclusioncriteria.

PROJECTGANTT

Intervention	Period(months)			
	1–12	13–24	25–36	37–48
Enlistment	-	-	--	
Informedconsent	-	-	-	
Inclusion/exclusion criteria	-	-	-	
Generationoforganoids	-	-	-	





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Evaluationofgeneexpressioninliveror ganoids	--	--	--	
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Evaluation of specific liver proteins in liver organoids	--	--	--	
Data correlation analysis		-	-	-

7. STATISTICAL ANALYSIS

7.1 Sample size and data analysis

Previously published works (7,8,9,10) have highlighted that the generation efficiency of liver

organoids is approximately 90% starting from surgical resections and 33% from biopsies (7,8). Considering these estimates, in order to isolate at least 6 independent organoid cultures of each type of condition of interest in our study, it is estimated to recruit at least 30 donors (10 for each condition studied). The intent is to obtain replicates for analyses with a sufficient representation of the different stages of liver damage (normal liver, inflammation/fibrosis and hepatocarcinoma) and to be able to map the impact of the most common hereditary genetic variants (present in at least 20% of individuals) in the population.

The main analyses will be conducted from cell cultures isolated from a single individual. Given the descriptive nature of mapping cellular phenotypes and gene expression, and the large number of organoids ($> 10^3$) and cells ($> 10^9$) from a single individual, we will have sufficient statistical power ($> 80\%$) to identify and analyze specific cellular subsets representing $> 1\%$ of the population overall.

8. ADVERSE EVENTS

The project does not involve the administration of drugs or other substances or ad hoc invasive intervention outside of normal clinical practices. Therefore, no adverse events are expected.

9. RISK/BENEFIT ASSESSMENT

The study does not predict an immediate benefit for the donors, but the results of this trial will have the potential to lead to a decoding of the mechanisms at the origin of tumor development, resolve the temporal order of the events that regulate the evolutionary trajectory, understand the progression, characterization and improve the therapeutic management of patients.



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10. PRACTICE MANAGEMENT

10.1 Data collection and management

For each participant will be assigned a unique code upon enrollment. The file associating the participant's code with the relevant identification data will be stored separately on a computer protected with a password. The study database will be protected with a different password and accessible only to staff designated by the principal investigator. The unique code will be used to prevent that unauthorized individual will be able to discover the identity of the enrolled patient. Only designed investigators will be able to know the identity of the enrolled subjects.

Regarding data storage: data that can be made publicly accessible, to all partners will be required to use an open access repository, linked to the tools proposed by the European Community (e.g. OpenAIRE) to ensure access to publications and a bibliographic metadata archive in a standard format, including the information required by the EC.

10.2 Regulatory aspects and ethical considerations

10.2.1 Approval of the Competent Authority

In accordance with current regulations, the principal investigator must obtain approval from the appropriate Competent Authority before starting the clinical study.

This study will be conducted in accordance with the rules of the ICH/GCP (International Conference of Harmonization/Good Clinical Practice) and all applicable laws, including the Declaration of Helsinki of June 1964, as amended by the latest World Medical Association General Assembly in Seoul, 2008.

10.2.2 Approval of the Ethics Committee

The investigator must ensure that the protocol has been seen and approved by the local independent Ethics Committee (EC) before starting the study.

The EC

must also review and approve the informed consent (IC) form and all written information received from the patient prior to enrollment in the study.

If it is necessary to modify the protocol and/or the IC during the study, the investigator will be the guarantor and therefore the person in charge of ensuring the review and approval of this modified document according to the EC's request.

The content of these changes will be implemented only after the EC has approved them. Until that time, it will be necessary to refer to the previous version of the already approved document.





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10.2.3 Informed Consent (IC)

The investigator or other staff designed by the investigator should inform people about all aspects and procedures of the study. The process for obtaining informed consent must comply with current regulatory procedures. The investigator (or designated collaborator) and the subject must date and sign the informed consent form before that the patient initiates any study-related procedures. The subject will receive a copy of the CI dated and signed by both parties; the original copy will be kept in the archives designated for study. Neither the investigator nor designated personnel should in any way exert any coercion or influence on a subject to participate or continue to participate in the study. A subject's decision to participate in the study must be completely voluntary. The investigator and designated personnel should emphasize to the subject that they may withdraw consent at any time without penalty or loss of any benefit to which they may be entitled.

Written or oral information relating to the study, including the written consent form, must not contain any linguistic expression that forces the subject to (even apparently) waive his or her legal rights, or that would exonerate the investigator, the institution or the sponsor from liability for negligence.

10.3 Duties of the experimenter

In accordance with applicable local regulations, the investigator must send periodic reports regarding the progress of the study in his center to the EC and notify the EC of the closure of the study. Periodic reporting and closure notification are part of the investigator's responsibilities.

10.4 Study monitoring

In accordance with applicable regulations and good clinical practice (GCP), the clinical monitor must visit or contact the center periodically. The duration, nature and frequency of such visits/contacts depend on the frequency of recruitment, the quality of the documents held by the center and their adherence to the protocol.

Through these contacts, the monitor must:

- monitor and evaluate the progress of the study
- examine the data collected
- conduct verification of the source document
- identify each problem and its solutions





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The purposes of the monitoring activity are to verify that:

- the rights and welfare of the subject are respected
- The study data are accurate, complete, and can be verified from the original documents
- the study is conducted in accordance with the protocol and any approved amendments, GCP and applicable regulations

The investigator must:

- give to the clinical monitor direct access to all relevant documentation
- dedicate part of his and his staff's time to the clinical monitor to discuss the monitoring results and any other possible aspects.

The clinical

monitor should also contact the center prior to the start of the study to discuss the protocol and data collection procedures with the staff.

10.5 Quality assurance of the study

As Promoter, Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico can carry out a quality control check on the study at its discretion. In this case, the investigator must allow the clinical monitor direct access to all relevant documentation and dedicate some of his or her time and staff to the review and to discuss the monitoring results and any other aspects of the study.

Furthermore, Regulatory Authorities can carry out inspections. In this case, the investigator must grant the inspector direct access to all relevant documentation, and dedicate part of his time and staff to the inspector to discuss the monitoring results and any other aspects of the study.

10.6 Closure of the study

At the time of study closure, the monitor and the investigator must activate a series of procedures:

- review all study documentation
- reconcile study data
- reconcile all clarification reports.





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10.7 Archiving of documents

In accordance with current national regulations, the investigator must keep a copy of all documentation and store it in a dry and safe place after the study is closed.

10.8 Disclosure of information regarding scientific discovery

10.8.1 Confidentiality

The investigator and other personnel involved in the study must process all information relating to the study (including the protocol, data obtained and all documentation produced during the study) and must not use such information, data or reports for purposes other than those described in the protocol.

These restrictions do not apply to:

- 1) information that becomes publicly available, not due to negligence on the part of the investigator or his staff;
- 2) information requiring confidential disclosure to CE for the sole purpose of evaluating the study;
- 3) information that must be disclosed in order to obtain appropriate medical care for a study subject.

10.8.2 Publications

The IRCCS Ca' Granda Foundation, Ospedale Maggiore Policlinico is the sole owner of the data. A website dedicated to the project will be built with private sections that will serve the project partners as update and exchange channels, as well as to build a data archive to be used in the post-project exploitation phase. The public section will contain the project description (e.g. objectives, partners, funding source, etc.), published articles and ongoing work. The scientific director of the study will write a final report and make the results public at the end of the study. The data will be made public anonymously and presented as required in aggregate mode. In accordance with the provisions of the Grant Agreement, the dissemination of the results will take place as soon as possible, by appropriate means, including scientific publications.

10.9 Intellectual property rights on the study results

Most data will be associated with results that may have commercial or industrial protection potential and therefore cannot be made accessible for verification and re-use generally due to intellectual property protection measures.

However, relevant data necessary for verifying results published in scientific journals can be made accessible on a case-by-case basis.





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A Consortium Agreement will be negotiated and signed by all parties in order to specify, among other things, the terms and conditions relating to ownership, access rights, exploitation of the background and results as well as the dissemination of the results, in accordance with the grant agreement and Regulation n° 1290/2013 of 11 December 2013. The consortium agreement will be based on the DESCA Horizon 2020 Model Consortium Agreement with the necessary adaptations considering the specific context and parties involved in the project and

will be based on the following principles:

- The parties will identify the knowledge, information, data (of any form or nature), know-how, technical material and/or assets protected by industrial and/or intellectual property rights or susceptible to protection, (Background) held and/or developed in any capacity independently by each of the Parties at a time prior to the start of the Project which they will make available for the implementation of the project and will evaluate their availability for access rights with regard to the rights of potential third parties on such background to the extent that this information is known to the respective party at the time of the assessment;
- Ownership of results, including joint results generated by two or more parties, will go to the party or parties that generated such results;
- The proprietary parties will take all appropriate measures to protect the results susceptible to commercial or industrial exploitation, in particular through intellectual property rights, where appropriate;
- The parties will use a reasonable effort to exploit and disseminate the results, directly or indirectly, for example by licensing such results;
- Each party will grant the other parties access rights (through licenses) to its Background and the results generated during the Project, only if strictly necessary for the implementation of the project itself and/or for the exploitation of the results of such other parties (on fair and reasonable terms).

11. INDEMNITY AND COMPENSATION IN CASE OF DAMAGES

In the event of unwanted events or any damage that may arise from participation in research, our Institute's Insurance Policy is also extended to cover individual participating in research projects.

12. FINANCIAL AGREEMENTS

The costs of study procedures exceeding normal clinical practice will be entirely covered by the funds deriving from the Horizon 2020–Europe EU financing.





Department of Services and Preventive Medicine
UOCTransfusion Center – Director: Dr. Daniele Prati
Tel. 0255036595–email:luca.valenti@unimi.it

13. DISCLOSURESONCONFLICTSOFINTEREST

Theinvestigatorsdeclarenonoconflictsofinterest.

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